

Published by Ege Animal Science Association
Ege Zootekni Derneđi Yayınıdır.

ISSN 1301-9597



JOURNAL OF ANIMAL PRODUCTION

Hayvansal Üretim

ISSN 1301-9597

JOURNAL OF ANIMAL PRODUCTION

Hayvansal Üretim

YEAR
YIL

2018

VOLUME
CİLT

59

NUMBER
SAYI

1



Published by Ege Animal Science Association
Ege Zootekni Derneği Yayınıdır



IMPORTANT INFORMATION (Önemli Bilgi)

Number of citations is a vital criterion for not only the articles but also evaluation of the journals. It's noticed that there have been some wrong citations in the Journal of Animal Production.

*Atıf sayısı hem makalelerin hem de dergilerin değerlendirilmesinde önemli bir kriterdir. Yapılan atıflar incelendiğinde **Hayvansal Üretim** dergisindeki makalelere bazen doğru atıf yapılmadığı saptanmıştır.*

It must be written the name of the journal as “**Hayvansal Üretim**” when used for citation. If used in English, the name of the journal must be “**Journal of Animal Production**”.

*Atıflarda derginin adı “**Hayvansal Üretim**” olarak yazılmalıdır. Dergi adı İngilizce olarak yazılacaksa “**Journal of Animal Production**” kullanılmalıdır.*

Journal name of abbreviation must be “**Hay. Üret.**” as Turkish, but in English “**J. Anim. Prod.**” Except for obligatory situations, Turkish name of the journal and abbreviation should be preferred.

*Dergi adı kısaltmaları Türkçe olarak “**Hay. Üret.**”, İngilizce olarak ise “**J. Anim. Prod.**” şeklinde olmalıdır. Zorunlu haller dışında Türkçe isim ve kısaltma tercih edilmelidir.*



Journal of Animal Production

indexed by

Hayvansal Üretim aşağıdaki indekslerce taranmaktadır

- Ulusal Akademik Ağ ve Bilgi Merkezi (ULAKBİM), 2001
- CAB Abstracts, 2001
- AgBiotechNet, 2001
- Index Copernicus Journal Master List, 2008
- EBSCO, 2018
- Bielefeld Academic Reserch Engine (BASE), 2018
- ResearchBib, 2018



JOURNAL OF ANIMAL PRODUCTION

(HAYVANSAL ÜRETİM)

Year (Yıl): 2018 Volume (Cilt): 59 Number (Sayı): 1

Publisher on Behalf of Turkish Animal Science Association

(Ege Zootekni Derneği Adına Sahibi)

Prof. Dr. Nedim KOŞUM
Dernek Başkanı

Editor in Chief

(Baş Editör)

Prof. Dr. Nedim KOŞUM

Managing Editors

(Editör Yardımcısı)

Arş. Gör. Dr. Çağrı KANDEMİR

Editorial Board in Alphabetical Order of Name

(Editörler Kurulu)

Prof. Dr. Abdullah CAN
Dr. Öğr. Üye. Abdullah Nuri ÖZSOY
Prof. Dr. Ahmet GÜLER
Prof. Dr. Ahmet ŞAHİN
Prof. Dr. Atakan KOÇ
Prof. Dr. Banu YÜCEL
Prof. Dr. Cemal ÜN
Prof. Dr. Çiğdem TAKMA
Prof. Dr. Ethem AKYOL
Prof. Dr. Figen KIRKPINAR
Prof. Dr. Güldehen BİLGİN
Prof. Dr. Hayati KÖKNAROĞLU
Prof. Dr. Hayrettin OKUT
Prof. Dr. Hatice B. MALAYOĞLU
Prof. Dr. İbrahim CEMAL
Doç. Dr. İbrahim KAYA
Doç. Dr. İsmail DURMUŞ
Prof. Dr. Mahmut KESKİN
Prof. Dr. Mesut TÜRKOĞLU
Prof. Dr. Mehmet İhsan SOYSAL
Prof. Dr. Mehmet KOYUNCU
Prof. Dr. Mehmet KURAN
Doç. Dr. Muazzez CÖMERT
Prof. Dr. Muhiittin ÖZDER
Prof. Dr. Muhammet ALAN
Prof. Dr. M. Soner BALCIOĞLU
Prof. Dr. Mustafa AKŞİT
Prof. Dr. Muzaffer DENLİ
Prof. Dr. Mürsel ÖZDOĞAN
Prof. Dr. Numan ÖZCAN
Prof. Dr. Ömer Cevdet BİLGİN
Prof. Dr. Servet YALÇIN
Prof. Dr. Sezen ÖZKAN
Prof. Dr. Sinan Sefa PARLAT
Prof. Dr. Şenay SARICA
Prof. Dr. Turgay ŞENGÜL
Prof. Dr. Turgay TAŞKIN
Prof. Dr. Turgut AYGÜN
Prof. Dr. Türker SAVAŞ
Prof. Dr. Yusuf KONCA
Prof. Dr. Zafer ULUTAŞ

acan@harran.edu.tr
nuriozsoy@sdu.edu.tr
aguler@omu.edu.tr
ahmet.sahin@ahievran.edu.tr
akoc@adu.edu.tr
banu.yucel@ege.edu.tr
cemal.un@ege.edu.tr
cigdem.takma@ege.edu.tr
eakyol@ohu.edu.tr
figen.kirkpinar@ege.edu.tr
guldehen.bilgen@ege.edu.tr
hayatikoknaroglu@sdu.edu.tr
hokut@yyu.edu.tr
hatice.basmacioğlu@ege.edu.tr
icemal@adu.edu.tr
ibrahim.kaya@ege.edu.tr
idurmus@odu.edu.tr
mkeskin@mku.edu.tr
mturk@agri.ankara.edu.tr
misoysal@nku.edu.tr
koyuncu@uludag.edu.tr
mkuran@omu.edu.tr
muazzez.comert@ege.edu.tr
mozder@nku.edu.tr
muhammetalan@ogu.edu.tr
msoner@akdeniz.edu.tr
maksit@adu.edu.tr
mdenli@dicle.edu.tr
mozdogan@adu.edu.tr
nozcan@cu.edu.tr
ocbilgin@atauni.edu.tr
servet.yalcin@ege.edu.tr
sezen.ozkan@ege.edu.tr
sparlat@selcuk.edu.tr
senay.sarica@gop.edu.tr
tsengül@bingol.edu.tr
turgay.taskin@ege.edu.tr
taygunyyu.edu.tr
tsavas@comu.edu.tr
yusufkonca@erciyes.edu.tr
zaferulutas@ohu.edu.tr

Harran University, ŞANLIURFA
Süleyman Demirel University, ISPARTA
Ondokuz Mayıs University, SAMSUN
Ahi Evran University, KIRŞEHİR
Adnan Menderes University, AYDIN
Ege University, IZMİR
Ege University, IZMİR
Ege University, IZMİR
Ömer Halisdemir University, NİĞDE
Ege University, IZMİR
Ege University, IZMİR
Süleyman Demirel University, ISPARTA
Yüzüncü Yıl University, VAN
Ege University, IZMİR
Adnan Menderes University, AYDIN
Ege University, IZMİR
Ordu University, ORDU
Mustafa Kemal University, HATAY
Ankara University, ANKARA
Namık Kemal University, TEKİRDAĞ
Uludağ University, BURSA
Ondokuz Mayıs University, SAMSUN
Ege University, IZMİR
Namık Kemal University TEKİRDAĞ
Osmangazi University, ESKİŞEHİR
Akdeniz University, ANTALYA
Adnan Menderes University, AYDIN
Dicle University, DİYARBAKIR
Adnan Menderes University, AYDIN
Çukurova University, ADANA
Atatürk University, ERZURUM
Ege University, IZMİR
Ege University, IZMİR
Selçuk University, KONYA
Gaziosmanpaşa University, TOKAT
Bingöl University, BİNGÖL
Ege University, IZMİR
Yüzüncü Yıl University, VAN
Onsekiz Mart University, ÇANAKKALE
Erciyes University, KAYSERİ
Ömer Halisdemir University, NİĞDE

The referees list / Hakem listesi

Journal of Animal Production is a peer-reviewed journal. List of referees is given in the last press issue of the year.

Hayvansal Üretim hakemli bir dergi olup, hakem listesi her yılın son sayısında basılı yayınlanmaktadır.

Journal of Animal Production is published two times in a year (May and November) by Ege Animal Science Association in Turkey. Detail information about Ege Animal Science Association and Journal of Animal Science could be finding from the web site of the Ege Animal Science Association or correspondence address of the journal given below. Guidelines to authors are also given at the end of each issue of the journal.

Hayvansal Üretim dergisi, Ege Zootekni Derneği'nin "yaygın süreli" bir yayınıdır. Yılda iki kez (Mayıs ve Kasım aylarında) yayınlanmaktadır. Ege Zootekni Derneği ve Hayvansal Üretim dergisine ilişkin ayrıntılı ve güncel bilgiler Ege Zootekni Derneği'nin internet sitesinden veya dergi yazışma adresinden öğrenilebilir. Yazım kuralları derginin her sayısının sonunda verilmektedir.

Correspondence Address (Dergi İçin Yazışma Adresi):

Prof. Dr. Nedim KOŞUM

Journal of Animal Production Editor in Chief

Ege Universty, Faculty of Agriculture, Deperment of Animal Science

35100 Bornova, İzmir-TURKEY

Phone (Tel): +90 (232) 311 2718 (sekreter) **Fax:** +90 (232) 388 1867

E-posta (e-mail): nedim.kosum@ege.edu.tr, cagri.kandemir@ege.edu.tr

All rights reserved. No part of this publication may be reproduced in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without the prior permission of the publisher.

Bu derginin yayın hakları Ege Zootekni Derneği'ne aittir. Derginin hiçbir bölümü, yayıncının izni olmaksızın, elektronik, mekanik veya başka bir yöntemle, herhangi bir şekilde çoğaltılamaz.

Ege Zootekni Derneği Yönetim Adresi:

Fevzipaşa Bulvarı No: 17 Azim Han K:4 D:408 Konak / İZMİR

Basım Yeri : Ege Üniversitesi Rektörlüğü Basımevi Müdürlüğü

No: 172/134 Kampüsü / Bornova, İzmir

Tel : 0232 311 18 19

e-mail : bsmmd@mail.ege.edu.tr

TC Kültür ve Turizm Bakanlığı Sertifika No: 18679

Baskı Tarihi:

31 Temmuz 2018



JOURNAL OF ANIMAL PRODUCTION

(Hayvansal Üretim)

YEAR 2018
YIL

VOLUME 59
CİLT

NUMBER 1
SAYI

CONTENTS (İçindekiler)

RESEARCH ARTICLES (Araştırma Makaleleri)

- A Study on the Change in Postpartum Immunoglobulins of Goats and Kids**
Keçi ve Oğlaklarda Doğum Sonrası İmmünoglobulinlerin Değişimi Üzerine Bir Araştırma
Nedim KOŞUM, Turgay TAŞKIN, Özer KINIK, Çağrı KANDEMİR, Ecem AKAN..... 1
- Yumurtacı Tavuk Rasyonlarına İlave Edilen Esansiyel Yağ ve Organik Asit Karışımının Performans, Yumurta Verimi ve Kalite Parametreleri Üzerine Etkisi**
Effect of Supplementation Essential Oil and Organic Acid Mixture on Performance, Egg Production and Egg Quality Parameters in Laying Hens
Özlem KARADAĞOĞLU, Mükremin ÖLMEZ, Bülent ÖZSOY, Tarkan ŞAHİN..... 9
- Effects of Pre-milking Resting on Some Lactation Characteristics in Damascus (Shami) and Kilis Goats**
Şam ve Kilis Keçilerinde Sağım Öncesi Dinlendirmenin Bazı Süt Verim Özellikleri Üzerine Etkileri
Sabri GÜL, Mahmut KESKİN, Zehra GÜLER, Ahmet DURSUN, Zuhal GÜNDÜZ,
Süleyman Ercüment ÖNEL, Dilek TÜNEY BEBEK..... 17
- Hatay İli Damızlık Sığır Yetiştiricileri Birliği Üyesi İşletmelerin Sosyo-Ekonomik Özellikleri**
Socio-Economic Characteristics of Members of Cattle Breeders Association in Hatay Province
Nuran TAPKI, İbrahim TAPKI, Erdal DAĞISTAN, Muhammet Hanifi SELVİ, Aybüke KAYA, Yusuf Ziya GÜZEY,
Bekir DEMİRTAŞ, Ahmet Duran ÇELİK..... 25
- Yetiştirici Koşullarında Kıvrırcık Koyunlarının Bazı Döl Verimi Özellikleri**
Some Fertility Traits of Kıvrırcık Sheep in Rural Farms
Mehmet KOYUNCU, Hilal AKGÜN..... 33
- Saanen Keçilerinin Entansif Koşullarda Bazı Verim Özelliklerinin Belirlenmesi Üzerine Bir Araştırma**
A Study on the Determination of Some Production Characteristics of Saanen Goats in Intensive Conditions
Çağrı KANDEMİR, Turgay TAŞKIN, Nedim KOŞUM..... 41

REVIEWS (Derlemeler)

- Alternatif Protein Kaynaklarının Hayvan Beslemede Kullanım Olanakları**
Possible Usage of Alternative Protein Sources in Animal Nutrition
Hasan Hüseyin İPÇAK, Sema ÖZÜRET MEN, Ahmet ALÇİÇEK, Hülya ÖZELÇAM..... 51
- Importance of Characterization of the Vaginal Microbiota in Ewes and Nannies**
Koyun ve Keçilerde Vaginal Mikrobiota Karakterizasyonun Önemi
Şeniz ÖZİŞ ALTINÇEKİÇ, Mehmet KOYUNCU..... 59
- Buzağılarda Yaşama Gücünün Anahtarı “Kolostrum”**
Key of Survival in Calves “Colostrum”
Mehmet KOYUNCU, Merve KARACA..... 67
- Effects of Feed Additives Used As an Alternative to Antibiotics on Mineral Absorption and Bone Characteristics in Poultry: A Review**
Antibiyotiklere Alternatif Olarak Kullanılan Yem Katkı Maddelerinin Kanatlılarda Mineral Emilimine ve Kemik Karakteristiklerine Etkileri: Derleme
Ahmet Önder ÜSTÜNDAĞ, Mürsel ÖZDOĞAN..... 79

Research Article
(Araştırma Makalesi)



J. Anim. Prod., 2018, 59 (1):1-8
DOI: 10.29185/hayuretim.394535

Nedim KOŞUM¹

Turgay TAŞKIN¹

Özer KINIK²

Çağrı KANDEMİR¹

Ecem AKAN²

¹Ege Üniversitesi, Ziraat Fakültesi, Zootekni Bölümü, 35000, İzmir / Türkiye

²Ege Üniversitesi, Ziraat Fakültesi, Süt Teknolojisi Bölümü, 35000, İzmir / Türkiye

sorumlu yazar: turgay.taskin@ege.edu.tr

A Study on the Change in Postpartum Immunoglobulins of Goats and Kids

Keçi ve Oğlaklarda Doğum Sonrası İmmünoglobulinlerin Değişimi Üzerine Bir Araştırma

Alınış (Received): 13.01.2018

Kabul tarihi (Accepted): 20.05.2018

Key Words:

Saanen goat, intensive conditions, fertility, growth traits, milk yield

Anahtar Kelimeler:

Saanen keçisi, entansif koşullar, döl verimi, gelişme özellikleri, süt verimi

ABSTRACT

Objective: In the study, the changes in the immunoglobulin levels of the Saanen goats fed with colostrum were determined during the three days after birth.

Material and Methods: The animal material of the research consists of 11 goats and 11 of their kids. Colostrum samples were taken from the goats in three repetitions (at birth, at 24th and 48th hours after birth). Changes in IgA, IgM, and IgG levels were examined individually in 33 (11 x 3) colostrum samples taken at birth and 24th and 48th hours after birth.

Results: IgA, IgM and IgG levels at birth and 24 and 48 hours after birth for Saanen goats were 0.11-1.98-1.88 mg/ml, 0.28-0.95-14.01 mg/ml and 0.25-0.96-13.53 mg/ml, respectively. The IgA, IgM, IgG levels of kids at birth, 24, and 48 hours after birth were 0.76-1.11-19.22 mg/ml, 0.58-1.02-18.42 mg/ml and 0.53-1.24-21.60 mg/ml, respectively. The effect of birth type and gender and parity on IgA, IgM, and IgG levels were not significant, while the effect of the time-dependent change was linearly and quadratically significant ($P < 0.01$).

Conclusion: In the colostrum secreted in the postpartum period in goats, it is necessary for the immune substances to be taken as soon as possible since the rate of passage of the immune substances transferred through the intestinal epithelium by this way decreases in time.

ÖZ

Amaç: Bu çalışmada; doğumu izleyen üç gün boyunca kolostrumla beslenen Saanen oğlaklarında kolostrumda bulunan bağışıklık maddelerinin değişimi belirlenmiştir.

Materyal ve Metot: Araştırmancın hayvan materyalini; 11 baş saf Saanen keçisi ile bunlardan doğan 11 baş oğlak olmak üzere toplam 22 baş hayvan oluşturmaktadır. Her keçiden toplam 3 kez (doğumda ve bunu izleyen saatlerde 24. ve 48. saatte) kolostrum örneği alınmıştır. Çalışmada, doğum, 24. ve 48. saatte alınan toplam 33 (11 x 3) ağız sütü (kolostrum) örneğinde IgA, IgM, IgG düzeylerinin değişimleri bireysel olarak incelenmiştir.

Bulgular: Saanen keçilerinde IgA, IgM, IgG düzeyleri doğumda; 0.11-1.98-1.88mg/ml, 24 saat sonra 0.28-0.95-14.01 mg/ml ve 48 saat sonra 0.25-0.96-13.53 mg/ml değerleri arasında değişmiştir. IgA, IgM, IgG nin oğlak kanlarında doğum, 24 ve 48. saatlerdeki düzeyleri sırasıyla; 0.76-1.11-19.22 mg/ml, 0.58-1.02-18.42 mg/ml ve 0.53-1.24-21.60 mg/ml değerleri arasında değişmiştir. IgA, IgM, IgG düzeyleri üzerinde oğlak doğum tipi, ve cinsiyeti ile laktasyon sırasının etkisi önemsiz, zamana bağlı olarak değişimin etkisi gerek lineer gerekse kuadratik olarak önemli bulunmuştur ($P < 0.01$).

Sonuç: Keçilerde doğum sonrası dönemde salgılanan kolostrumda yavruya bu yolla aktarılan bağışıklık maddelerinin bağırsak epitel dokudan geçiş hızı azaldığı için oğlakların bağışıklık maddelerinin en kısa sürede alması sağlanmalıdır.



INTRODUCTION

In recent years, the importance placed on small ruminants has increased in comparison to other domestic animals (Lerias et al. 2014). In developing countries, small ruminant dairy farms have been applying artificial rearing methods, depending on the increase in marketable milk amount (Morales et al. 2014). There are three critical periods in the newborn ruminants' immune system development during the first two months after birth (Sherman et al. 1990), which are the colostrum-feeding period, milk-feeding period and the post-weaning period, respectively. Flock management methods applied during these periods affect the growth performance of animals (Demirören et al. 1995; Mastellone et al. 2011).

Colostrum intake time and amount during the first 72 hours are critical in the transfer of passive immune substances to ruminant farm animals, such as, cattle, sheep and goat, and in the resultant viability of the offspring (Stelwagen et al. 2009; Hernandez et al. 2014a). This is due to the hypogammaglobulinemia of the offspring. Artificially reared offspring may require bottle-feeding during the first couple of days postpartum to allow sufficient immunoglobulin intake through colostrum intake (Morales et al. 2011). However, the amount and composition of colostrum production vary depending on various factors including feeding type and the number of offspring per birth (Banchemo et al. 2004). Insufficient colostrum intake in lambs and goat kids during the first hours after birth results in increased susceptibility to diseases and increased mortality (Ahmad et al. 2000; Novak and Poindron, 2006).

Colostrum feeding time is another factor affecting the immune status, considering the fact that using undernourished offspring as breeding stock may sometimes cause certain problems (Hernandez et al. 2014b). IgG levels are high in the blood during the first 12-36 hours after birth, and for ruminant animals, absorption of IgG in the body, which is transferred to colostrum, is of great importance (Chen et al. 1999). Researches on the effect of possible delays in colostrum intake on immunoglobulins in the bloodstream continue (Ahmadi et al. 2009). Two of the most important variables regarding immunoglobulins are the levels of IgG and IgM in the blood. In newborn goat kids, passive immunity is obtained through colostrum, which protects the animals that were the first to respond to any pathogen type or suspected of carrying infectious diseases (Ahmadi et al. 2009). Until the immune system of the offspring is developed, immunoglobulin level in the blood continues to increase for a certain

period and increasing colostrum increases the viability of the offspring (Alves et al. 2015).

In intensive goat dairy farms, goat kids are immediately separated from their mothers to avoid or minimize mother-offspring bond (Ramirez et al. 1996). However, colostrum sanitation is imperative in locations where the risk of goat disease transmission, such as, CAE (caprine arthritis encephalitis), is high (Rachman et al. 2015). It was reported that pasteurization and alternative methods used in colostrum hygiene decreased the IgG amounts in colostrum by 20-30% (Arguello, 2011; Trujilo et al. 2007) and thereby IgG concentration in either lyophilized or atomized colostrum remained unchanged (Castro et al. 2011). On the other hand, it was also reported that this recently highlighted negative condition may be avoided using certain methods. CAE virus can be neutralized through heat treatment to colostrum (Castro et al. 2011), although IgG and other colostrum components are affected by heat and immune system of animals may be hindered. For example, in a study carried out using pasteurized cow colostrum, IgG and lactoferrin density and neutrophil activity were decreased in calves (Aldomy et al. 2014). Morin et al. (2007) in their study on calves, and, Constant et al. (1994), in their study on goat kids, reported insufficient IgG absorption in colostrum. However, whether maternal and exogenous IgG have a synergetic effect on the gamma-globulin concentration of newborn goat kids is not well known. This study aimed to determine the changes in immunoglobulin levels in Saanen goats during colostrum feeding.

MATERIAL and METHOD

Animal Material

In the study, totally 22 head animals, 11 head Saanen goats, 11 heads Saanen kids were used as experimental materials.

Method

Colostrum Feeding to Goat Kids

After birth, the kids were immediately separated from their mothers and fed colostrum weighing at least 10% of their live weight with feeding bottles. During the first hour postpartum, after the umbilical cords of the goat kids were disinfected, ear tags of the goat kids were placed and their birth weights were individually measured immediately after birth (Marounek et al. 2012; Chigerwe et al. 2005).



Blood Sample Collection

To individually determine the changes in IgA, IgM and IgG levels, 2 cc blood samples were taken from the vena jugularis of the goats, starting from at least 3 days prior to birth and during the 4 days after birth, and 1 cc blood samples were taken from the goat kids during the 3 days after birth (Quantispeed goat test, QGT) (Chigerwe et al. 2005).

Determination of the Colostrum Samples and IgG

Prior to the first milk feeding to goat kids, 500-ml colostrum samples were taken from each goat; then, they were taken from each goat in every 24 hours and thereby, a total of 3 colostrum samples were collected per a goat during the trial. The changes in IgA, IgM and IgG levels were individually determined for each colostrum sample, a total of 33 samples collected at the birth, 24th and 48th hours (Quantispeed goat test, QGT) (Dale et al. 2009).

Milk Analyses

The dry matter and fat values of milk were determined in accordance with TS 1018 (Anonym, 1989); determination of lactose was carried out by using the photometric method [30]; determination of mineral matter was carried out in accordance with AOAC (2000); determination of ash was carried out by following the method proposed by Kurt et al. (2007); the Kjeldahl method was used in the calculation of protein ratio (Renner, 1993). The determined nitrogen amount was multiplied by 6.38 to calculate the percentage protein amount.

Data Analyses

In the study, Repeated Measures Factorial Analysis of Variance was used to determine the difference between birth type, gender and parity with respect to investigated properties. Among the multiple comparison tests, the Duncan test was used to determine the differences between the averages

of the days in reference to the variance analysis (Gürbüz ve ark., 2003). In the calculations, SPSS 15 (2007) statistical package program was used.

RESULTS

Table 1 shows the means and standard errors of the immunoglobulin amounts in goat colostrum at birth, 24th, and 48 hours after birth.

Table 1. Least square means and standard errors of the immunoglobulins in goat colostrum

Çizelge 1. Keçi kolostrumundaki bağışıklık maddelerine ait en küçük kareler ortalamaları ve standart hataları

Sampling Time	Investigated Properties	N	Mean (mg/ml)	Standard Error
At birth	IgA	11	0.1105	0.179
	IgM	11	1.9870	0.025
	IgG	11	1.8850	0.041
24 hours after birth	IgA	11	0.2850	0.078
	IgM	11	0.9580	0.031
	IgG	11	14.0160	0.027
48 hours after birth	IgA	11	0.2535	0.435
	IgM	11	0.9640	0.017
	IgG	11	13.5395	0.089

IgA: Immunoglobulin A,

IgM: Immunoglobulin M,

IgG: Immunoglobulin G

Immunoglobulin levels in goat colostrum showed that IgM had the highest level in the colostrum at birth, followed by IgG, while IgA had the lowest level. At the 24th hour after birth, the IgG level increased and reached 14.016 ml, followed by IgM with 0.958 ml. At the 48th hour after birth, immunoglobulin levels in colostrum followed a similar trend to those at the 24th hour; in other words, IgG had the highest level. Table 2 shows the means and standard errors of the immunoglobulin levels determined in the bloods of goat kids at birth and during the hours following birth.

Table 2. Least square means and standard errors of the immunoglobulins in goat kid bloods

Çizelge 2. Oğlak kanlarındaki bağışıklık maddelerine ait en küçük kareler ortalamaları ve standart hataları

Sampling Time	Investigated Properties	N	Mean (mg/ml)	Standard error
At birth	IgA	11	0.7664	0.253
	IgM	11	1.1191	0.050
	IgG	11	19.2295	0.025
24 hours postpartum	IgA	11	0.5895	0.591
	IgM	11	1.0295	0.046
	IgG	11	18.4282	0.026
48 hours postpartum	IgA	11	0.5314	0.535
	IgM	11	1.2482	0.038
	IgG	11	21.6091	0.026

IgA: Immunoglobulin A IgM: Immunoglobulin M IgG: Immunoglobulin G

In the blood samples of the goat kids, IgA level at birth was 0.76 mg/ml and decreased to 0.58 mg/ml

24 hours after birth and reached 0.53 mg/ml 48 hours after birth; in other words, initially, it rapidly



decreased and after 48 hours, its decrease nearly come to a standstill. On the other hand, the IgM level first showed a decreasing trend after 24 hours and, then, showed a tendency to increase around the 48th hour. IgG level in the blood samples of the goat kids firstly decreased until the 24th hour after birth and then, by contrast with other immunoglobulins, increased by almost 50% around the 48th hour after birth.

Table 3 shows the significance levels and least square average and standard error of the significance levels of some of the factors affecting the investigated properties of the blood samples

obtained from the goats. Among the investigated properties, effects of birth type, kid gender and parity were not significant, whereas the effect of time-dependent changes was both quadratically and linearly significant ($P<0.01$).

Table 4 shows the means and standard errors of some factors, which were considered to affect the immunoglobulins in goat kid blood. Among the investigated factors, only the effect of time-dependent changes was significant ($P<0.01$); the effects of birth type, kid gender and parity were not significant. The changes in immunoglobulin levels in kid blood samples from birth to postpartum were both quadratically and linearly significant ($P<0.01$).

Table 3. Least square means and standard errors of birth type, gender, parity and time-dependent immunoglobulin changes in goat bloods

Çizelge 3. Keçi kanlarındaki bağışıklık maddelerinin doğum tipi, cinsiyet, laktasyon sırası ve zamana bağlı olarak değişimine ait en küçük kareler ortalamaları ve standart hataları

Effects	n	IgA (mg/ml)	IgM(mg/ml)	IgG(mg/ml)
Birth Type		NS	NS	NS
Single	5	0.176 ± 0.026	1.358 ± 0.048	9.597 ± 0.374
Twins	6	0.264 ± 0.025	1.283 ± 0.046	9.799 ± 0.361
Gender		NS	NS	NS
Male	5	0.182 ± 0.034	1.387 ± 0.062	10.261 ± 0.485
Female	6	0.259 ± 0.040	1.254 ± 0.074	9.134 ± 0.563
Parity		NS	NS	NS
2	3	0.182 ± 0.032	1.162 ± 0.059	9.311 ± 0.462
3	3	0.114 ± 0.048	1.414 ± 0.048	9.381 ± 0.689
4	5	0.165 ± 0.044	1.385 ± 0.080	10.581 ± 0.623
Time(hour)				
0	11	0.111 ± 0.020	1.978 ± 0.039	1.820 ± 0.072
24	11	0.295 ± 0.027	0.953 ± 0.019	13.730 ± 0.534
48	11	0.255 ± 0.017	1.029 ± 0.069	13.543 ± 0.232
Linear		**	**	**
Quadratic		**	**	**
Overall Mean	11	0.220 ± 0.018	1.320 ± 0.032	9.698 ± 0.250

** : ($P<0.01$) NS: Not Significant

Table 4. Least square means and standard errors of birth type, gender, parity and time-dependent immunoglobulin changes in goat kid bloods

Çizelge 4. Oğlak kanlarındaki bağışıklık maddelerinin doğum tipi, cinsiyet, laktasyon sırası ve zamana bağlı olarak değişimine ait en küçük kareler ortalamaları ve standart hataları

Effects	n	IgA(mg/ml)	IgM(mg/ml)	IgG(mg/ml)
Birth Type		NS	NS	NS
Single	5	0.586 ± 0.111	1.141 ± 0.041	19.295 ± 0.961
Twins	6	0.649 ± 0.073	1.124 ± 0.035	20.009 ± 0.818
Gender		NS	NS	NS
Male	5	0.586 ± 0.111	1.134 ± 0.053	18.382 ± 1.240
Female	6	0.650 ± 0.126	1.131 ± 0.060	20.922 ± 1.409
Parity		NS	NS	NS
2	3	0.681 ± 0.106	1.091 ± 0.051	19.999 ± 1.185
3	3	0.554 ± 0.140	1.163 ± 0.067	19.905 ± 1.568
4	5	0.618 ± 0.142	1.146 ± 0.068	19.052 ± 1.586
Time(hour)				
0	11	0.751 ± 0.059	1.111 ± 0.032	19.209 ± 0.683
24	11	0.575 ± 0.056	1.027 ± 0.032	18.319 ± 0.642
48	11	0.527 ± 0.047	1.259 ± 0.030	21.429 ± 0.618
Linear		**	**	**
Quadratic		**	**	**
Overall Mean	11	0.618 ± 0.054	1.133 ± 0.026	19.652 ± 0.598

NS: Not Significant

** : ($P<0.01$)



Properties of Goat Milk

Table 5 shows descriptive statistics on the changes in the chemical composition of goat milk occurring after birth. Dry matter, fat, protein, casein, lactose, ash and specific weight values of goat colostrum were determined. The average dry matter value of goat colostrum was 24.28%; dry matter values after 24 hours and 48 hours were 17.26% and 13.24%, respectively. As can be seen in Table 5, dry matter values steadily decreased after birth. Fat values in goat colostrum at birth and 24 and 48 hours after birth were 8.27%, 6.76% and 5.59%, respectively. Average protein values of goat colostrum were 10.55%, 6.09%, and 4.23% at birth, 24 hours after birth and 48 hours, respectively.

Table 5. Some descriptive statistics on the chemical composition of goat milk

Çizelge 5. Keçi sütünün kimyasal bileşimine ait bazı tanımlayıcı istatistikler

Period/time	Investigated Property	N	Minimum	Maximum	Mean	Standard Error
At birth	Dry matter (%)	11	18.88	29.08	24.28	0.95
	Fat (%)	11	5.70	9.85	8.27	0.43
	Protein (%)	11	7.04	15.32	10.55	0.78
	Casein (%)	11	4.76	12.14	7.80	0.67
	Lactose (%)	11	3.79	5.83	4.86	0.20
	Ash (%)	11	0.82	2.07	1.18	0.10
	Specific weight (g)	11	1.029	1.074	1.04	0.01
24 hours postpartum	Dry matter (%)	11	13.76	23.07	17.26	0.77
	Fat (%)	11	5.30	8.75	6.76	0.37
	Protein (%)	11	3.81	8.72	6.09	0.43
	Casein (%)	11	3.08	6.91	4.70	0.14
	Lactose (%)	11	2.58	4.30	3.37	0.03
	Ash (%)	11	0.82	1.14	0.90	0.01
	Specific weight (g)	11	1.024	1.040	1.030	0.25
48 hours postpartum	Dry matter (%)	11	12.33	14.83	13.24	0.26
	Fat (%)	11	3.20	7.50	5.59	0.43
	Protein (%)	11	2.76	5.06	4.13	0.22
	Casein (%)	11	2.72	4.01	3.31	0.13
	Lactose (%)	11	2.29	3.79	2.81	0.14
	Ash (%)	11	0.70	1.07	0.84	0.04
	Specific weight (g)	11	1.022	1.034	1.026	0.01

DISCUSSION and CONCLUSION

A general review of the results shows that IgG, IgM and IgA levels determined in our study are in agreement with the results reported in the relevant literature (Singh, 2010; Berry and Broughan, 2007; Keskin ve ark., 2007; Rodrigez et al. 2008, 2009). The source of IgM and IgA in colostrum is not well known. In a study on human colostrum (Moro et al. 1985), researchers reported that phagocytes transferred these immunoglobulins. Some researchers (Islam et al. 2006) reported the presence of plasma cells in human colostrum and determined that these cells produced IgM and IgA. However, it is difficult to adapt

these results to ruminants because, in addition to differences among species in terms of colostrum composition, pre-birth immunoglobulin transfer to offspring is quite hard (Langer, 2009; Moreno et al. 2012b). Another measure of the immune system development in animals is the time of first colostrum feeding to offspring, which also affects the future productivity of adult animals (Pakkanen and Aalto, 1997). For ruminant animals, the first 12-36 hours postpartum is of great importance in terms of IgG intake from colostrum (Castro et al. 2011). A delay in



this period results in decreased viability and increased susceptibility to diseases (Hernandez et al. 2011).

Colostrum intake is of great importance in the viability and passive immunity development in newborn animals (Stelwagen et al. 2009; Hernandez et al. 2014a,b). Therefore, especially artificially reared animals should receive a sufficient intake of colostrum (Morales et al. 2011, 2014). However, the amount and composition of colostrum produced by mothers are affected by a variety of environmental factors including feeding and number of offspring per a birth (Besser and Gay, 1994). Another important issue is the decreased viability due to insufficient colostrum intake during the first couple of hours after birth, which may cause increased death rates (Quigley and Drewry, 1998). Therefore, supplying the most appropriate colostrum source is of great importance. Some studies have focused on using cow milk as the colostrum source in lamb and goat kid rearing (Quigley et al. 2000,2002); however, the most important risk in here is posed by iron deficiency, which may result in anemia (Rodriguez et al. 2009). Therefore, for the easy transfer of immunoglobulins to offspring, species-specific colostrum should be provided to each species (Korhonen, 1998). The most important change in immunoglobulin levels occur in IgG and IgM. Goat kid mortality is closely associated with the IgG level in blood (Moreno et al. 2012a). In addition to optimum colostrum intake amount and composition, optimum temperature, humidity and hygiene conditions should also be provided.

In goat kids, there is a correlation between birth weight and IgG level in blood (Rodriguez et al. 2008). IgG levels in the animals with birth weights below 2.5 kg are especially low during the 12-84 hours after birth (Arguello et al. 2005). Arguello et al. (2004) determined that the effect of birth weight on feeding with colostrum was not significant, whereas, in 1979, Halliday and William (Halliday and Williams, 1979) and, in 2003, Christley et al. (2003) reported that birth weight significantly affected the IgG levels in the blood of the lambs of low-colostrum-producing sheep. In a study on goat kids of French Alpine, researchers reported that IgG levels in death goat kids were below 75.1 mg/ml and thereby showed the importance of IgG levels in blood (O'Brien and Sherman, 1993) in healthy goat kids, IgG levels was over 143 mg/ml, while these levels were below 75 mg/ml in dead goat kids.

It was observed that the effect of postpartum period on the chemical composition of goat colostrum

was significant. In 1995, in their study on goat colostrum, Hadjipanayiotou (1995) determined that dry matter value at birth was 13.20% and the fat content, protein content and ash content of the goat milk used in the study were 4.26%, 4.90%, and 0.83%, respectively. Marounek et al. (2012) carried out a study on goat colostrum and investigated the properties of goat colostrum during the 30 days after birth. Fat content of the colostrum used in this study varied between 3.48% and 5.67%. The highest fat content was determined at birth in that study. Decreases and increases in the fat content of colostrum were distinguished during the 30-day observation. Rachman et al. (2015) studied the colostrum and dry matter amounts in the milk of Peranakan Etawah, Saanen and Jawarandu goat genotypes and determined that dry matter amount in Peranakan Etawah colostrum was 38.96% at the first day and it was 37.49% in Jawarandu colostrum and 47.09% in Saanen goats colostrum. In general, dry matter amount in colostrum decreases as the time after birth passes.

In goats, colostrum production and protein ratio in its composition are proportionate to the size of the mammary, in addition to varying depending on species (Marnet and Komara, 2008). Another important issue is the tension occurring due to the secretion level of the alveolar cells in mammary glands and the resultant increase in the time spent to remove the remaining colostrum from the mammary glands. Capote et al. (1992) reported that the size of mammary at the first three hours after birth was close to the size of the mammary of Tinerfena genotype goats at the fourth week of lactation. However, in their study on cow colostrum, Ontsouka et al. (2011) reported that protein levels in the mammary glands were higher than those in colostrum. This is attributed to the high levels of IgG in total protein. The immunoglobulin levels found in our study (IgG, IgM, IgA) are in agreement with the results reported in the relevant literature. Linzell and Peaker (1974) and Arguello et al. (2006a, 2008) reported that the fat content of colostrum increased after 24 hours postpartum and attributed this to the higher levels of fat in the remaining milk as a result of the high amounts of milk removed from mammary glands. The change in the lactose content of colostrum is similar to the change in fat and protein contents. This change is in agreement with the results reported by Hadjipanayiotou (1995), Arguello et al. (2004), Moreno-Indias et al. (2012a,b) and Piccone et al. (2011b).



In conclusion, the immunoglobulin levels in the blood do not change with changing birth weight, colostrum source and first colostrum intake time, whereas, during the days following the birth, the amount of immunoglobulin intake per live weight is significant. In other words, feeding goat kids with 80 mg/ml IgG-containing colostrum or milk substitute

REFERENCES

- Ahmad R, Khan A, Javed MT, Hussain I. 2000. The level of immunoglobulins in relation to neonatal lamb mortality in Pak-Karakul sheep. *Vet Arhiv*, 70, 129-139.
- Ahmadi M, Velciov A, Riviş A, Traşcă T, Hărmănescu M, Antoanela C, Scurtu M. 2009. Physico-chemical and Nutritional Characterization of Bovine Colostrum, 16th Symposium on Analytical and Environmental Problems, 28 September SZAB Szeged, Hungary. 430- 433.
- Aldomy Hussein NO, Sawalha L, Khatatbeh K, Aldomy A. 2009. A national survey of perinatal mortality in sheep and goats in Jordan. *Pak Vet J*, 29, 102-106.
- Alves AC, Alves NG, Ascari IJ, Junqueira FB, Coutinho AS, Lima RR, Perez JRO, De Paula SO, Furusho-Garcia IF, Abreu LR. 2015. Colostrum composition of Santa Ines sheep and passive transfer of immunity to lambs, *J Dairy Sci*, 68(6): 3706-3716.
- Anonim. 1981. TS 1018 Çiğ Süt Standardı, Türk Standartları Enstitüsü, Ankara.
- Arguello A. 2000. Artificial rearing of kids. Colostrum feed, growth, carcass quality and meat quality. PhD Thesis Las Palmas de Gran Canary University, Spain.
- Arguello A, Castro N, Capote J. 2004. Growth of milk replacer kids fed under three different managements. *J Appl Anim Res*, 25, 37-40.
- Arguello A, Castro N, Alvarez S, Capote J. 2005. Effects of the number of lactations and litter size on chemical composition and physical characteristics of goat colostrum. *Small Rumin Res*, 64, 53-59.
- Arguello A, Castro N, Alvarez S, Capote J. 2006a. Effects of the number of lactations and litter size on chemical composition and physical characteristics of goat colostrum. *Small Rumin Res* 64, 53-59.
- Arguello A, Castro N, Batista M, Moreno-Indias I, Morales-DelaNuez A, Sanchez-Macias D, Quesada E, Capote J. 2008. Chitotriosidase activity in goat blood and colostrum. *J Dairy Sci*, 91, 2067-2070.
- Arguello A. 2011. Trends in goat research. *J Appl Anim Res*, 39, 429-434
- Banchero GE, Quintans G, Martin GB, Lindsay DR, Milton JT. 2004. Nutrition and colostrum production in sheep. Metabolic and hormonal responses to a high energy supplement in the final stages of pregnancy. *Reprod Fertil Dev*, 16, 633-643.
- Berry E, Broughan J. 2007. Use of the DeLaval cell counter (DCC) on goats' milk. *J Dairy Res*, 74(3): 345-348.
- Besser TE, Gay CC. 1994. The importance of colostrum to the health of the neonatal calf. *Vet. Clin North Am. Food Anim Prac*, 10, 107-117.
- Capote J, Darmanin N, Delgado JV, Fresno MR, Lo'pez JL. 1992. *Agrupación Caprina Canaria Ed. Consejería de Agricultura y Pesca del Gobierno de Canarias, Canary Islands, Spain.*
- Castro N, Capote J, Alvarez S, Arguello A. 2005. Effects of lyophilized colostrum and different colostrum feeding regimens on passive transfer of immunoglobulin G in Majorera goat kids. *J Dairy Sci*, 88, 3650-3654.
- Castro N, Capote J, Bruckmaier RM, Arguello A. 2011. Management effects on colostrumogenesis in small ruminants. *J Appl Anim Res*, 39, 85-93.
- Chen JC, Chang CJ, Peh HC, Chen SY. 1999. Serum protein levels and neonatal growth rate of Nubian goat kids in Taiwan area. *Small Rumin Res*, 32, 153-160.
- Chigerwe M, Dawes ME, Tyler JW, Middleton JR, Moore MP, Nagy DM. 2005. Evaluation of a cow-side immunoassay kit for assessing Ig G concentration in colostrum. *J Am Vet Med Assoc*, 27, 129-131.
- Christley RM, Morgan KL, Parkin TDH, French NP. 2003. Factors related to the risk of neonatal mortality, birth-weight and serum immunoglobulin concentration in lambs in the UK. *Prev Vet Med*, 57, 209-226.
- Constant SB, LeBlanc MM, Klapstein EF, Beebe DE, Leneau HM, Nunier CJ. 1994. Serum immunoglobulin G concentration in goat kids fed colostrum or a colostrum substitute. *J Am Vet Med Assoc*, 205, 1759-1762.
- Dale LG, Stephen DA, Deborah MH. 2009. Failure of passive transfer and effective colostrum management in calves. from http://www.larounds.ca/crus/laveng_1203.pdf, Accessed:16/11/2009
- Demiroren E, Shrestha JNB, Boylan WJ. 1995. Breed and environmental effects on components of ewe productivity in terms of multiple births, artificial rearing and 8-month breeding cycles. *Small Rumin Res*, 16, 239-249.
- Emsen E, Yaprak M, Bilgin OC, Emsen B, HW Ockerman. 2004. Growth performance of Awassi lambs fed calf milk replacer. *Small Rumin Res*, 53, 99-102.
- Gürbüz F, Başpınar E, Çamdeviren H, Keskin S. 2003. Tekrarlanan ölçümlü deneme düzenlerinin analizi. *Yüzüncü Yıl Üniversitesi Matbaası, Van.*
- Hadjipanayiotou M. 1995. Composition of ewe, goat and cow milk and of colostrum of ewes and goats. *Small Rumin Res*, 18, 255-262.
- Halliday R, Williams MR. 1979. Absorption of immunoglobulin from colostrum by bottle-fed lambs. *Ann Rech Vet*, 10, 549-556.
- Hernandez-Castellano LE, Almeida AM, Castro N, Arguello A. 2014a. The colostrum proteome, ruminant nutrition and immunity. *Curr Protein Pept Sci*, 15, 64-74.
- Hernandez-Castellano LE, Almeida AM, Ventosa M, Coelho AV, Castro N, Arguello A. 2014b. The effect of colostrum intake on blood plasma proteome profile in newborn lambs: Low abundance proteins. *BMC Vet Res*, 10, 85.
- Hernandez-Castellano LE, Torres A, Alavoine A, Ruiz-Diaz MD, Arguello A, Capote J, Castro N. 2011. Effect of milking frequency on milk immunoglobulin concentration (IgG, IgM and IgA) and chitotriosidase activity in Majorera goats. *Small Rumin Res*, 98, 70-72.
- Islam N, Ahmed L, Khan NI, Huque S, Begun A, Yunus ABM. 2009. Immune components (IgA, IgM, IgG immune cells) of colostrum of Bangladeshi mothers. *Pediatr Int*, 48(6): 543-548.
- Keskin M, Güler Z, Gül S, Biçer O. 2007. Changes in Gross Chemical Compositions of Ewe and Goat Colostrum during Ten Days Postpartum. *J App Anim Res*, 32(1): 25-28.



- Korhonen H. 1998. Colostrum immunoglobulins and the complement system potential ingredients of functional foods. *Bulletin Intl Dairy Fed*, 336, 36–40.
- Kurt A, Çakmakçı S, Çağlar A: Süt ve mamülleri muayene ve analiz metotları rehberi (9. Baskı). Atatürk Üniversitesi Ziraat Fakültesi Yayınları, No: 18, 254s, Erzurum, 2007.
- Langer, P. 2009. Differences in the composition of colostrum and milk in eutherians reflect differences in immunoglobulin transfer. *J Mammal*, 90, 332–339.
- Lerías JR, Hernandez Castellano LE, Suarez-Trujillo A, Castro N, Poulis A, Almeida AM. 2014. The mammary gland in small ruminants; Major morphological and functional events underlying milk production. *J Dairy Res*, 81, 304–318.
- Linzell JL, Peaker M. 1974. Changes in colostrum composition and in the permeability of the mammary epithelium at about the time of parturition in the goat. *J Physiol*, 243, 129–151.
- Marnet PG, Komara M. 2008. Management systems with extended milking intervals in ruminants: regulation, production, and quality of milk. *J Anim Sci*. 86, 47–56.
- Marounek M, Pavlata L, Misurova L, Volek Z, Dvorak R. 2012. Changes in the composition of goat colostrum during the first month of lactation, *Czech J Anim Sci*, 57(1): 28–33.
- Mastellone V, Massimini G, Pero ME, Cortese L, Piantedosi D, Lombardi P, Britti D, Avallone L. 2011. Effects of passive transfer status on growth performance in buffalo calves. *Asian Aust J Anim Sci*, 24, 952–956.
- Morales-delaNuez A, Moreno-Indias I, Sanchez-Macias D, Capote J, Juste MC, Castro N, Hernandez-Castellano LE, Arguello A. 2011. Sodium dodecyl sulphate reduces bacterial contamination in goat colostrum without negative effects on immune passive transfer in goat kids. *J Dairy Sci*, 94, 410–415.
- Morales-delaNuez A, Moreno-Indias I, Sanchez-Macias D, Hernandez-Castellano L E, Suarez-Trujillo A, Assuncao P, Arguello A, Castro N, Capote J. 2014. Effects of *Cryptocodium* spp, *Chlorella* spp and *Isochrysis galbana* addition to milk replacer on goat kids and lambs growth. *J Appl Anim Res*, 42, 213–216.
- Moreno-Indias I, Morales-delaNuez A, Hernandez-Castellano LE, Sanchez-Macias D, Capote J, Castro N, Arguello A. 2012a. Docosahexaenoic acid in the goat kid diet: Effects on immune system and meat quality. *J Anim Sci*, 90, 3729–3738.
- Moreno-Indias, I, Sanchez-Macias D, Castro N, Morales-delaNuez A, Hernandez-Castellano LE, Capote J, Arguello A. 2012b. Chemical composition and immune status dairy goat colostrum fractions during the first 10h after partum. *Small Rumin Res*, 103, 220–224.
- Morin DE, McCoy GC, Hurley WL. 1997. Effects of quality, quantity, and timing of colostrum feeding and addition of a dried colostrum supplement on immunoglobulin G₁ absorption in Holstein bull calves. *J Dairy Sci*, 80, 747–753.
- Moro I, Abo T, Crago SS, Komiyama K, Mestecky J. 1985. Natural killer cells in human colostrum. *Cell Immunol*, 93, 467–474.
- Nobrega MA, Ovcharenko I, Afzal V, Rubin EM. 2003. Scanning human gene deserts for long-range enhancers. *Science* 302, 413.
- Nowak R, Poindron P. 2006. From birth to colostrum, Early steps leading to lamb survival. *Reprod Nutr Dev*, 46, 431–446.
- O'Brien J, Sherman D. 1993. Serum immunoglobulin concentrations of newborn goat kids and subsequent kid survival through weaning. *Small Rumin Res*, 11, 71–77.
- Official Methods of Analysis of AOAC: http://www.aoac.org/aoac_prod_imis/AOAC/Publications/Official_Methods_of_Analysis/AOAC_Member/Pubs/OMA/AOA_C_Official_Methods_of_Analysis.aspx?hkey=5142c478-ab50-4856-8939-a7a491756f48, 2000, Accessed: 01.07.2016
- Ontsouka QRU, Bruckmaier J, Blum W. 2003. Fractionized milk composition during removal of colostrum and mature milk. *J Dairy Sci*, 86, 2005–2011.
- Pakkanen R, Aalto J. 1997. Growth factors and antimicrobial factors of bovine colostrum. *Intl Dairy J*, 7, 285–297.
- Piccione G, Scianò S, Messina V, Casella S, Zumbo A. 201b. Changes in serum total proteins, protein fractions and albumin-globulin ratio during neonatal period in goat kids and their mothers after parturition. *Ann Anim Sci*, 11, 251–260.
- Quigley JD, Carson AF, Polo J. 2002. Immunoglobulin derived from bovine plasma as a replacement for colostrum in newborn lambs. *Vet Ther*, 3, 262–269.
- Quigley JD, Drewry JJ. 1998. Nutrient and immunity transfer from cow to calf pre- and post-calving. *J Dairy Sci*, 8, 79–90.
- Quigley JD, French P, James RE. 2000. Effect of pH on absorption of immunoglobulin G in neonatal calves. *J Dairy Sci*, 83, 1853–1855.
- Rachman AB, Maheswari RRA, Bachroem MS. 2015. Composition and Isolation of Lactoferrin from Colostrum and Milk of Various Goat Breeds. *Procedia Food Sci*, 3, 200–210.
- Ramirez A, Quileys A, Hevia ML, Sotillo F, Ramirez MC. 1996. Influence of forced contact on the maternal filial bond in the domestic goat after different periods of postpartum separation. *Small Rumin Res*, 23, 75–81.
- Renner E. 1993. *Practical handouts to the milk*, Justus Liebig Universität, Germany, 76.
- Rodriguez AB, Landa R, Bodas R, Prieto N, Mantecon AR, Giraldez FJ. 2008. Effect of rearing system carcass and meat quality Assaf milk fed lambs: *Meat Sci*, 80, 225–230.
- Rodriguez C, Castro N, Capote J, Morales-dela Nuez A, Moreno-Indias I, Sanchez-Macias D, Arguello A. 2009. Effect of colostrum immunoglobulin concentration on immunity in Majorera goat kids. *J Dairy Sci*, 92, 1696–1701.
- Ruiz P, Sesena S, Rieiro I, Llanos PM. 2014. Effect of postpartum time and season on physic chemical characteristics of Murcia-Granada goat colostrum. *Int J Dairy Tech*, 68(1): 88–96.
- Sherman DM, Arendt TD, Gay JM, Maefsky VA. 1990. Comparing the effects of four colostrum preparations on serum IgG levels of newborn kids. *Vet Med*, 85, 908–913.
- Singh AK. 2010. Studies on immune functions and metabolic status of growing buffalo calves in response to colostrum immunoglobulins. MSc Thesis submitted to NDRI Deemed University, Karnal (Haryana), India.
- SPSS 15. 2007. *Veri Analiz Yöntemleri*. Eymen, U.E. İstatistik Merkez Yayın No:1.
- Stelwagen K, Carpenter E, Haigh B, Hodgkinson A, Wheeler TT. 2009. Immune components of bovine colostrum and milk. *J Anim Sci*, 87(Suppl.), 3–9.
- Suraya MS, Yaakub H. 2011. Effect of colostrum feeding on the serum immunoglobulin level in Saanen crossbred kids. *Mal. J Anim Sci*, 14, 25–29.
- Trujillo AJ, Castro N, Quevedo JM, Arguello A, Capote J, Guamis B. 2007. Effect of heat and high pressure treatments on microbiological quality and immunoglobulin G stability of caprine colostrum. *J Dairy Sci*, 90, 833–839.