The effects of herbal cream and silymarin on liver in carbon tetrachloride-treated animals

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ABSTRACT

Many medical or pharmaceutical plants have been widely used for the treatment of the liver injury. Silymarin is now used as a food-supporting material for the liver as a patented product. Herbal cream has antiinflammatory and antioxidant effects on wound healing in a hemorrhoid model. This study aimed to evaluate the effects of herbal cream and silvmarin on the liver in carbon tetrachloride (CCl₄)-treated animals. Male Wistar rats were divided into seven groups as Intact control, Control, Herbal cream (0.5 ml, intrarectal), Silymarin (70 mg/kg, intrarectal), CCl₄ (2 ml/kg, intraperitoneal), CCl₄+Herbal cream (0.5 ml herbal cream for 21 days, 3 weeks after administration of CCl₄) and CCl₄+Silymarin (70 mg/kg silymarin for 21 days, 3 weeks after administration of CCl₄). Herbal cream reduced damage and leukocyte distribution induced by CCl4 and increased catalase. There was no significant change in superoxide dismutase (SOD) and glutathione peroxidase (GPx). The levels of SOD, catalase, and GPx in the liver increased significantly in the group treated only with herbal cream. These results point out that herbal cream may have antioxidant properties in the liver and a role in preventing liver damage. As a result, it has been detected that herbal cream is not a toxic agent and recovers liver damage with antioxidant properties.

Introduction

Liver diseases are a widespread public health problem all over the world. The use of herbal products is increasing worldwide. Many plants are used traditionally and commercially for treatment and support. *Silybum marianum* (Camel thistle) extract is used as a cytoprotectant in liver injury induced by carbon tetrachloride (CCl₄), alcohol, radiation, acetaminophen, iron overload, phenyl hydrazine, and *Amanita phalloides* (1, 27, 51, 61). *Silybum marianum* consists of flavanone derivatives such as silybin, silydianin, isosilybin, and silychristin (1, 58). *Silybum marianum* was suggested to be potentially beneficial for liver diseases because of its anti-inflammatory, antifibrotic, antioxidant, and hepatoprotective properties (2). In addition, silymarin exerts a significant anti-inflammatory effect by diminishing edema and inhibiting leukocyte migration to the inflammation region (14).

Artichoke (*Cynara scolymus* L.) leaf extracts were shown to have antimicrobial, antioxidative (64), hepatoprotective, and anti-cholestatic effects (22, 30). It was suggested to have antioxidative potential in hydroperoxide-induced oxidative stress (22). It was indicated to improve cadmium-induced hepatorenal oxidative injury in rats (19). The benefits of walnuts (*Juglans regia*) have been known for many years. The cosmetic and pharmaceutical industries use green walnuts, leaves, shells, and seeds (56). Walnuts may be a protective agent against the toxicity of chemicals and drugs (25). It has been proposed that the bioactive compounds of walnut green husks can trigger the death of prostate carcinoma cells through apoptotic pathways (7). It has been indicated that the extract of green walnut husks had potential antioxidant activity (43).

Ficus carica is used in traditional medicine to treat cardiovascular and respiratory disorders. It has been investigated whether fig (*Ficus carica*) leaves have anti-inflammatory and antioxidant properties (23). It has been reported that the extract of *Ficus carica* has the capacity of antioxidant and anti-inflammatory properties (4). The administration of *Ficus carica* leaf extracts has been shown to affect the oxidative stress in diabetes (48). It was observed that its leaves have hypoglycaemic activity in diabetic animals (49).

Aesculus hippocastanum (Hippocastanaceae) is known as horse chestnut tree. The therapeutic benefit of horse chestnut extract is supported by *in vivo* and *in vitro* experimental researches, and it has anti-inflammatory and antioedematous properties. Escin is the major active form of Aesculus hippocastanum and has pharmacodynamic actions (31, 34, 50, 55). Aescin have protective effects associated with anti-inflammatory effects on endotoxininduced liver injury (28). It has also been demonstrated that Aesculus hippocastanum seeds have beneficial effects on diabetic nephropathy (20). A gel formulation containing Aesculus hippocastanum was shown to have stability, and skin permeability ability (44).

The herbal cream used in this study contains the most active ingredients including *Cynara scolymus*, *Aesculus hippocastanum*, *Juglans regia*, and *Ficus carica* leaves. The herbal cream has been detected to be effective in wound healing and have antioxidant properties (24). The herbal cream was reported to have antioxidant, antiinflammatory, and wound-healing effects in our experimental studies for patent research. Moreover, a patents (European Patent No: 2022504) have been taken from the European Patent Office at the end of this study. Therefore, it is considered that different studies should be performed with this cream. For this purpose, it was decided to investigate whether this cream has a role in liver toxicity.

The aim of this study was to investigate whether herbal cream has an ameliorating effect against CCl₄induced liver injury.

Materials and Methods

Animals: All experiments in this study were approved and reviewed by the Istanbul University Local Committee on

Animal Research Ethics (Decision no: 2013/92). Care and handling of the animals were in accordance with the Institute for Laboratory Animal Research Guide for the Care and Use of Laboratory Animals. Experiments were performed with 3 months old male Wistar albino rats (250-300 g). Animals were obtained from Istanbul University, Aziz Sancar Experimental Medicine Research Institute. Animals were fed a pelleted diet and tap water as free access. Animals were divided into intact control, control, silymarin (70 mg/kg, intrarectal) (n=7), herbal cream (0.5 ml, intrarectal) (n=7), CCl₄ (2 ml/kg, intraperitoneal) (n=7), CCl₄+herbal cream (0.5 ml herbal cream for 21 days 3 weeks after administration of CCl₄) (n=7) and CCl₄+Silymarin (70 mg/kg silymarin for 21 days 3 weeks after administration of CCl₄) (n=7).

CCl₄ (2 ml/kg of the 30% CCl₄ solution) was dissolved in olive oil and administrated intraperitoneally twice weekly (Monday-Tuesday) for 3 weeks. Silymarin (Solgar®) was dissolved in isotonic saline solution. The herbal cream and silymarin were administrated intrarectally twice daily (09.00 and 17.00) for 21 days. The herbal cream or silymarin was administrated intrarectally by a syringe without a needle. The tip of the insulin syringe without a needle was inserted into the rectum reaching approximately 1 cm proximal to the anus. The herbal cream in the syringe was slowly injected into the rectum, and then the tip of the syringe was kept in this position for 1 minute to remain substances into the rectum. All herbs and plant oils used in this study were obtained from certified herb sellers. The herbal cream is comprised of an aqueous-based liquid containing herbal extracts, vegetable oils, and gelling agents according to the invention application (24). A more detailed description of the herbal cream is presented in the Supplementary Material. The control group was received twice daily (09.00 and 17.00) intrarectal injection of isotonic saline solution for 21 days after olive oil was given intraperitoneally twice weekly for 3 weeks. The intact group did not receive any substance administration. Animals were anesthetized at the end of the experiment. Blood and liver were taken for analysis.

Biochemical Analyses: Blood samples were centrifuged at 4000 x g for 10 minutes to obtain serum. Aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatases (ALP), lactate dehydrogenase (LHD), albumin, total bilirubin, and total cholesterol were measured in the serum sample by enzymatic methods on AutoAnalyzer (Roche Hitachi Cobas c311).

The liver samples were homogenized and centrifuged at 6000 x g for 10 minutes, and then supernatants were collected from homogenates. Malondialdehyde (MDA) levels (nmol/mg protein) were

measured according to the method of Ohkawa et al. (42). Total sialic acid (TSA) levels (nmol/mg protein) were determined according to the method of Katopodis et al. (29). SOD activity (U/mg protein) was calculated according to the method of Winterbourn et al. (59). SOD was defined as the amount of enzyme that causes the inhibition of nitroblue tetrazolium reduction by 50% and the enzyme activity. Catalase (CAT) activity (U/mg protein) was determined by the method of Beutler (10). The decrease in optical density per minute and enzyme activity were determined. GPx activity (U/mg protein) was measured using the methods of Paglia and Valentine (45). The protein levels of homogenates were detected using the Bradford assay (11).

Histological Analysis: The liver samples were fixed in 10% neutral buffer formalin solution and embedded in paraffin. Liver sections were stained with Hematoxylin-Eosin (HE). Liver damage was scored and evaluated in the section. The degree of damage was defined as follows: 0-normal (0-5%); 1-Mild (5-25%); 2-Moderate (25-75%); 3-Severe (>75%).

Immunohistochemical Analysis: Sections were prepared to evaluate the myeloperoxidase (MPO)-stained leukocytes as described previously (15, 32). The distribution of MPO-stained leukocytes was evaluated and scored in selected 210 areas of liver sections. The photographs of the sections were taken with Image Pro-Plus.

Statistical Analysis: The results are expressed as the mean \pm SEM. The Shapiro-Wilk test was used to test the normality of the data. Significant differences were evaluated using one-way ANOVA and Kruskal Wallis test with Bonferroni and Dunn post-tests using GraphPad Prism software (San Diego, CA, USA). A value of P<0.05 was considered significant.

Results

Biochemical Results: The albumin levels enhanced in the CCl_4+S (P<0.01) and CCl_4+HC (P<0.05) groups compared to the CCl_4 groups. Total bilirubin level was enhanced in the CCl_4 group compared to the HC group (P<0.01). Cholesterol in serum was not different between groups (Figure 1). Unlike albumin, bilirubin levels that increased with CCl_4 application decreased especially after cream application and became closer to the C group (Figure 1).

In the CCl₄ group, AST levels were similar to the C groups while ALT levels increased (P<0.01) (Figure 2). ALT and AST levels in the groups applied with herbal cream application resembled the control group.

ALP levels in the S group were increased compared to the control group (P<0.01). In addition, ALP levels in the CCl₄+S (P<0.05) and S groups (P<0.05) were higher according to the CCl₄ group. LDH level decreased in the S group (P<0.05) compared to the control group (Figure 2).

MDA levels increased in the CCl₄+S group compared with IC and the HC groups (P<0.01). TSA increased in the S group (P<0.01) compared to the IC group. Moreover, TSA increased in the CCl₄+S group compared to the control groups (P<0.05) (Figure 3).

SOD level increased in the HC (P<0.001, P<0.05, P<0.05, P<0.001, and P<0.05, respectively) or S groups (P<0.001, P<0.01, P<0.05, P<0.001, and P<0.05, P<0.001, and P<0.05, respectively) compared with IC, C, CCl₄, CCl₄+S, and CCl₄+HC groups. SOD levels in the CCl₄ group were similar to the control groups (Figure 4).

CAT levels increased in the HC groups compared with IC, C, CCl₄, and CCl₄+S groups (P<0.001, P<0.05, P<0.001, and P<0.001, respectively). CAT levels increased in the CCl₄+HC and S groups compared to the CCl₄ group (P<0.01 and P<0.001, respectively).

The levels of GPx increased in the HC group compared with IC, C, CCl₄, and CCl₄+HC groups (P<0.05, P<0.001, P<0.001, and P<0.05, respectively). GPx levels increased in the S group compared with the C and CCl₄ groups (P<0.01 and P<0.001, respectively) (Figure 4).

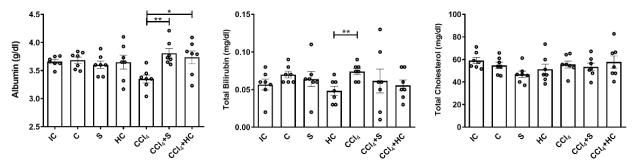


Figure 1. Albumin, total bilirubin and total cholesterol levels in the groups. *P<0.05, **P<0.01.

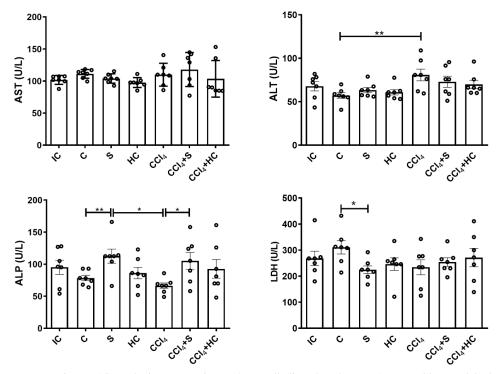


Figure 2. Aspartate transaminase (AST), alanine transaminase (ALT), alkaline phosphatase (ALP), and lactate dehydrogenase (LDH) levels in the groups. *P<0.05, **P<0.01.

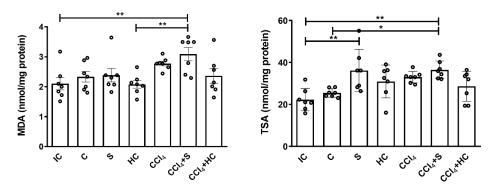


Figure 3. The malondialdehyde (MDA) and total sialic acid (TSA) levels in the groups. *P<0.05, **P<0.01.

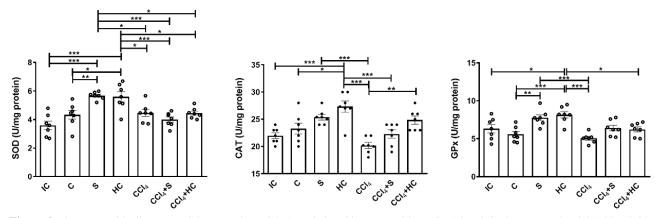


Figure 4. The superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx) levels in the groups. *P<0.05, **P<0.01, ***P<0.001.

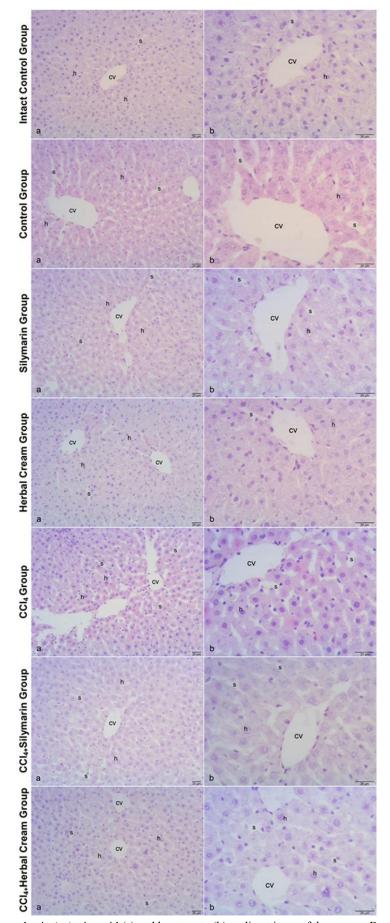


Figure 5. Central vein (cv), sinusoid (s) and hepatocyte (h) on liver tissue of the groups. Bar: 20 µm, HE.

Histological Results: The liver sections were examined under light microscopy. Histopathological changes were observed in HE-stained liver sections of CCl₄-treated animals. CCl₄ caused degeneration and expansion of sinusoids, damages in the endothelial layer, activation of Kupffer cells, and portal inflammation in liver tissue. The herbal cream or silymarin administration decreased CCl₄induced liver damage. The liver sections in the HC or S groups were similar to the control group (Figure 5, Table 1). *Immunohistochemical Results:* Leukocyte infiltration (MPO-stained leukocytes) in liver tissue increased in the CCl₄ groups compared to the IC and C groups (P<0.001 and P<0.01, respectively). The herbal cream or silymarin administration decreased leukocyte infiltration in the CCl₄ group (P<0.001) (Figure 6). Leukocytes were detected in portal areas, sinusoids, and vessels of liver tissue (Figure 7).

Parameters	Intact Control	Control	Silymarin	Herbal Cream	CCl4	CCl₄+ Silymarin	CCl₄+ Herbal cream
Degeneration and expansion of sinusoids	0.675	0.633	0.92	0.78	1.08	0.816	0.933
Damages in endothelial layer	0.275	0.233	0.26	0.22	0.9	0.3	0.254
Vacuolation of hepatocytes	0	0.366	0.02	0	0.2	0.016	0.133
The activation of Kupffer cells	0.675	0.366	0.38	0.44	0.86	0.65	0.683
Portal inflammation	0.475	0.666	0.36	0.62	1.18	0.516	0.181
Total score	2.1	2.25	1.94	2.06	4.22	2.298	2.184
Mean±SEM	0.42±0.13	0.45±0.08	0.39±0.15	0.41±0.14	0.84±0.17	0.46±0.14	0.44±0.16

Table 1. The histological scores of groups to evaluate the damage in the liver.

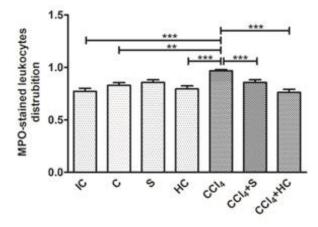
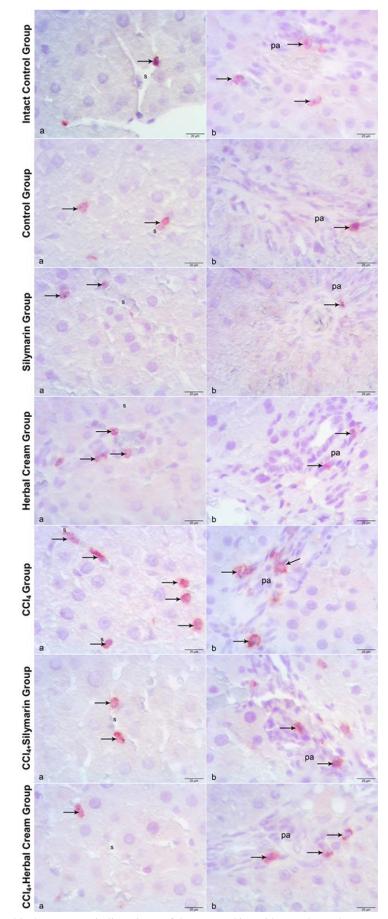
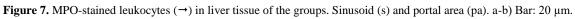


Figure 6. MPO-stained leukocyte distributions in liver tissues of groups. **P<0.01, ***P<0.001.





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Discussion and Conclusions

Liver diseases are among the most serious health problems worldwide. Liver disease is a term that indicates damage in cells, tissues, and structure, or liver dysfunction. Liver injury can occur by a variety of factors including viruses, bacteria, parasites, drugs, chemical compounds (CCl₄, dimethyl nitrosamine, thioacetamide, Dgalactosamine/lipopolysaccharide), and excessive alcohol consumption (33). Inflammation and oxidative stress participate in the pathogenesis and progression of liver diseases. For this reason, many treatment procedures including various plants such as silymarin (58), Cynara scolymus L. (22), and Physalis peruviana L. (5) are developed to prevent liver disease (57). Many plants and their extracts are used to prevent liver damage (33).

In our study, it was evaluated the potential effects of herbal cream on liver toxicity. It was previously determined that intrarectal administration of this cream was effective in wound healing and reduced inflammation and oxidative stress in rats (24). It suggests that the intrarectal application of the cream can be effective by penetrating the blood more quickly. Therefore, the protective properties of herbal cream may play an effective role in different pathological conditions in addition to hemorrhoids. As a result, it was investigated whether the herbal hemorrhoid cream has beneficial effects on the liver in the CCl₄-induced hepatotoxicity model in this study. Liver toxicity was also created with CCl₄, which is widely used in the literature (6, 35, 39). CCl₄ is used as a liver injury model to investigate the mechanism of hepatotoxic effects in animals (39). CCl₄ increases free radical formation (12). The free radicals induce lipid peroxidation, tissue damage, necrosis and releasing of AST, ALT, and ALP enzymes (16, 63). In addition, toxic agents induce an inflammatory response and activate especially Kupffer cells (47). CCl₄ increased ALT and AST levels and decreased GPx, CAT, and SOD levels. Moreover, CCl₄ caused fibrosis and necrosis in the liver (60). CCl₄ increased serum ALT and induced hepatic necrosis and inflammation (62). CCl₄ increased MDA levels (8). The administration of CCl₄ increased total bilirubin, amino transaminases, ALP, glutathione, and protein levels, while decreasing the TBARS and total protein level (26, 46). In this study, CCl₄ resulted in an increase in ALT in serum. Albumin, cholesterol, and bilirubin levels did not change in the CCl₄ group. It was observed that CCl₄ administration caused histopathological changes including degeneration and enlargement of sinusoids, endothelial damage and inflammatory cell infiltration in portal areas. The leukocyte infiltration in liver tissue indicated that CCl₄ induced the inflammatory response depending on its toxicity.

The effects of these herbal products and silymarin have been investigated in CCl₄-induced oxidative stress.

Milk thistle protected the liver against oxidative damage and decreased MDA in the liver in CCl4-administrated rats (8). The protective effect of Silybum marianum extracts was determined in CCl₄-treated animals (52). The nanoemulsion formulation of silymarin decreased alkaline phosphatase, total bilirubin, tissue lipid peroxides, pyruvate transaminase, and glutamate oxaloacetate transaminase, and increased tissue glutathione, total protein, albumin, and globulin in CCl₄-treated rats (46). Silybum marianum total extract, silymarin, and silibinin administrations improved the liver damage induced by diethylnitrosamine/2-acetylaminofluorene/CCl4, decreased ALT, AST, total bilirubin, ALP levels, and increased the serum total protein levels (61). Silymarin reduced the migrating neutrophils and caused a dose-dependent inhibition of leukocyte accumulation in carrageenaninduced inflammation (14). Beta-aescin was suggested to have antioxidative and antifibrotic properties on CCl₄induced liver injury (54). Extract of artichoke was detected to prevent hepatotoxicity on CCl₄-caused liver injury (36). Chicory and/or artichoke leaf extracts were reported to have a protective effect on CCl4 and gammairradiation-induced chronic nephrotoxicity in rats (17). Cynara scolymus leaf extract exhibited therapeutic effects on liver injury (13, 21). A chemically-characterized extract from artichoke (Cynara scolymus L.) showed hepatoprotective effects against aflatoxin B1-induced toxicity in rats (40). The walnut extract exhibited protective and antioxidant effects in CCl₄-induced oxidative liver damage (18). Aydın et al. (9) have reported that Juglans regia L. (walnut) extract has protective effects against lipid peroxidation formation in the brain, kidney, and liver tissues in CCl₄-applied Wistar rats. The leaf extract of Ficus carica had a preservative effect against CCl₄-induced hepatic damage (3, 37, 38, 41, 53). In our study, herbal cream increased CAT levels and reduced CCl₄-induced liver injury and leukocytes infiltration. The antioxidant enzymes in the liver increased significantly in the HC group. These results indicate that herbal cream may have antioxidant properties on the liver tissue and a role in preventing liver damage and also does not have hepatoxicity effects.

In conclusion, it has been determined that herbal cream contains the most active ingredients including *Cynara scolymus*, *Aesculus hippocastanum*, *Juglans regia* and *Ficus carica* leaves have antioxidant and antiinflammatory effects, and intrarectal application of natural and herbal compounds also has an important effect on pathophysiological processes such as liver damage. The researches that investigate whether there are other potential effects of the combination of fig leaves, artichoke leaves, walnut husks, and horse chestnut fruit and intrarectal therapies on different diseases may contribute to improving the life quality of patients. The authors would like to thank Feyyaz Küçükay for the drug preparation.

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Ethical Statement

This study was approved by the Istanbul University Local Committee on Animal Research Ethics (Decision no: 2013/92).

Conflict of Interest

The authors declared that there is no conflict of interest.

Author Contributions

AK, AK, EGG, IS, SU, MC, and CDT conceived and planned the experiments. AK, AK, EGG, IS, SU, and CDT carried out the experiments. AK, AK, EGG, IS, SU, SE, and CDT contributed to sample preparation. AK, AK, EGG, IS, SU, and CDT contributed to the interpretation of the results. AK took the lead in writing the manuscript. All authors provided critical feedback and helped shape the research, analysis and manuscript.

Data Availability Statement

The data supporting this study's findings are available from the corresponding author upon reasonable request.

Animal Welfare

The authors confirm that they have adhered to ARRIVE Guidelines to protect animals used for scientific purposes.

References

- 1. Abenavoli L, Capasso R, Milic N, et al (2010): *Milk thistle in liver diseases: past, present, future.* Phytother Res, 24, 1423-1432.
- 2. Abenavoli L, Izzo AA, Milić N, et al (2018): Milk thistle (Silybum marianum): A concise overview on its chemistry, pharmacological, and nutraceutical uses in liver diseases. Phytother Res, **32**, 2202-2213.
- **3.** Aghel N, Kalantari H, Rezazadeh S (2011): Hepatoprotective effect of Ficus carica leaf extract on mice intoxicated with carbon tetrachloride. Iran J Pharm Res, **10**, 63-68.
- 4. Ali B, Mujeeb M, Aeri V, et al (2012): Anti-inflammatory and antioxidant activity of Ficus carica Linn. Leaves. Nat Prod Res, 26, 460-465.
- 5. Al-Olayan EM, El-Khadragy MF, Aref AM, et al (2014): The potential protective effect of Physalis peruviana L. against carbon tetrachloride-induced hepatotoxicity in rats is mediated by suppression of oxidative stress and

downregulation of MMP-9 expression. Oxid Med Cell Longev, 2014, 381413.

- Al-Rasheed N, Faddah L, Al-Rasheed N, et al (2016): Protective effects of silymarin, alone or in combination with chlorogenic acid and/or melatonin, against carbon tetrachloride-induced hepatotoxicity. Pharmacogn Mag, 12, S337-S345.
- Alshatwi AA, Hasan TN, Shafi G, et al (2012): Validation of the antiproliferative effects of organic extracts from the green husk of Juglans regia L. on PC-3 human prostate cancer cells by assessment of apoptosis-related genes. Evid Based Complement Alternat Med, 2012, 103026.
- 8. Aslan A, Can MI (2014): Milk thistle impedes the development of carbontetrachloride-induced liver damage in rats through suppression of bcl-2 and regulating caspase pathway. Life Sci, 117, 13-18.
- **9.** Aydın S, Gökçe Z, Yılmaz Ö (2015): The effects of Juglans regia L. (walnut) extract on certain biochemical paramaters and in the prevention of tissue damage in brain, kidney, and liver in CCl4 applied Wistar rats. Turk J Biochem, **40**, 241-250.
- Beutler E (1982): Red cell metabolism: A manual of biochemical methods. 3rd edition. Grune and Stratton; New York.
- **11. Bradford MM** (1976): *A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding*. Anal Biochem, **72**, 248-254.
- 12. Clawson GA (1989): *Mechanisms of carbon tetrachloride hepatotoxicity*. Pathol Immunopathol Res, **8**, 104-112.
- 13. Colak E, Ustuner MC, Tekin N, et al (2016): The hepatocurative effects of Cynara scolymus L. leaf extract on carbon tetrachloride-induced oxidative stress and hepatic injury in rats. Springerplus, 5, 216.
- **14.** De La Puerta R, Martinez E, Bravo L, et al (1996): *Effect* of silymarin on different acute inflammation models and on leukocyte migration. J Pharm Pharmacol, **48**, 968-670.
- Demirci C, Gargili A, Kandil A, et al (2006): Inhibition of inducible nitric oxide synthase in murine visceral larva migrans: effects on lung and liver damage. Chin J Physiol, 49, 326-334.
- Donga D, Xua L, Yina L, et al (2015): Naringin prevents carbon tetrachloride-induced acute liver injury in mice. J Funct Foods, 12, 179-191.
- Eassawy MMT, Ismail AFM (2024): Protective effect of chicory and/or artichoke leaves extracts on carbon tetrachloride and gamma-irradiation-induced chronic nephrotoxicity in rats. Environmental Toxicology, 39, 1666-1681.
- **18. Eidi A, Moghadam JZ, Mortazavi P, et al** (2013): Hepatoprotective effects of Juglans regia extract against CCl4-induced oxidative damage in rats. Pharm Biol, **51**, 558-565.
- El-Boshy M, Ashshi A, Gaith M, et al (2017): Studies on the protective effect of the artichoke (Cynara scolymus) leaf extract against cadmium toxicity-induced oxidative stress, hepatorenal damage, and immunosuppressive and hematological disorders in rats. Environ Sci Pollut Res Int, 24, 12372-12383.
- **20. Elmas O, Erbas O, Yigitturk G** (2016): The efficacy of Aesculus hippocastanum seeds on diabetic nephropathy in

http://vetjournal.ankara.edu.tr/en/

a streptozotocin-induced diabetic rat model. Biomed Pharmacother, **83**, 392-396.

- Florek E, Szukalska M, Markiewicz K, et al (2023): Evaluation of the protective and regenerative properties of commercially available artichoke leaf powder extract on plasma and liver oxidative stress parameters. Antioxidants (Basel), 12, 1846.
- Gebhardt R (1997): Antioxidative and protective properties of extracts from leaves of the artichoke (Cynara scolymus L.) against hydroperoxide-induced oxidative stress in cultured rat hepatocytes. Toxicol Appl Pharmacol, 144, 279-286.
- 23. Guarrera PM (2005): Traditional phytotherapy in Central Italy (Marche, Abruzzo, and Latium). Fitoterapia, 76, 1-25.
- 24. Gurel E, Ustunova S, Ergin B, et al (2013): Herbal haemorrhoidal cream for haemorrhoids. Chin J Physiol, 56, 253-262.
- 25. Haque R, Bin-Hafeez B, Parvez S, et al (2003): Aqueous extract of walnut (Juglans regia L.) protects mice against cyclophosphamide-induced biochemical toxicity. Hum Exp Toxicol, 22, 473-480.
- 26. Hosseini SA, Mohammadi J, Delaviz H, et al (2018): Effect of Juglans regia and Nasturtum officinalis biochemical parameters following toxicity of kidney by CCl4 in Wistar rats. Electronic Journal of General Medicine, 15.
- 27. Jiang G, Sun C, Wang X, et al (2022): Hepatoprotective mechanism of Silybum marianum on nonalcoholic fatty liver disease based on network pharmacology and experimental verification. Bioengineered, 13, 5216-5235.
- Jiang N, Xin W, Wang T, et al (2011): Protective effect of aescin from the seeds of Aesculus hippocastanum on liver injury induced by endotoxin in mice. Phytomedicine, 18, 1276-1284.
- **29.** Katopodis N, Stock CC (1980): Improved method to determine lipid bound sialic acid in plasma or serum. Res Commun Chem Pathol Pharmacol, **30**, 171-180.
- **30.** Kraft K (1997): Artichoke leaf extract Recent findings reflecting effects on lipid metabolism, liver and gastrointestinal tracts. Phytomedicine, **4**, 369-378.
- **31.** Küçükkurt I, Ince S, Keleş H, et al (2010): Beneficial effects of Aesculus hippocastanum L. seed extract on the body's own antioxidant defense system on subacute administration. J Ethnopharmacol, **129**, 18-22.
- **32.** Legrand M, Bezemer R, Kandil A, et al (2011): The role of renal hypoperfusion in development of renal microcirculatory dysfunction in endotoxemic rats. Intensive Care Med, **37**, 1534-1542.
- **33.** Madrigal-Santillán E, Madrigal-Bujaidar E, Álvarez-González I, et al (2014): *Review of natural products with hepatoprotective effects*. World J Gastroenterol, **20**, 14787-14804.
- 34. Matsuda H, Li Y, Murakami T, et al (1997): Effects of escinsIa, Ib, IIa, and IIb from horse chestnut, the seeds of Aesculus hippocastanum L., on acute inflammation in animals. Biol Pharm Bull, 20, 1092-1095.
- **35.** Mbarki S, Alimi H, Bouzenna H, et al (2017): Phytochemical study and protective effect of Trigonella foenum graecum (Fenugreek seeds) against carbon tetrachloride-induced toxicity in liver and kidney of male rat. Biomed Pharmacother, **88**, 19-26.

- **36.** Mehmetçik G, Ozdemirler G, Koçak-Toker N, et al (2008): Effect of pretreatment with artichoke extract on carbon tetrachloride-induced liver injury and oxidative stress. Exp Toxicol Pathol, **60**, 475-480.
- 37. Mohan GK, Pallavi E, Kumar R, et al (2007): Hepatoprotective activity of Ficus carica Linn leaf extract against carbon tetrachloride-induced hepatotoxicity in rats. DARU Journal of Pharmaceutical Sciences, 15, 162-166.
- **38.** Mujeeb M, Alam Khan S, Aeri V, et al (2011): Hepatoprotective activity of the ethanolic extract of Ficus carica Linn. leaves in carbon tetrachloride-induced hepatotoxicityin rats. Iran J Pharm Res, **10**, 301-306.
- **39. Muriel P** (2007): Some experimental models of liver damage. S. Sahu (Ed.), Hepatotoxicity: from genomics to in vitro and in vivo models. John Wiley and Sons LTD, West Sussex, UK p. 119-137.
- 40. Nasef MA, Yousef MI, Ghareeb DA, et al (2023): Hepatoprotective effects of a chemically-characterized extract from artichoke (Cynara scolymus L.) against AFB1induced toxicity in rats. Drug and Chemical Toxicology, 46, 1070-1082.
- **41.** Nawaz H, Rehman T, Shahzad H, et al (2024): Hepatoprotective activity of fruit and leaf extracts of Ficus carica and Ficus benghalensis in experimental rats. JAPS, **34**, 168-176.
- Ohkawa H, Ohishi N, Yagi K (1979): Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal Biochem, 95, 351-358.
- Oliveira I, Sousa A, Ferreira IC, et al (2008): Total phenols, antioxidant potential and antimicrobial activity of walnut (Juglans regia L.) green husks. Food Chem Toxicol, 46, 2326-2331.
- 44. Özkan EE, Mesut B, Polat A, et al (2016): In vitro skin permeation of escin in the new gel formulation of Aesculus hippocastanum (Horse Chestnut). Journal of Faculty of Pharmacy of Istanbul University, 46, 79-88.
- **45.** Paglia DE, Valentine WN (1967): Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. J Lab Clin Med, **70**, 158-169.
- 46. Parveen R, Baboota S, Ali J, et al (2011): Effects of silymarin nanoemulsion against carbon tetrachlorideinduced hepatic damage. Arch Pharm Res, N34, 767-774.
- **47.** Peng W, Zhang C, Lv H, et al (2010): Comparative evaluation of the protective potentials of human paraoxonase 1 and 3 against CCl4-induced liver injury. Toxicol Lett, **193**, 159-166.
- **48.** Pèrez C, Canal JR, Torres MD, et al (2003): Experimental diabetes treated with Ficus carica extract: effect on oxidative stress parameters. Acta Diabetol, **40**, 3-8.
- **49.** Pérez C, Domínguez E, Canal JR, et al (2000): Hypoglycaemic activity of an aqueous extract from Ficus carica (fig tree) leaves in streptozotocin diabetic rats. Pharm Biol, **38**, 181-186.
- **50.** Pittler MH, Ernst E (1998): Horse-chestnut seed extract for chronic venous insufficiency, a criteria-based systematic review. Arch Dermatol, **134**, 1356-1360.
- **51. Rainone F** (2005): *Milk thistle*. Am Fam Physician, **72**, 1285-1288.
- **52.** Shaker E, Mahmoud H, Mnaa S (2010): Silymarin, the antioxidant component and Silybum marianum extracts prevent liver damage. Food Chem Toxicol, **48**, 803-806.

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- **53.** Singab AN, Ayoub NA, Ali EN, et al (2010): Antioxidant and hepatoprotective activities of Egyptian moraceous plants against carbon tetrachloride-induced oxidative stress and liver damage in rats. Pharm Biol, **48**, 1255-1264.
- 54. Singh H, Sidhu S, Chopra K, et al (2017): The novel role of β -aescin in attenuating CCl₄-induced hepatotoxicity in rats. Pharm Bio, 55, 749-757.
- Sirtori CR (2001): Aescin: pharmacology, pharmacokinetics and therapeutic profile. Pharmacol Res, 44, 183-193.
- Stampar F, Solar A, Hudina M, et al (2006): Traditional walnut liqueur-cocktail of phenolics. Food Chemistry, 95, 627-631.
- **57.** Tacke F, Luedde T, Trautwein C (2009): *Inflammatory pathways in liver homeostasis and liver injury*. Clin Rev Allergy Immunol, **36**, 4-12.
- Wellington K, Jarvis B (2001): Silymarin: a review of its clinical properties in the management of hepatic disorders. BioDrugs, 15, 465-489.
- **59.** Winterbourn CC, Hawkins RE, Brian M, et al (1975): The estimation of red cell superoxide dismutase activity. J Lab Clin Med, **85**, 337-341.
- **60.** Yang CC, Fang JY, Hong TL, et al (2013): Potential antioxidant properties and hepatoprotective effects of an aqueous extract formula derived from three Chinese

medicinal herbs against CCl(4)-induced liver injury in rats. Int Immunopharmacol, **15**, 106-113.

- **61.** Yassin NYS, AbouZid SF, El-Kalaawy AM, et al (2022): Silybum marianum total extract, silymarin and silibinin abate hepatocarcinogenesis and hepatocellular carcinoma growth via modulation of the HGF/c-Met, Wnt/β-catenin, and PI3K/Akt/mTOR signaling pathways. Biomed Pharmacother, **145**, 112409.
- Zhang DG, Zhang C, Wang JX, et al (2017): Obeticholic acid protects against carbon tetrachloride-induced acute liver injury and inflammation. Toxicol Appl Pharmacol, 314, 39-47.
- **63.** Zhang R, Hu Y, Yuan J, et al (2009): Effects of Puerariae radix extract on the increasing intestinal permeability in rat with alcohol-induced liver injury. J Ethnopharmacol, **126**, 207-214.
- **64.** Zhu X, Zhang H, Lo R, et al (2005): Antimicrobial activities of Cynara scolymus L. leaf, head, and stem extracts. Journal of Food Science, **70**, M149-M152.

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