



# Ankara Üniversitesi Veteriner Fakültesi Dergisi

# Veterinary Journal of Ankara University

Cilt/Volume 67 • Sayı/Number 2 • 2020

#### Ankara Üniversitesi Veteriner Fakültesi Dergisi

Cilt / Volume: 67 • Sayı / Number: 2 • 2020

#### Veterinary Journal of Ankara University

Üç ayda bir yayımlanır / Published three monthly

E-ISSN 1308-2817

#### Sahibi

Ankara Üniversitesi Veteriner Fakültesi Adına Prof. Dr. Belgin SARIMEHMETOĞLU

Dekan

Danışma Kurulu / Advisory Board Prof. Dr. Hikmet Altunay, Ankara University Prof. Dr. Ali Aydın, İstanbul University Prof. Dr. Wolfgang Bäumer, Berlin Freie University Prof. Dr. Gerhard Breves, Hannover Veterinary Medicine University Prof. Dr. Heiner Bollwein, Zurich University Prof. Dr. Ali Bumin, Ankara University Prof. Dr. Yavuz Cevger, Ankara University Prof. Dr. R. Teodor Cristina, Banat's University Prof. Dr. Ahmet Çakır, Ankara University Prof. Dr. Roman Dabrowski, Lublin Life Science University Prof. Dr. Cornelia Deeg, Münih Ludwig Maximilian University Prof. Dr. Bilal Dik, Selçuk University Prof. Dr. Levent Dirikolu, Louisiana University Prof. Dr. Marc Drillich, Vienna Veterinary Medicine University Prof. Dr. Bülent Ekiz, Istanbul-Cerrahpaşa University Prof. Dr. Emel Ergün, Ankara University Prof. Dr. Frank Gasthuys, Gent University Prof. Dr. Tamay Başağaç Gül, Ankara University Dr. Paweł Górka, Krakow Agriculture University Prof. Dr. Berrin Kocaoğlu Güçlü, Erciyes University Prof. Dr. Shimon Harrus, Jerusalem Hebrew University Prof. Dr. R1fk1 Haz1roğlu, Ankara University Assoc. Prof. Dr. Jia-Qiang He, Virginia Polytechnic Institute and State University Assoc. Prof. Dr. Laura Hurtado, Elvas Agrarian School Prof. Dr. Şeref İnal, Selçuk University Prof. Dr. M. Taner Karaoğlu, Ankara University Prof. Dr. Abdullah Kaya, Selçuk University Prof. Dr. Arif Kurtdede, Ankara University Prof. Dr. Mariusz P. Kowalewski, Zurich University Prof. Dr. Osman Kutsal, Ankara University Prof. Dr. Erdoğan Memili, Mississippi State University Prof. Dr. A. Serpil Nalbantoğlu, Ankara University Prof. Dr. F. Seda Bilir Ormancı, Ankara University Prof. Dr. Hakan Öztürk, Ankara University Prof. Dr. Dušan Palić, Münich Ludwig-Maximilians University Ankara Üniversitesi Veteriner Fakültesi Dekanlığı Prof. Dr. Lazo Pendovski, Skopje Ss. Cyril and Methodius University Prof. Dr. H. P. Salmann, Hannover Veterinary Medicine University Prof. Dr. Barış Sareyyüpoğlu, Ankara University Prof. Dr. Sabine Schäfer-Somi, Vienna Veterinary Medicine University Prof. Dr. Franz Schwarzenberger, Vienna Veterinary Medicine University Prof. Dr. Antti Sukura, Helsinki University Prof. Dr. Hamdi Uysal, Ankara University Prof. Dr. Ongun Uysal, Ankara University Prof. Dr. Rıfat Vural, Ankara University Prof. Dr. Sakine Yalcın, Ankara University Prof. Dr. Ender Yarsan, Ankara University Assoc. Prof. Dr. Ebru Yıldırım, Kırıkkale University

"Bu dergi Thomson Reuters®'in SCI-EXP ve JCR ile Uluslararası CABI yayınlarının CAB Abstracts, Global Health, CAB Direct, Database Subsets; Scopus ve Ulakbim (Yaşam Bilimleri) veri tabanları kapsamındadır.

This journal is covered by SCI-EXP and JCR of Thomson Reuters®, CAB Abstracts, Global Health, CAB Direct, Database Subsets; Scopus and Ulakbim (Life Sciences) database systems.

© Ankara Üniversitesi Veteriner Fakültesi Dergisi

Tüm hakları saklıdır. Bu Derginin tamamı ya da Dergide yer alan bilimsel çalışmaların bir kısmı ya da tamamı 5846 sayılı yasanın hükümlerine göre Ankara Üniversitesi Veteriner Fakültesi Dekanlığının yazılı izni olmaksızın elektronik, mekanik, fotokopi ya da herhangi bir kayıt sistemiyle çoğaltılamaz, yayımlanamaz.

> Internet Adreslerimiz / Web Addresses http://vetfakderg.ankara.edu.tr http://vetjournal.ankara.edu.tr Yayım Tarihi: 03.03.2020

#### Editörler Kurulu / Editorial Board

#### Sorumlu Yazı İşleri Müdürü

Baş Editör / Editor-in Chief Prof. Dr. E. Ebru ONBAŞILAR

Editörler / Editors Prof. Dr. Aykut ÖZKUL Prof. Dr. Bahadır GÖNENÇ Prof. Dr. Necmettin ÜNAL Prof. Dr. İ. Safa GÜRCAN Doç. Dr. Levent ALTINTAŞ Doç. Dr. Halit KANCA Doç. Dr. Mert PEKCAN Doç. Dr. Bengi ÇINAR KUL Doç. Dr. İrem ERGİN Dr. E. Çağatay ÇOLAKOĞLU Dr. Güzin İPLİKÇİOĞLU ÇİL Dr. Ali ÇALIK Dr. Ahmet CEYLAN Dr. Gökben ÖZBAKIŞ BECERİKLİSOY

Sekreterya: Havva Yenidoğan

#### Yönetim Yeri

Adres / Address

Yayın Alt Komitesi 06 110 Ankara, Türkiye Tel: 90 312 317 03 15, Fax: 90 312 316 44 72 Sürdürüm ve iletişim: vfdergi@veterinary.ankara.edu.tr URL: http://www.veterinary.ankara.edu.tr/vdergi Yayın Türü: Yaygın süreli ve hakemli

## Ankara Üniversitesi Veteriner Fakültesi Dergisi

Cilt / Volume: 67 • Sayı / Number: 2 • 2020

### Veterinary Journal of Ankara University

# İÇİNDEKİLER / CONTENTS

Investigating various performance traits of Karakul sheep Karagül koyunlarında çeşitli verim özelliklerinin araştırılması Halil Erol, Ceyhan Özbeyaz, Necmettin Ünal	113
Determination of 8-hydroxy-2'deoxyguanosine, malondialdehyde levels and antioxidant enzyme activities in Kangal dogs with venereal tumour Venereal tümörlü Kangal köpeklerinde 8-hidroksi-2'deoksiguanozin, malondialdehit düzeyleri ile antioksidan enzim	
aktivitelerinin belirlenmesi Nazlı Ercan, Murat Yüksel, Mustafa Koçkaya	121
Determination of serum amyloid A, haptoglobin and hepcidin levels in calves with endemic viral pneumonia Yaygın viral pnömonili buzağılarda serum amiloid A, haptoglobin ve hepsidin düzeylerinin belirlenmesi Ali Burak Dörtkardeş, Şima Şahinduran	127
Antioxidant activities, total phenolic and flavonoid contents of honey collected from different botanical origins Farklı botanik kaynaklardan elde edilen balların antioksidan aktiviteleri, toplam fenolikleri ve flavonoid içerikleri <b>Deniz Aker, Cevat Nisbet</b>	133
Investigation of enrofloxacin and ciprofloxacin residues by LC-MS/MS in chicken liver marketed in Afyonkarahisar Afyonkarahisar'da satışa sunulan tavuk karaciğerlerinde enrofloksasin ve siprofloksasin kalıntılarının LC-MS/MS ile araştırılması Damla Arslan Acaröz, Nalan Bayşu Sözbilir	<sup>1</sup> 137
Honey price estimation for the future in Turkey; example of 2019- 2020 Türkiye'de bal fiyatlarının geleceğe yönelik kestirimi; 2019-2020 örneği Mustafa Bahadır Çevrimli, Mehmet Saltuk Arıkan, Mustafa Agah Tekindal	143
Knowledge and attitudes in food safety and the occurrence of indicator bacteria on hands of food handlers at the point of	
pastrami sale Pastırma satış noktalarındaki gıda çalışanlarının gıda güvenliği bilgi ve tutumları ve ellerindeki indikatör bakteri varlığının belirlenmesi	
Yeliz Yıldırım, Nurhan Ertaş Onmaz, Zafer Gönülalan, Harun Hızlısoy, Serhat Al, Fulden Karadal, Erhan Kum, Aytaç Akçay	153
Three-dimensional tomographic reconstruction and morphometric analysis of skull in gazelles ( <i>Gazella subgutturosa</i> ) Ceylanlarda ( <i>Gazella subgutturosa</i> ) kafatasının üç boyutlu tomografik rekonstruksiyonu ve morfometrik analizi <b>Bestami Yılmaz, İsmail Demircioğlu, Faruk Bozkaya, Nazan Gezer İnce</b>	161
First molecular data on the validity of <i>Myxobolus ichkeulensis</i> (Cnidaria: Myxosporea) from <i>Mugil cephalus</i> (Mugilidae) in Turkish waters	
Türkiye sularında Mugil cephalus'tan (Mugilidae) Myxobolus ichkeulensis'in (Cnidaria: Myxosporea) geçerliliğine ilişkin ilk moleküler veriler	
Cenk Soner Bölükbaş, Gökmen Zafer Pekmezci, Banu Yardımcı, Melek Özpıçak, Savaş Yılmaz, Nazmi Polat	169
Influence of different water access status of suckling calves on performance, water intake behavior and some blood metabolites Mohsen Zarei, Jamal Seifdavati, Gholam Reza Ghorbani, Hosein Abdi-Benemar, Reza Seyedsharifi, Abdolhamid Karimi	175
The effects of fumaric and malic acids on the <i>in vitro</i> true digestibility of some alternative feedstuffs for ruminants Ruminantlar için fumarik ve malik asitlerin bazı alternatif yem maddelerinin <i>in vitro</i> gerçek sindirilebilirliği üzerine etkileri <b>Buğra Genç, Mustafa Salman, Bora Bölükbaş, İsmail Kaya, Mustafa Açıcı</b>	185
Insulin-like growth factor-1 gene expression in the endometrium of repeat breeder cows after the administration of presynch-10 and ovsynch protocol	
Presynch-10 ve ovsynch protokolleri uygulamasından sonra repeat breeder ineklerde endometriyal insülin benzeri büyüme faktörü-1 gen ekspresyonu	102
Ali Reha Ağaoğlu, Özgecan Korkmaz Ağaoğlu, Özge Sidekli, Mesih Kocamüftüoğlu Short Communication / Kısa Bilimsel Çalışma	193
PCR detection of Mycobacterium genavense DNA in fecal samples of caged birds Kafes kuşu dışkı örneklerinde Mycobacterium genavense DNA'sının PZR ile saptanması	
Orkun Babacan, Bülent Baş, Barış Sareyyüpoğlu	201
Case Report / Olgu Sunumu	
The first case of <i>Bovicola (Werneckiella) ocellatus</i> (Piaget, 1880) (Phthiraptera: Ischnocera: Trichodectidae) on a donkey ( <i>Equus asinus</i> Linnaeus, 1758) in Turkey Türkiye'de bir eşekte ( <i>Equus asinus</i> Linnaeus, 1758) ilk <i>Bovicola (Werneckiella) ocellatus</i> (Piaget, 1880) (Phthiraptera: Ischnocera: Trichodectidae) olgusu	
Bilal Dik, İpek Erdem, Aykut Zerek, Mustafa Karagöz, Mehmet Yaman	205
Review / Derleme	
The role of micronutrients in high-yielding dairy ruminants: Choline and vitamin E Luciano Pinotti, Michele Manoni, Francesca Fumagalli, Nicoletta Rovere, Marco Tretola, Antonella Baldi	209

Ankara Üniv Vet Fak Derg, **67**, 113-120, 2020 DOI: 10.33988/auvfd.491960

### Investigating various performance traits of Karakul sheep

Halil EROL<sup>1,a,⊠</sup>, Ceyhan ÖZBEYAZ<sup>2,b</sup>, Necmettin ÜNAL<sup>2,c</sup>

<sup>1</sup>Edremit Directorate of District Agriculture and Forestry, Balıkesir; <sup>2</sup>Ankara University, Faculty of Veterinary Medicine, Department of Animal Breeding and Husbandry, Ankara, Turkey.

<sup>a</sup>ORCID: 0000-0001-9728-7244; <sup>b</sup>ORCID: 0000-0002-3748-9992; <sup>c</sup>ORCID: 0000-0001-5250-7063.

<sup>⊠</sup>Corresponding author: erol.halil@tarimorman.gov.tr Received date: 04.12.2018- Accepted date: 20.09.2019

**Abstract:** Karakul sheep in Turkey is an endangered breed according to the total number of ewes. Two herds of Karakul sheep in Tokat Province, reared similar environmental conditions, kept *in situ in vivo* conservation as a gene resource. This study aimed to determine various performance traits of Karakul sheep reared in breeder conditions as a gene resource. Least squares means for lambing rate (LR), litter size (LS) and lamb production (LP) were found  $95.8\pm0.60\%$ ,  $1.04\pm0.01$  and  $100.0\pm0.80\%$ , respectively. It was determined that farm and age had significant effect on LR and LP, while farm and year had significant impact on LS. Survival rate in lambs were found 95.2 and 94.1%, respectively on  $90^{\text{th}}$  and  $180^{\text{th}}$  days. Means of least squares for live weights of lambs at birth and on  $90^{\text{th}}$  and  $180^{\text{th}}$  days were identified as  $3.35\pm0.02$ ,  $21.52\pm0.27$  and  $30.34\pm0.32$  kg, respectively. Live weight after shearing and greasy fleece weight values were  $40.73\pm0.12$  and  $2.04\pm0.01$  kg for females and  $62.65\pm0.53$  and  $3.48\pm0.06$  kg for males. Lactation milk yield (LMY) was  $104.85\pm3.73$  kg in ewes and lactation duration was  $159.01\pm1.70$  days. LMY was affected by the lactation number and farm while lactation duration was affected by lactation number. Results revealed that various performance traits of Karakul sheep conserved in breeder conditions were similar to or better than those previously reported for this breed. Also, the breed is similar to medium size native sheep breeds in terms of growth and mature live weight.

Keywords: Genetic resource, Karakul sheep, production traits

#### Karagül koyunlarında çeşitli verim özelliklerinin araştırılması

Özet: Türkiye'de Karagül koyunu, yok olma tehlikesi altında bulunan bir ırktır. Tokat ilinde birbirine yakın ve benzer çevresel koşullarında yetiştirilen iki Karagül koyun sürüsü, yetiştirici koşullarında gen kaynağı olarak korunmaktadır. Bu araştırma, yetiştirici koşullarında gen kaynağı olarak korunmaktadır. Bu araştırma, yetiştirici koşullarında gen kaynağı olarak korunmaktadır. Bu araştırma, yetiştirici koşullarında gen kaynağı olarak korunmaktadır. Bu araştırma, yetiştirici koşullarında gen kaynağı olarak korunan Karagül koyunlarında çeşitli verim özelliklerinin incelenmesi amacıyla yapılmıştır. Koyunlarda doğum oranı, bir doğuma kuzu sayısı ve kuzu verimi için en küçük kareler ortalamaları % 95,8±0,60; 1,04±0,01 ve % 100,0±0,80 olmuştur. İşletme ve yaşın doğum oranına ve kuzu verimine; işletme ve yılın ise bir doğuma kuzu sayısına etkileri önemli bulunmuştur. Kuzularda 90. ve 180. günde yaşama gücü sırasıyla % 95,2 ve 94,1 olarak tespit edilmiştir. Kuzularda doğum, 90. ve 180. gün canlı ağırlıkları sırasıyla 3,35±0,02; 21,52±0,27 ve 30,34±0,32 kg belirlenmiştir. Kırkım sonrası canlı ağırlık ve yapağı verimi dişilerde 40,73±0,12 ve 2,04±0,01 kg, erkeklerde 62,65±0,53 ve 3,48±0,06 kg tespit edilmiştir. Koyunlarda laktasyon süt verimi 104,85±3,73 kg ve laktasyon süresi 159,01±1,70 gün olmuştur. Laktasyon süt verimine laktasyon sırası ve işletme, laktasyon süresine ise laktasyon sırasının etkisi önemli olmuştur. Sonuç olarak yetiştirici koşullarında korunan Karagül koyunlarında genel olarak çeşitli verim özelliklerinin ırk için bildirilen değerlere benzer veya daha iyi olduğu belirlenmiştir. Ayrıca Karagül ırkı, canlı ağırlık ve büyüme bakımından orta yapılı yerli koyun ırklarına benzerlik göstermektedir.

Anahtar sözcükler: Gen kaynağı, Karagül koyunu, verim özellikleri

#### Introduction

Karakul is a fat-tailed sheep breed with coarse fleece (1). This breed is called after Karakul town in Turkmenistan (1) or Karagöl located in the city of Bukhara in Uzbekistan (2). The most significant characteristic of this breed is the Astrakhan fur obtained from the newborn lambs. It has been reported that Karakul breed was first brought to Tokat and Antalya in Turkey by the families who migrated from Caucasia at the end of 19<sup>th</sup> century (22). Later, rams and ewes were brought from Turkistan in 1929 to start breeding (2, 11). Breeding was undertaken in many state institutions led by Çifteler (Eskişehir) and Kazova (Tokat); however, subsequently breeding in these state farms was terminated. Today, Karakul sheep are bred only in Tokat vicinity at a small scale. Because total number of Karakul ewes has decreased considerably, Karakul breed in Turkey was accepted as an endangered breed (8). Therefore, two herds (a total of 320-head) were conserved in the framework of the project to conserve the genetic resources of domesticated animals. Previous studies, carried out in the 1960s, investigating Karakul sheep in Turkey were mostly related to curl forms and skin structure (6, 7, 11). Some studies performed in 2000s were published on milk (16), fleece (17), fertility (23), and some production traits (13). A number of production traits of ewes and lambs in the Karakul herds during the period of 2005-2008, in which the current research was conducted, were reported (13). The current study includes fertility, live weight after shearing, greasy fleece yield, milk yield and some udder measurements along with lamb survival rates and growth characteristics of the herds from 2011 to 2015.

Karakul sheep breed has been reared for a long time in Anatolia and it is one of components of biodiversity of Turkey. This breed faces a challenge of the need to increase production traits to provide sustainable production. Astrakhan fur, the most important yield of the breed, is not generally utilized in Turkey, and so the existence of the breed depends on the use of other yield characteristics. For this reason, it is important to know the current information about production traits of the breed.

The study aimed to investigate fertility, survival ability, growth, production of milk and wool of Karakul sheep under *in situ in vivo* conservation.

#### **Material and Methods**

The study was conducted on two Karakul sheep herds reared in Gülpınar and Ulaş villages of Tokat province (Gülpınar and Ulaş villages are situated between 40° 18' 03"- 40° 18' 48" east longitude and 36° 26' 11"- 36° 23' 07" north latitude and elevation from sea level is 630 m). Data on the number of ewes, the performance traits of which were under investigation are presented in the form of tables. Husbandry and feeding conditions were generally similar in the farms where the project was carried out. The distance between the two farms is 7 km. Ewes in both herds were fed on pasture during the first 3/5period of gestation, and were offered with 400 g of concentrate feed (14.0% crude protein and 2400 kcal ME/kg) and roughage (400 g sugar beet pulp and 500 g vetch plus barley straw) per ewe daily during the last 2/5 period of gestation. The ewes were kept indoors during the first 6 weeks of lactation period and fed similar to the last period of gestation. After indoor keeping, the ewes were fed on pasture during the rest of lactation period.

Ewes mating was conducted as random mating. Lambing rate (LR) and lamb production (LP) were calculated according to the number of ewes exposed to rams and litter size (LS) was calculated based on the number of ewes lambing. Survival rate (SR) of lambs was based on lambs born alive. The date of birth, gender and age of dam were recorded at birth along with their birth weight no later than 24 hours after the birth. The lambs were received alfalfa hay and lamb grower feed. The growth of lambs was recorded and live weight on the 90<sup>th</sup> and 180<sup>th</sup> days was calculated by using interpolation method. Ewes were shorn in June every year. Greasy fleece weight and live weight after shearing were identified with the help of electronic bascules sensitive to 50 g.

Milk yield controls were conducted in 2013 on a total of 30 ewes per herd randomly selected from both herds with single births at the second week of February and were in the 1<sup>st</sup>, 2<sup>nd</sup> and 3<sup>rd</sup> lactation. The first milk control was conducted about on the 45<sup>th</sup> day after the birth and controls were continued about 90 and 135 days after parturition. The data for the lactation milk yield (LMY) were calculated by interpolation and extrapolation methods. The lactation duration (LD) was calculated as the period between the date of birth and the end of lactation. The end of lactation was determined by extrapolation method based on the last milk control day.

The lambs were separated from their mother one day before the milk control day at 17<sup>.00</sup> and the ewes were milked by hand on the milk control day around at 08.<sup>00</sup> and 17.<sup>00</sup>. Lambs in both herds were not weaned until the last control milking and went to the pasture with the ewes. LMY was calculated by using Fleischmann's method (TrapezII). Udder measurements were collected right before the 1<sup>st</sup> and the 3<sup>rd</sup> milk control days of lactation with the help of measuring tape and digital calipers (18).

*Statistical analysis:* Fertility, greasy fleece weight, lactation milk yield, lactation duration and udder measurements in ewes and growth characteristics in lambs were examined with the Least Squares Method. Duncan's Multiple Comparison Test was used to compare more than two groups with significant differences. Survival rate in lambs was analyzed using Chi-Square Method (9).

#### Results

Table 1 presents the ewes fertility characteristics of which were assessed and Table 2 presents the least squares means for fertility characteristics. Mean least squares for LR, LS and LP in ewes were  $95.8\pm0.60\%$ ,  $1.04\pm0.01$  and  $100.0\pm0.80\%$ , respectively. It was found that farm and age (P <0.05) affected LR, farm (P <0.001) and year (P <0.01) affected LS and farm (P <0.001) and age (P <0.05) affected LP (Table 2).

Survival rates of lambs on the 90<sup>th</sup> and 180<sup>th</sup> days were found 95.2 and 94.1% (Table 3) and the impact of the examined factors were generally insignificant. Table 4 presents the means of the live weights of the lambs at different stages. Live weights at birth and the 90<sup>th</sup> and 180<sup>th</sup> days of lambs were found  $3.35\pm0.02$ ,  $21.52\pm0.27$ and  $30.34\pm0.32$  kg, respectively. The effects of the examined factors on the live weight of lambs at different periods were found significant at different levels (P <0.05; P <0.01; P <0.001).

Farm	Age	2011-2012	2012-2013	2013-2014	2014-2015	General
	2	38	26	16	35	115
	3	46	34	27	16	123
C#1	4	24	47	33	27	131
Gülpınar	5	22	21	43	32	118
	6≥	20	29	38	67	154
	Total	150	157	157	177	641
	2	35	18	27	25	105
	3	35	36	18	27	116
	4	24	31	35	18	108
Ulaș	5	22	23	30	35	110
	6≥	24	32	39	53	148
	Total	140	140	149	158	587
	2	73	44	43	60	220
	3	81	70	45	43	239
C 1	4	48	78	68	45	239
General	5	44	44	73	67	228
	6≥	44	61	77	120	302
	Total	290	297	306	335	1228

Table 1. Number of ewes investigated for the fertility traits by the farm and year subclasses

Table 2. Numerical values and the least squares means (±SEM) for fertility traits

Terrer	NIME	NI E	NCLE	NIMLE	NT DA		LSM±SE	
Items	NME	NLE	NSLE	NMLE	NLBA	LR (%)	LP (%)	LS (x100)
Farm						*	***	***
Gülpınar	641	606	592	14	620	$94.4{\pm}0.80$	96.7±1.10	102.4±0.80
Ulaş	587	571	535	36	607	$97.1 {\pm} 0.80$	$103.4{\pm}1.20$	$106.5 \pm 0.80$
Age						*	*	-
2	220	202	197	5	207	91.9±1.30 <sup>a</sup>	$94.4{\pm}1.90^{a}$	102.6±1.40
3	239	232	224	8	240	$97.3{\pm}1.30^{b}$	$100.3{\pm}1.80^{b}$	103.1±1.30
4	239	228	215	13	241	$95.5{\pm}1.30^{b}$	$100.9 \pm 1.80^{b}$	105.6±1.30
5	228	221	206	15	236	$96.8{\pm}1.30^{b}$	$103.9{\pm}1.90^{b}$	107.3±1.30
6≥	302	294	285	9	303	$97.2 \pm 1.20^{b}$	$100.6 \pm 1.70^{b}$	103.6±1.20
Year						-	-	**
2011-2012	290	275	264	11	286	95.1±1.20	99.2±1.70	104.3±1.20at
2012-2013	297	283	260	23	306	95.2±1.20	$103.1{\pm}1.70$	$108.3 \pm 1.20^{b}$
2013-2014	306	296	288	8	304	96.5±1.10	98.9±1.60	$102.4{\pm}1.20^{a}$
2014-2015	335	323	315	8	331	96.3±1.10	98.9±1.60	102.6±1.10ª
Interactions								
FxA						-	-	-
FxY						-	-	-
AxY						-	-	-
General	1228	1177	1127	50	1227	95.8±0.60	$100.0 \pm 0.80$	104.4±0.60

F: Farm, A: Age, Y: Year, - P>0.05, \* P<0.05, \*\* P<0.01, \*\*\* P<0.001

<sup>a, b</sup> Differences between the means with unlike letters in the same column are significant at P < 0.05.

LSM: Least squares means, SE: Standard error, NME: Number of mating ewes, NLE: Number of lambing ewes, NSLE: Number of single lambing ewes, NMLE: Number of multiple lambing ewes, NLBA: Number of lambs born alive, LR: Lambing rate, LP: Lamb production, LS: Litter size

Items		Number of lambs (	(n)		rate (%)
	Live birth	90 <sup>th</sup> day	180 <sup>th</sup> day	90 <sup>th</sup> day	180 <sup>th</sup> day
Farm			-	*	*
Gülpınar	620	602	596	96.9	96.0
Ulaș	607	567	560	93.4	92.3
Dam Age				-	-
2	207	194	192	93.7	92.8
3	240	230	227	95.8	94.2
4	241	233	232	96.7	96.3
5	236	228	226	96.2	95.8
6≥	303	284	279	93.7	92.1
Birth Year				-	-
2012	286	271	268	94.8	93.4
2013	306	288	283	94.1	92.5
2014	304	294	293	96.4	96.4
2015	331	316	312	95.5	94.3
Gender				-	-
Female	603	572	565	94.7	93.7
Male	624	597	591	95.7	94.6
Birth Type				-	-
Single	1127	1077	1066	95.5	94.5
Twin	100	92	90	92.0	90.0
General	1227	1169	1156	95.2	94.1
$D = 0.05 + D_{-0.05}$					

Table 3. Number of lambs born alive and survival rates at different periods

- P>0.05, \* P<0.05

Table 4. The least squares means (±SEM) for live weight at different ages (kg)

Items	n	Birth	n	90 <sup>th</sup> day	n	180 <sup>th</sup> day
Farm		**		*		**
Gülpınar	620	$3.28 \pm 0.04$	602	22.01±0.41	596	29.55±0.49
Ulaș	607	$3.41 \pm 0.03$	567	$21.03 \pm 0.30$	560	31.13±0.35
Dam Age		-		-		-
2	207	$3.22 \pm 0.07$	194	21.55±0.71	192	29.67±0.84
3	240	$3.34{\pm}0.05$	230	$21.90{\pm}0.48$	227	$30.88 \pm 0.57$
4	241	$3.42 \pm 0.04$	233	22.15±0.44	232	$30.92 \pm 0.53$
5	236	$3.34 \pm 0.04$	228	21.58±0.44	226	$30.92 \pm 0.54$
5≥	303	$3.41 \pm 0.04$	284	$20.42 \pm 0.46$	279	29.32±0.57
Birth Year		***		***		***
2012	286	3.18±0.04 <sup>a</sup>	271	$19.14 \pm 0.47^{a}$	268	$28.08 \pm 0.57^{a}$
2013	306	$3.41 \pm 0.03^{b}$	288	22.41±0.35 <sup>b</sup>	283	$30.00 \pm 0.43^{b}$
2014	304	$3.48{\pm}0.05^{b}$	294	22.78±0.53b	293	$30.88 {\pm} 0.63^{b}$
2015	331	3.32±0.05°	316	21.75±0.54°	312	32.40±0.63°
Gender		***		*		***
Female	603	$3.23 \pm 0.03$	572	21.16±0.33	565	29.53±0.39
Male	624	$3.47 \pm 0.03$	597	21.88±0.35	591	31.15±0.42
Birth Type		***		*		***
Single	1127	$3.76 \pm 0.01$	1077	22.10±0.11	1066	31.72±0.13
Гwin	100	$2.93 \pm 0.05$	92	20.94±0.53	90	28.96±0.63
Interactions						
FxDA		***		-		-
FxBY		***		***		***
FxG		-		*		***
FxBT		-		-		-
DAxBY		**		-		-
DAxG		-		-		-
DAxBT		-		-		-
BYxG		**		-		-
3YxBT		***		-		-
GxBT		-		-		-
Regression				1.603±0.291***		1.564±0.356***
General	1227	$3.35 \pm 0.02$		21.52±0.27		$30.34 \pm 0.32$

F: Farm, DA: Dam Age, BY: Birth Year, G: Gender, BT: Birth Type, - P>0.05, \* P<0.05, \*\*\* P<0.001 <sup>a, b, c</sup> Differences between the means with unlike letters in the same column are significant at P<0.05.

Regression: Partial regression of live weight on birth weight.

Table 6 presents the least squares means for LMY and LD, and Table 7 shows the least squares means for udder measurements. LMY and LD were determined to be

104.85 $\pm$ 3.73 kg and 159.01 $\pm$ 1.70 days, respectively. Udder circumference on the 45<sup>th</sup> day of lactation was found 44.20 $\pm$ 0.37 cm and teat-floor distance 26.48 $\pm$ 0.15 cm. It was identified that lactation number and farm affected LMY (P <0.01) and lactation number affected the LD (P <0.05). The udder measurement values except teat-floor distance was found to decrease when lactation progressed (Table 7).

Table 5. The least squares means	(±SEM) for live we	eight after shearing and	greasy fleece weight (kg)

Items		Live w	eight afte	er shearing		Greasy fleece we	ight	
	n	Ewes	n	Rams	n	Ewes	n	Rams
Farm		***		***		***		*
Gülpınar	692	40.23±0.13	49	$60.78 {\pm} 0.73$	685	$1.88 \pm 0.02$	49	$3.43{\pm}0.09$
Ulaș	628	41.23±0.14	60	$64.52 \pm 0.79$	628	$2.20 \pm 0.02$	60	$3.54{\pm}0.08$
Age		***		***		***		***
1.5	164	35.54±0.28ª	48	42.59±0.95ª	164	$1.91{\pm}0.03^{a}$	48	2.86±0.10 <sup>a</sup>
2.5	206	$40.50{\pm}0.23^{b}$	26	$63.29{\pm}0.99^{b}$	206	$2.03{\pm}0.03^{b}$	26	$3.46{\pm}0.11^{b}$
3.5	229	42.60±0.22°	18	69.72±1.15°	228	$2.12{\pm}0.03^{bc}$	18	3.97±0.12°
4.5	233	$42.43 \pm 0.22^{\circ}$	17	$74.99 {\pm} 1.23^{d}$	230	2.14±0.03°	17	$3.65 {\pm} 0.13^{b}$
5.5	248	42.01±0.22°			248	$2.03{\pm}0.02^{b}$		
6≥	240	$41.29 \pm 0.24^{bc}$			237	$2.00{\pm}0.03^{b}$		
Year		***		***		***		***
2012	329	39.59±0.19ª	18	57.96±1.15ª	329	$1.99{\pm}0.02^{a}$	18	3.23±0.12 <sup>a</sup>
2013	328	$40.82{\pm}0.18^{b}$	29	$62.31{\pm}1.09^{b}$	327	$2.05{\pm}0.02^{ab}$	29	3.24±0.12 <sup>a</sup>
2014	331	$40.44{\pm}0.20^{b}$	31	$63.52 \pm 1.12^{b}$	327	$1.97{\pm}0.02^{a}$	31	$3.70{\pm}0.12^{b}$
2015	332	$42.07 \pm 0.20^{\circ}$	31	$66.81 \pm 0.92^{\circ}$	330	$2.15{\pm}0.03^{b}$	31	$3.76 \pm 0.10^{b}$
Interactions								
FxA		-		***		***		***
FxY		***		***		***		-
AxY		**		-		***		-
General	1320	40.73±0.12	109	62.65±0.53	1313	2.04±0.01	109	3.48±0.06

F: Farm, A: Age, Y: Year, - P>0.05, \* P<0.05, \*\* P<0.01, \*\*\* P<0.001

<sup>a, b, c, d</sup> Differences between the means with unlike letters in the same column are significant at P<0.05.

Items	n	LMY (kg)	Min	Max	LD (day)	Min	Max
LN		**			*		
1	20	92.12±4.56 <sup>a</sup>	44	128	153.41±2.91ª	130	169
2	19	$112.09 \pm 4.68^{b}$	74	174	$158.24{\pm}2.99^{ab}$	136	187
3	20	$110.33 \pm 4.56^{b}$	85	141	165.38±2.91 <sup>b</sup>	143	202
Farm		**			-		
Gülpınar	29	97.50±3.79	44	128	156.21±2.42	130	202
Ulaş	39	112.19±3.73	74	174	161.81±2.38	136	187
Interaction							
LNxF		-			-		
General	59	104.85±3.73	44	174	159.01±1.70	130	202

LMY: Lactation milk yield, LD: Lactation duration, LN: Lactation number, F: Farm, Min: Minimum, Max: Maximum, - P>0.05, \* P<0.05, \*\* P<0.01

<sup>a, b</sup> Differences between the means with unlike letters in the same column are significant at P<0.05.

Items	n -	Days								
Items	п -	45 <sup>th</sup> day	135 <sup>th</sup> day	45 <sup>th</sup> day	135 <sup>th</sup> day	45 <sup>th</sup> day	135 <sup>th</sup> day			
		Udder	width	Udder	depth	Udder circ	umference			
LN		*	**	-	-	-	-			
2	22	13.17±0.21ª	$9.48{\pm}0.28^{a}$	$14.70 \pm 0.18$	$11.64 \pm 0.56$	42.98±0.63	28.73±0.88			
3	20	13.95±0.22 <sup>b</sup>	$10.09{\pm}0.30^{ab}$	$15.25 \pm 0.18$	$11.35 \pm 0.60$	44.78±0.65	30.34±0.95			
4	22	13.92±0.21 <sup>b</sup>	$10.74{\pm}0.27^{b}$	$14.94{\pm}0.18$	$12.37 \pm 0.56$	44.84±0.62	31.14±0.88			
Farm		**	-	-	-	***	-			
Gülpınar	31	$13.31 \pm 0.18$	9.99±0.24	14.86±0.15	11.16±048	45.74±0.53	30.50±0.75			
Ulaș	33	$14.05 \pm 0.17$	$10.22 \pm 0.22$	15.06±0.14	12.41±0.45	42.66±0.51	29.64±0.72			
Interaction										
LNxF		**	-	-	-	*	-			
General	64	13.68±0.12	10.10±0.16	14.96±0.10	11.79±0.33	44.20±0.37	30.07±0.52			
		Length of	f right teat	Length o	f left teat	Diameter o	f right teat			
LN		**	*	*	-	*	*			
2		2.93±0.12 <sup>a</sup>	$2.25{\pm}0.07^{a}$	3.00±0.13 <sup>a</sup>	$2.34 \pm 0.07$	$1.64{\pm}0.05^{a}$	$1.31 \pm 0.03$			
3		3.41±0.13 <sup>b</sup>	2.46±0.07 <sup>ab</sup>	3.38±0.13 <sup>ab</sup>	$2.48 \pm 0.07$	$1.87{\pm}0.05^{b}$	$1.43 \pm 0.04^{1}$			
4		$3.53{\pm}0.12^{b}$	$2.54{\pm}0.07^{b}$	$3.47{\pm}0.12^{b}$	$2.50 \pm 0.06$	$1.80{\pm}0.05^{b}$	$1.42{\pm}0.03^{1}$			
Farm		-	-	-	-	***	-			
Gülpınar		3.27±0.10	2.36±0.06	3.24±0.11	2.41±0.06	$1.88 \pm 0.04$	$1.40{\pm}0.03$			
Ulaș		3.31±0.10	$2.47 \pm 0.06$	3.32±0.10	$2.47 \pm 0.05$	$1.63 \pm 0.04$	$1.38 \pm 0.03$			
Interaction										
LNxF		-	*	-	-	-	**			
General		3.29±0.07	2.42±0.04	3.28±0.073	$2.44 \pm 0.04$	1.76±0.03	1.39±0.02			
		Diameter	of left teats	Distance between teats		Teat-floor	r distance			
LN		-	**	-	-	-	-			
2		$1.61 \pm 0.05$	1.27±0.03ª	16.83±0.38	13.22±0.23	26.64±0.26	29.94±0.46			
3		$1.71 \pm 0.05$	1.41±0.03 <sup>b</sup>	$17.71 \pm 0.40$	13.29±0.25	26.45±0.27	29.66±0.49			
4		$1.72{\pm}0.05$	1.42±0.03 <sup>b</sup>	17.75±0.38	13.76±0.23	26.34±0.26	28.77±0.45			
Farm		***	-	-	-	-	*			
Gülpınar		$1.80{\pm}0.04$	$1.40{\pm}0.03$	17.29±0.32	13.27±0.20	26.53±0.22	30.04±0.39			
Ulaş		$1.56 \pm 0.04$	$1.34{\pm}0.02$	17.56±0.31	13.58±0.19	26.43±0.21	28.87±0.37			
Interaction										
LNxF		-	*	-	-	-	-			
General		1.68±0.03	1.37±0.02	17.43±0.22	13.42±0.14	26.48±0.15	29.46±0.27			

Table 7. The least squares means (±SEM) for some udder measurements (cm)

LN: Lactation number, F: Farm, - P>0.05, \* P<0.05, \*\* P<0.01, \*\*\* P<0.001

<sup>a, b</sup> Differences between the means with unlike letters in the same column are significant at P<0.05.

#### **Discussion and Conclusion**

Lambing rate (LR) (95.8%) obtained in this study is a rather high value for native sheep breeds. As a matter of fact, LR values were found higher than the values reported for the same breed (85.0 - 91.4%) (13, 15, 23) and some of the other fat tailed breeds (Akkaraman, Morkaraman) (67.6 - 90.5%) (3, 12). The high lambing rate in both farms indicates that environmental conditions were taken into consideration during the mating period. Fertility characteristics were considered to be the lowest in two years old ewes. Litter size (LS) obtained from Karakul ewes (1.04) was identified similar to the values reported for the same breed (1.00 - 1.18) (11, 13, 15, 23). The Karakul breed has of low value in terms of LS and is similar to the Dağlıç (1.05) (10) and Karayaka (1.03 -1.08) (5, 20) in this respect. The low LS in Karakul herds is due to the low rate of twin birth; this shows that although the breed has a high LR, it is not a prolific breed.

There is only one study in the literature on the survival rate (SR) of Karakul lambs in Turkey (13). In the present study, SR values at 90<sup>th</sup> and 180<sup>th</sup> days (95.2 and 94.1%) were found similar to those identified for the same herds from 2006 to 2008, reported as 96.0 and 90.0%, in general (13). SR values obtained from lambs on the 90<sup>th</sup> and 180<sup>th</sup> days can be regarded as optimal and this is important for the sustainability of the herds.

The findings of the birth weight for female lambs (3.23 kg) and male lambs (3.47 kg) and the average live weight on the 180<sup>th</sup> days (30.34 kg) in this study were similar (3.24 and 3.47 kg) (11) or higher (3.03 and 3.23 kg) (13) than the those of values for birth weight of female and male lambs and live weight on the 180<sup>th</sup> day (24.62 kg) (13) of the same breed. When the live weights at birth, 90<sup>th</sup> and 180<sup>th</sup> days of the breed are evaluated together, it could be said that Karakul breed is similar to medium size breeds (Bafra, Dağlıç, Karayaka), but lower than the large size breeds (Akkaraman, Chios) in terms of the growth (1,

The means obtained for the ewes live weight after shearing (40.73 kg) and greasy fleece weight (2.04 kg) were consistent with the those of results reported for the same breed (36.81 - 42.95 kg and 1.84 - 2.84 kg) (11, 13). In addition, live weight after shearing value was similar to the lower limit of range (42.70 - 62.60 kg) reported for some native breeds (5, 19).

4, 5, 12, 19, 21).

LMY (104.85 kg) obtained in the current study was found higher than the means reported for the same breed (61.5 and 60.0 kg) (16, 22). The herd in Ulaş village is regularly milked each year; however, the herd in Gülpınar village was milked for the first time in the framework of the study. The fact that Ulaş herd gave 14.69 kg more milk than Gülpınar herd might be due to accustomed to hand milking. LMY and LD differed from the lactation number groups, while the first lactation ewes had numerically the lowest values than those of the other groups. This is in the line with the general understanding that milk production of ewes generally increases by lactation numbers. In this research, high lactation milk yield of the ewes shows that breeders can utility from Karakul breed in terms of milk yield. LD determined in this study (159.01 days) was found in the range of values reported for the same breed (137.9 - 168.0 days) (2, 16, 22). On the other hand, LD was similar in both herds. There was a wide variation in LMY and LD in both herds. This shows that it is possible to make improvement in the direction of milk yield in Karakul breed.

The longer the lactation duration was in Karakul sheep, the more decreasing was observed in udder measurement values other than the teat-floor distance. This is related to decreased milk yield due to progression of lactation. As a matter of fact, similar situation has been reported for various native breeds (3, 18). In general, udder measurement values obtained for Karakul ewes in this study were higher than those found for Tushin and Morkaraman ewes (14). Udder measurement values obtained for Bafra sheep (18) on the 42<sup>nd</sup> day of lactation were similar to or higher than the values found in the current study other than the values for right and left teat length and the distance between teats.

In conclusion, the performance traits of Karakul sheep were similar to or better than the those of results reported before for the same breed, and the breed was alike to medium size native breeds in terms of growth and mature live weight.

#### Acknowledgements

The authors would like to thank General Directorate of Agricultural Research and Policies, Ministry of Agriculture and Forestry, for their contribution to the implementation of the project.

#### **Financial Support**

This research received no grant from any funding agency/sector.

#### **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### References

- Akçapınar H (2000): Koyun yetiştiriciliği. ISBN: 975-96978-1-5, Ankara
- Aköz K (1961): Karagül koyunu ve Karagül yetiştiriciliği. Lalahan Zootekni ve Araştırma Enstitüsü, Ankara.
- Akçapınar H, Kadak R, Odabaşıoğlu F (1982): Morkaraman ve Kangal-Akkaraman koyunlarının döl verimi ve süt verimi üzerinde karşılaştırmalı araştırmalar. Ankara Univ Vet Fak Derg, 3-4, 379-391.
- 4. Akçapınar H, Özbeyaz C, Ünal N, et al (2000): The possibilities of developing dam and sire lines using Akkaraman, Sakız and Kıvırcık sheep breeds for lamb production I. Fertility in Akkaraman sheep, survival rate and growth characteristics of Sakız x Akkaraman F<sub>1</sub> and Kıvırcık x Akkaraman F<sub>1</sub> lambs. Turk J Vet Anim Sci, 24, 71-79.
- 5. Akçapınar H, Ünal N, Atasoy F, et al (2002): Adaptation capability of Karayaka and Bafra (Chios x Karayaka B<sub>1</sub>) genotypes reared in Lalahan Livestock Research conditions. Lalahan Hay Araşt Enst Derg, **1**, 11-24.
- Aköz K, Öznacar K (1960): A comparative study on the forms of curly, brightness of the curls and birth weigths of purebred Afgan Karakul lambs' raised at Lalahan Animal Breeding Station and purebred Karakul lambs' raised at Çifteler Hara. Lalahan Zoot Araş Enst Derg, 7, 65-78.
- Aköz K (1960): The relationship between finest of the Karakul ewes wool fibers and curle conformation of the new born Karakul lambs. Lalahan Zoot Araşt Enst Derg, 7, 21-27.
- Anonymous (2015): Animal genetic resources for food and agriculture, The second report on the state of the world's. FAO Commission on Genetic Resources for Food and Agriculture Assessments, Rome.
- Anonymous (2008): SPSS Software, Statistical Package for the Social Sciences for Windows. Statistical Innovations Inc (Version 14.01, No: 9869264), USA.
- **10. Başpınar H** (1985): A comparative study on the fertility, milk yield and fleece yield performances of main sheep

breeds in Turkey, kept under semi-intensive conditions. İstanbul Üniv Vet Fak Derg, **2**, 43-66.

- **11.** Batu S, Özcan H (1966): The most important morphological characters and yields of the Karakül sheep and the breeding system at Çifteler Stud Farm. Lalahan Zoot Araşt Enst Derg, **1-2**, 135-139.
- **12.** Çolakoğlu N, Özbeyaz C (1999): Comparison of some production traits in Malya and Akkaraman Sheep. Turk J Vet Anim Sci, **23**, 351-360.
- **13. Erol H, Akçadağ Hİ** (2009): Some production characteristics of Karagül sheep in situ conditions. Lalahan Hay Araşt Enst Derg, **46**, 91-104.
- 14. Kırmızıbayrak T, Aksoy AR, Saatci M, et al (2005): *Milk* yield and udder charecteristics in Tuj and Morkaraman ewes and the relationships between them. Kafkas Üniv Vet Fak Derg, 1, 11-15.
- **15.** Köseoğlu H (1978): Studies on the improvement of twinning rate with the use of hormones in Karakul sheep. Lalahan Zoot Araşt Enst Derg, **3-4**, 64-67.
- 16. Küçük M, Öztürk Y, Bayram D (2000): Comparison of milk yield characteristics on Hamdani, Karagül and Morkaraman breeds in semi-intensive conditions. Y.Y.Ü. Vet Fak Derg, 1, 44-48.

- Küçük M, Yılmaz O, Ateş CY (2000): The evaluation of Morkaraman, Hamdani and Karakul wool for carpet wool type. YYÜ Vet Fak Derg, 2, 54-59.
- **18.** Ünal N, Akçapınar H, Atasoy F, et al (2008): Some udder traits and growth of lambs and phenotypic correlations between those of traits with milking traits and milk production measured by various milk estimation methods in Bafra sheep. Ankara Univ Vet Fak Derg, **55**, 117-124.
- Ünal N, Akçapınar H, Atasoy H, et al (2004): The body weight and fleece traits of White Karaman, Chios x White Karaman F1, B1, Kıvırcık x White Karaman F1, B1, Karayaka and Bafra sheep. Lalahan Hay Araşt Enst Derg, 2, 15-22.
- 20. Ünal N, Atasoy F, Akçapınar H, et al (2003): Fertility traits, survival rate and growth characteristics of Karayaka and Bafra (Chios x Karayaka B<sub>1</sub>) genotypes. Turk J Vet Anim Sci, 27, 265-272.
- Unal N (2002): Survival rate, growth characteristics and some body measurements of Akkaraman and Sakiz (Chios) x Akkaraman F<sub>1</sub> lambs. Turk J Vet Anim Sci, 26, 109-116.
- **22.** Yalçın BC (1986): Sheep and goats in Turkey. FAO Animal Production and Health Papaer, Number **60**, Rome.
- 23. Yılmaz O, Odabaşıoğlu F (2006): Hamdani, Morkaraman ve Karagül koyunlarında kuzulatma sıklığının artırılması olanakları. YYÜ Sağlık Bil Derg, 1, 16-126.

Ankara Üniv Vet Fak Derg, **67**, 121-125, 2020 DOI: 10.33988/auvfd.492765

### Determination of 8-hydroxy-2'deoxyguanosine, malondialdehyde levels and antioxidant enzyme activities in Kangal dogs with venereal tumour

#### Nazlı ERCAN<sup>1,a, ⊠</sup>, Murat YÜKSEL<sup>2,b</sup>, Mustafa KOÇKAYA<sup>3,c</sup>

<sup>1</sup>Sivas Cumhuriyet University, Faculty of Veterinary Medicine, Department of Biochemistry, Sivas; <sup>2</sup>Hatay Mustafa Kemal University, Faculty of Veterinary Medicine, Department of Obstetrics and Gynecology, Hatay, <sup>3</sup>Sivas Cumhuriyet University, Faculty of Veterinary Medicine, Department of Physiology, Sivas, Turkey.

<sup>a</sup>ORCID: 0000-0003-3542-3743; <sup>b</sup>ORCID: 0000-0001-6634-0559; <sup>c</sup>ORCID: 0000-0001-5173-0853.

Corresponding author: nazliercan@yahoo.com
Received date: 05.12.2018- Accepted date: 12.11.2019

**Abstract:** Canine transmissible venereal tumour (TVT) is a contagious cancer and sexually transmitted one dog to another by allogenic transfer of living cancer cells. The aim of this study is to examine the relationship between antioxidative metabolism and venereal tumour at Kangal Dogs and to show the size of oxidative damage it causes through 8-hydroxy-2'deoxyguanosine (8-OHdG). The materials of this study included the blood sera of 15 Kangal Dogs with TVT, and as a control group of 15 healthy Kangal Dogs. The sera levels of 8-OHdG were determined by competitive enzyme linked immunosorbent assay (ELISA) method. The level of malondialdehyde (MDA), superoxide dismutase (SOD), glutathione peroxidase (GPx) and catalase (CAT) which are antioxidative enzymes were investigated by spectrophotometric methods as described. Serum 8-OHdG and MDA levels of dogs with TVT were found to be significantly increased compared to the control group (P<0.001). The levels of GPx were statistically different (P<0.05). In the light of the findings, which are obtained in this study, the transmissible venereal tumour, which is an infectious and treatable type of cancer, can lead to new studies that may be caused by oxidant-antioxidant metabolism.

Keywords: Antioxidant enzyme, Kangal dog, transmissible venereal tumour, 8-OHdG.

# Venereal tümörlü Kangal köpeklerinde 8-hidroksi-2'deoksiguanozin, malondialdehit düzeyleri ile antioksidan enzim aktivitelerinin belirlenmesi

Özet: Transmissible (bulaşıcı) venereal tümör (TVT), canlı kanser hücrelerinin allojenik transferiyle bir köpekten diğer bir köpeğe çiftleşme ile bulaşan bir kanser türüdür. Bu çalışmada, Kangal köpeklerinde önemli bir bulaşıcı kanser türü olan venereal tümör ile antioksidatif metabolizmanın arasındaki ilişkinin incelenmesi ve oksidatif hasarın boyutunun 8-hidroksi-2'deoksiguanozin (8-OHdG) düzeyleri üzerinden gösterilmesi amaçlandı. Bu amaçla, 15 adet TVT'li, kontrol grubu olarak 15 adet sağlıklı Kangal köpeklerinden alınan toplam 30 adet kan serumu çalışmanın materyalini oluşturdu. Bu hayvanlara ait kan serumlarında 8-OHdG kompetitif enzim bağımlı immunosorbent analiz (ELISA) yöntemi ile belirlendi. Malondialdehit (MDA), antioksidan enzimler olan süperoksit dismutaz (SOD), glutatyon peroksidaz (GPx) ve katalaz (CAT) seviyeleri ise spektrofotometrik yöntemlerle analiz edildi. TVT teşhisli köpeklerin serum 8-OHdG ve MDA düzeyleri kontrol grubundaki köpeklere göre istatiksel olarak anlamlı derecede artış gösterdiği tespit edildi (P<0,001). Antioksidan enzim olan GPx düzeylerinde istatiksel farklılık tespit edilmiştir (P<0,05). Bu çalışmada elde edilen sonuçlar neticesinde bulaşıcı ve tedavi edilebilir bir kanser türü olma niteliğindeki köpek bulaşıcı venereal tümörün oksidan antioksidan metabolizma kaynaklı yapılacak yeni çalışmalara yön gösterebilecek niteliktedir.

Anahtar sözcükler: Antioksidan enzim, bulaşıcı venereal tümör, Kangal köpeği, 8-OHdG.

#### Introduction

Transmissible venereal tumour (TVT) is a contagious tumour that affects female and male dogs by mating. The tumour is transmitted effectively by contact of mucosa. The surface membranes enable to the transmission because of disruption of mucosal integrity, abrasions or ruptures. In addition to genital contact,

extragenital TVT cases can also be seen such as skin, face, nasal cavity, oral cavity, rectum and perineum are transmitted by social behavior (eg, sniffing and licking). The metastasis rate is 5% and usually occurs in regional lymph nodes (3, 18, 27).

The fact that contagious venereal tumour is a cancer type which can be transmitted easily from one dog to another by the allogenic transfer of live cancer cells that increase the importance of this disease (22). While deaths can usually be seen in cancer patients, it should not be ignored that in many of the TVT cases can gain of lifetime immunity and this may contribute to cancer studies by revealing the mechanism (29).

The increase in the cancer incidence can be due to the increased DNA damage or reduced repair mechanism. The oxidative damage occurs because of altered DNA's. The DNA damage products are resulting in endogenously formed by oxygen radicals. These products are known to cause degenerative diseases such as aging, cancer and heart disease (7). Many studies that aim to research the markers for early diagnosis of a disease with poor prognosis such as cancer have focused on free radicals and oxidative stress especially in recent years (8). Endogenic formation of reactive oxygen species (ROS) leads to oxidation of DNA. Beside the factors that cause cancer, the cancer formation mechanism includes the DNA damage formed by ROS (17). 8-hydroxy-2'deoxyguanosine (8-OHdG) which is one of the DNA damage products of reactive oxygen species, is a commonly used marker in determining the extent of the DNA damage (31). Although there are over more than 20 oxidative damage products, due to its potential for susceptibility and mutagenicity are focused on most and have been the subject of many studies that 8hydroxyguanine or its deoxynucleotide 8-OHdG (6, 14, 16, 20, 28, 29).

Oxygen radicals, usually products of metabolism *in vivo* are also formed by phagocyte cells and lipid peroxidation (7). The measurement markers of lipid peroxidation are pentane, malondialdehyde (MDA), lipid hydroperoxides and isoprost (26). MDA which is a lipid peroxidation product is known as an important and most commonly used marker in evaluating oxidative stress (25). The free radicals that form the oxidative damage are defused by catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPx) and nonenzymatic antioxidants which are enzymes of defense system (26).

TVT is an important cancer type of cancer among cancer types because of a kind of cancer that is transmitted by mating of dogs from one to another by means of allogenic transfer of living cancer cells and also it shows fast endemic spread and resulting in restraints dogs for breeding. At the same time, it can that by enlightening the mechanism of a cancer with this etiology can contribute to studies conveyed on other types of cancer. Therefore, this study aims to examine the relationship between venereal tumour cases that are clinically significant and known as contagious cancer and antioxidative metabolism on Kangal Dogs peculiar to Sivas province which are grown in public farms and raised by people and to show the extent of oxidative damage via 8-OHdG.

#### **Material and Methods**

This study was approved by the Sivas Cumhuriyet University Animal Research Ethics Committee dated 23.02.2016 and numbered 23. The materials of this study included the blood sera of 15 dogs with TVT which are cytological (Figure 1) and clinically examined, and of 15 healthy dogs, all in Kangal breeding farm and people in Sivas.



Figure 1. TVT cell via cytological. Giemsa staining technique microscopy magnification 400x, scale bar: 20µm.

Blood for biochemical analysis was taken from each dog by *vena cephalica antebrachii* into biochemistry tubes, brought to the Sivas Cumhuriyet University Faculty of Veterinary Medicine Department of Biochemistry lab and centrifuged at 4,000 rpm for 10 min. Sera were stored at -20 °C until the day of analysis.

The serum levels of 8-OHdG were determined by competitive ELISA (enzyme linked immunosorbent assay) method. Analyzes were performed with the ELISA device (Thermo Scientific, Multiskan Go, USA), according to the kit procedures using SUNRED Canine 8-OHdG (Cat. No. DZE201151217) assay.

According to test procedure, added to microplate wells from 50 µl each standard (4, 8, 16, 32, 64 ng/ml) and 50 µl streptavidin-HRP included in the kit. And 40 µl of each sample was transferred to micro wells and 10 µl of 8-OHdG antibody and 50 µl of streptavidin-HRP were added. The microplate was covered and incubated for 1 hour at 37°C in a dark environment. At the end of the incubation, the wells were washed 3 times with wash buffer. After washing, were added 50 µl of chromogen solution A and 50 µl of chromogen solution B to each well and incubated for 10 min. At the end of the incubation, 50 µl of stop solution was added to each well to change the blue color to yellow. Finally, the absorbance of the standard and samples was measured on an ELISA device at a wavelength of 450 nm. Calibration curve was calculated for the absorbance values obtained and the 8OHdG levels were calculated as ng/ml against the absorbance of the samples.

MDA measurement was performed according to the Janero (11) method. The principle of this method is based on measuring the absorbance of the color produced by MDA with thiobarbituric acid (TBA) in acidic medium. For this purpose, 50  $\mu$ l of sample was taken, 250  $\mu$ l of TCA and 100  $\mu$ l of TBA solutions were added and incubated at 95°C for 30 minutes. After cooling the samples, the absorbance at 535 nm was measured spectrophotometrically in a microtiter plate reader (Thermo Scientific, Multiskan Go, USA). Standards were prepared using 1,1,3,3, tetramethoxypropane, formed by hydrolysis of this compound. The standard graph was drawn using these results and MDA results were calculated with this graph.

Determination of SOD activity according to Sun et al. (23) and Durak et al. (5) which the superoxide produced by the xanthine / xanthine oxidase system is based on reduction of nitro blue tetrazolium (NBT). The resulting superoxide radicals ( $O_2$ ) reduce NBT and form a colored formazan. This complex gives maximum absorbance at 560 nm. When SOD is present in the medium, there is no NBT reduction and does not turn blue-violet and a light color is formed depending on the amount and activity of the enzyme. Determined in a microtiter plate reader (Thermo Scientific, Multiskan Go, USA).

Determination of GPx activity according to Paglia and Valentine (19) was determined by kinetic, spectrophotometric analysis in which the incubation mixture containing phosphate buffer solution (PBS), nicotinamide adenine dinucleotide phosphate (NADPH), glutathione disulfide (GSSG), reduced glutathione (GSH), Na-azide and ethylenediaminetetraacetic acid (EDTA) was pre-incubated for 10 minutes at 37°C. The addition of hydrogen peroxidase and the decrease in absorbance at 340 nm were determined kinetically in a microtiter plate reader (Thermo Scientific, Multiskan Go, USA).

Catalase activity was calculated using the method of Aebi (1). 50 mM phosphate buffer was prepared with pH 7.0; further, 30 mM hydrogen peroxide solution was prepared. 1 mL of  $H_2O_2$  was added to a 2 mL sample, 1 mL of phosphate buffer was added to the resulting solution

and measurements were made at 230 nm in a microtiter plate reader (Thermo Scientific, Multiskan Go, USA).

The data were compared in SPSS 22.00 statistical analysis program (21). The normality of data distributions was investigated by Kolmogorov Smirnov test. Student *t*-test was used for parametric variables for the comparison of the studied parameters whether there is a statistical difference or not. Differences were considered with significant at P <0.05.

#### Results

The serum levels of 8-OHdG, MDA, SOD, GPx and CAT the dogs with TVT and control group shown in Table 1. According to Table 1, it was found that serum levels of 8-OHdG dogs ( $55.46\pm5.02$  ng/ml) with TVT diagnosis were significantly increased compared to the control group ( $8.78\pm0.70$  ng/ml) dogs (P <0.001).

The levels of MDA were found to be statistically significant increase in dogs  $(3.32\pm0.29 \text{ nmol/ml})$  with TVT diagnosis compared to control dogs  $(1.12\pm0.14 \text{ nmol/ml})$  as shown in Table 1 (P <0.001).

The levels of GPx were statistically significant between TVT ( $0.565\pm0.122$  U/ml) and control groups ( $1.512\pm0.362$  U/ml) (P <0.05). The levels SOD did not show any statistical difference between TVT ( $66.2\pm2.31$  U/L) and control groups ( $71.09\pm1.0$  U/L) (P >0.05). The levels CAT did not show any statistical difference between TVT ( $43.43\pm4.13$ U/ml) and control groups ( $48.93\pm8.79$ U/ml) (P >0.05).

#### **Discussion and Conclusion**

Antioxidant defense system is in a condition of equilibrium with free radicals formed in various ways in the organism. However, in cases where free radicals cannot be cleaned adequately from the organism it damages to the DNA and damage products are formed. These damage products can lead to various metabolic diseases, such as aging, cancer and even modifications in DNA structure. The most well-known damage product is 8-OHdG which is mutagenic (9).

**Table 1.** Serum 8-hydroxy-2'-deoxyguanosine (8-OHdG), malondialdeyde (MDA), superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPx) levels of transmissible venereal tumour (TVT) and control groups.

Parameters	<b>TVT group</b> (Mean±SE)	Control group (Mean±SE)	Р
8-OHdG (ng/ml)	55.46±5.02	$8.78{\pm}0.70$	0.001
MDA (nmol/ml)	3.32±0.29	$1.12 \pm 0.14$	0.001
SOD (U/L)	66.2±2.31	$71.09{\pm}1.0$	0.065
CAT (U/ml)	43.43±4.13	48.93±8.79	0.576
GPx (U/ml)	0.565±0.122	1.512±0.362	0.020

Data are presented as (Mean $\pm$ SE). SE: Standard error. Statistical comparisons were performed using Student *t*-test. Differences were considered statistically significant at P < 0.05.

The increase of reactive species can raise the development of malignancy and can be responsible for the rise in growth of cancer risk. The ineffectiveness of various antioxidant defense system enzymes can further increase the levels of oxidative damage and increase the cancer development in animals. Particular attention should be paid on DNA oxidative damage formed by a reactive kind such as hydroxyl radical (9). In many studies in the field of humanities and veterinary medicine increased oxidative damage in cancer patients has been associated with increased cancer risk (6, 10, 14, 16, 20, 28, 29, 30).

In a study which determines the malondialdehyde, antioxidant activity and adenosine levels in serum of dogs with TVT, serum MDA levels were found significantly higher in dogs with TVT than in healthy dogs but antioxidant activities were lower. MDA levels were found as 2.347nmol/mL in dogs with TVT and 1.512nmol/mL in healthy dogs (2). In this study, MDA levels at TVT diagnosed dogs were found to be 3.32nmol/mL and 1.12nmol/mL at healthy dogs. The results of Aydın et al. (2) are similar to the results of this study with increased MDA levels in TVT patients.

It is claimed that oxidant-antioxidant balance is an important factor in cancer formation and progression. An 8-OHdG level that is product of DNA oxidation and SOD, GPx and glutathione S transferase (GST) activities were measured as antioxidant activity in patients with bladder cancer. By means of serum 8-OHdG level and GPx activities, there were not any significant differences between the patient and control group. However, SOD activity were significantly lower and GST activity was significantly higher in the patient group (4).

In a study conducted by Macotpet et al. (15), malondialdehyde levels were evaluated in blood samples from dogs having cancer (n=80) and clinically healthy dogs (n=101). Serum MDA was significantly higher in dogs with cancer than in clinically healthy dogs.

In another study, two control and patient groups were formed according to healthy female dogs (n=13) and female dogs (n=16) diagnosed with pyoderma and healthy male dogs (n=14) and male dogs (n=12) diagnosed with. MDA were measured in plasma and 8-hydroxy-2'deoxyguanosine levels were measured in urine. MDA and 8-OHdG levels were found to be statistically significant in male and female dogs diagnosed with pyoderma (6).

The results of high MDA and 8-OHdG levels found in TVT diagnosed dogs obtained from this study are similar to the results of Macotpet et al. (15) and Ercan and Fidanci (6). It leads to the conclusion that oxidative stress can be associated with cancer types in dogs.

Antioxidant levels were researched at multicentric lymphoma, oral fibrosarcoma, mast cell tumours,

malignant melanoma, appendicular ostosarcoma, nasal tumours and peripheral ameloblastoma at dogs with cancer. Total antioxidant capacity (TAC), antioxidant enzymes which are GPx, CAT and SOD levels were measured. As a result, except for patients with nasal tumour, increased levels of SOD in patients with malignant melanoma, mast cell tumour, multicentric lymphoma and oral fibrosarcoma, increased levels of GPx and total antioxidant capacity in osteosarcoma and multicentric lymphoma patients, increased in levels of CAT and SOD at 42 dogs with cancer (24). In this study, it was observed that there was no increase at the levels of SOD, CAT and GPx in dogs with TVT but the levels were lower compared to the control group. The decrease, which is not reflected to the statistical difference, composes the result of increasing oxidative damage.

In a study in which lipid peroxidation and antioxidant enzyme levels were compared between dogs diagnosed with breast tumour and healthy dogs. The levels of lipid peroxidation have shown a significant increase in dogs diagnosed with breast tumour. It was determined that glutathione reductase levels showed significant increase in breast tumour patients when the antioxidant levels are evaluated (12).

Kumaraguruparan et al., (13) have observed in their study about breast tumour diagnosed dogs that increased antioxidant levels have an effect of reducing the peroxidation levels. SOD, CAT and GPx levels in dogs with tumours were found to be doubled compared to the normal tissues.

This study, in means of antioxidant enzyme levels, does not resemble to the results of studies of Jayasri et al., (12), Kumaraguruparan et al., (13). This study indicates that increased oxidative damage in TVT patients leads to a decrease in antioxidant enzyme.

In conclusion, the levels of 8-OHdG and MDA were found high in dogs with transmissible venereal tumours. Although a statistical difference was detected only at GPx levels, the significant decrease in antioxidant enzymes of SOD, GPx and CAT levels in dogs with TVT was remarkable. Increased oxidative damage and lipid peroxidation levels affect the changes in formation of antioxidant enzyme levels. In the light of the findings, which are obtained in this study, the transmissible venereal tumour, which is a contagious and treatable type of cancer, can lead to new studies that may be caused by oxidant-antioxidant metabolism. It can be concluded that the use of additional oxidative stress markers and antioxidant parameters may be useful in order to reveal the unknown activation mechanisms with more patients and more studies although activation of oxidant-antioxidant defense is observed in dogs with TVT.

#### Acknowledgements

The present study was presented in Mediterranean Veterinary Congress coupled with 7<sup>th</sup> REEV-Med General Assembly, 13-14 December, 2018, Kırıkkale, Turkey.

#### **Financial Support**

This study was supported by the Sivas Cumhuriyet University Scientific Research Project Coordination Unit under the project number 'V-042'.

#### **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### References

- **1.** Aebi H (1984): *Catalase in vitro*. Methods Enzymol, **105**, 121-6.
- 2. Aydın İ, Bülbül A, Avcı GE, et al (2009): Serum oxidative status and adenosine deaminase activity in dogs with transmissible venereal tumour. Bull Vet Inst Pulawy, 53, 771-774.
- **3.** Baştan A, Baki Acar D, Cengiz M (2008): Uterine and ovarian metastasis of transmissible venereal tumour in a bitch. Turk J Vet Anim Sci, **32**, 65-66.
- 4. Dincer Y, Akçay T, Kural AR, et al (2011): Evaluation of 8-hydroxy-2'-deoxyguanosine concentration and antioxidant enzyme activities in bladder cancer patients. Turkiye Klinikleri J Med Sci, 31, 553-558.
- 5. Durak I, Yurtaslani Z, Canbolat O, et al (1993): A methodological approach to superoxide dismutase (SOD) activity assay based on inhibition of nitroblue tetrazolium (NBT) reduction. Clin Chim Acta, 214,103-104.
- 6. Ercan N, Fidancı UR (2012): Piyodermalı köpeklerde 8hidroksi-2'-deoksiguanozin (8-OHdG) düzeyleri. Ankara Üniv Vet Fak Derg, **59**, 163-168.
- Fraga C, Shigenaga M, Park J, et al (1990): Oxidative damage to DNA during aging: 8-Hydroxy-2'deoxyguanosine in rat organ DNA urine. Proc Natl Acad Sci, 87, 4533-4537.
- 8. Gencer M, Ceyla E, Aksoy A, et al (2005): Oksidatif stres benign ve malign akciğer hastalıklarının ayırıcı tanısında belirteç olabilir mi? Akciğer Arşivi, 6, 89-92.
- **9.** Haliwell B (2007): Oxidative stress and cancer: have we moved forward? Biochem J, **401**, 1-11.
- **10.** Hattori Y, Nishigori C, Tanaka T, et al (1996): 8hydroxy-2'-deoxyguanosine is increased in epidermal cells of hairless mice after chronic ultraviolet B exposure. J Invest Dermatol, **107**, 733-737.
- 11. Janero DR (1990): Malondialdehyde and thiobarbituric acid-reactivy as diagnostic indices of lipid peroxidation and peroxidative tissue injury. Free Radic Biol Med, 9, 515-540.
- 12. Jayasri K, Padmaja K, Saibaba M (2016): Altered oxidative stress and carbohydrate metabolism in canine mammary tumors. Veterinary world, 9, 1489.
- **13.** Kumaraguruparan R, Balachandran C, Manohar BM, et al (2005): *Altered oxidant-antioxidant profile in canine mammary tumours*, Vet Res Commun, **29**, 287-96.

- 14. Lin H, Jenner A, Ong C, et al (2004): A high-throughput and sensitive methodology for the quantification of urinary 8-hydroxy-2'-deoxyguanosine: measurement with gas chromatography-mass spectrometry after single solidphase extraction. Biochem J, **380**, 541-548.
- **15.** Macotpet A, Suksawat F, Sukon P, et al (2013): Oxidative stress in cancer-bearing dogs assessed by measuring serum malondialdehyde. BMC Vet Res, *9*, 101.
- **16.** Martinis B, Bianchi M (2002): *Methodology for urinary* 8*hydroxy-2'-deoxyguanosine analysis by HPLC with electrochemical detection.* Pharmacol Res, **46**, 129-131.
- Matsufuji H, Ochi H, Shibamoto T (2006): Formation and inhibition of genotoxic malonaldehyde from DNA oxidation controlled with EDTA. Food Chem Toxicol, 44, 236-241.
- Nak D, Mısırlıoğlu D, Nak Y, et al (2004): Bir köpekte meme metastazlı transmissible venereal tümör olgusu. Vet Bil Derg, 20, 99-102.
- **19.** Paglia DE, WN Valentine (1967): Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. J Lab Clin Med, **70**, 158-169.
- 20. Pilger A, Ivancsits S, Germadnik D, et al (2002): Urinary excretion of 8-hydroxy-2'-deoxyguanosine measured by high-performance liquid chromatography with electrochemical detection. J Chromatogr B, 778, 393-401.
- SPSS (2014): IBM SPSS Statistics for Windows, version 22.00. SPSS Inc., Chicago, IL.
- 22. Strakova A, Murchison EP (2014): The changing global distribution and prevalence of canine transmissible venereal tumour. BMC Vet Res, 10, 168.
- Sun Y, Oberley LW, Li Y (1988): A simple method for clinical assay of superoxide dismutase. Clin Chem, 34, 497-500.
- Tanja P, Alenka NS, Butinar J, et al (2008): Antioxidant status in canine cancer patients. Acta Vet Beograd, 58, 275-286.
- 25. Tüközkan N, Erdamar H, Seven I (2006): Measurement of total malondialdehyde in plasma and tissues by highperformance liquid chromatography and thiobarbituric acid assay. Firat Tip Derg, 11, 88-92.
- Urso ML, Clarkson PM (2003): Oxidative stress, exercise and antioxidant supplementation. Toxicology, 189, 41-54.
- 27. Welsh JS (2011). *Contagious Cancer*. The Oncologist, 16, 1-4.
- Winter JL, Barber LG., Freeman L, et al (2009): Antioxidant status and biomarkers of oxidative stress in dogs with Lymphoma. J Vet Intern Med, 23, 311-316.
- Wiseman H, Kaur H, Halliwell B (1995): DNA damage and cancer: measurement and mechanism. Cancer Letters, 93, 113-120.
- **30.** Wu LL, Chiou CC, Chang PY, et al (2004): Urinary 8-OHdG: a marker of oxidative stress to DNA and a risk factor for cancer, atherosclerosis and diabetics Clin Chim Acta, **339**, 1-9.
- **31.** Zhang F, Stott WT, Clark AJ, et al (2007): *Quantitation* of 8-hydroxydeoxyguanosine in DNA by liquid chromatography/positive atmospheric pressure photoionization tandem mass spectrometry. Rapid Commun Mass Spectrom, **21**, 3949-3955.

Ankara Üniv Vet Fak Derg, **67**, 127-131, 2020 DOI: 10.33988/auvfd.523958

# Determination of serum amyloid A, haptoglobin and hepcidin levels in calves with endemic viral pneumonia

#### Ali Burak DÖRTKARDEŞ<sup>a</sup>, Şima ŞAHİNDURAN<sup>b,⊠</sup>

<sup>1</sup> Burdur Mehmet Akif Ersoy University, Faculty of Veterinary Medicine, Department of Internal Medicine, Burdur, Turkey. <sup>a</sup>ORCID: 0000-0002-9239-1226; <sup>b</sup>ORCID: 0000-0002-7718-2020

<sup>⊠</sup>Corresponding author: sahinuran@mehmetakif.edu.tr Received date: 07.02.2019- Accepted date: 20.12.2019

**Abstract:** The aim of this study was to investigate serum amyloid A, haptoglobin and hepcidin levels in respiratory infections caused by various viral agents in calves. Pneumonia is an infectious or non-infectious disease affecting the lungs of all animals, and during the inflammation, blood levels of acute phase proteins increase rapidly. Hepcidin is a hormone that has many functions and is known as type II acute phase reactant. The blood samples were collected from a total of 20 calves to form the study group. The control group consisted of 10 healthy animals. Sera were analyzed with ELISA for antigen detection. According to the results, Bovine herpesvirus type-1 (BHV-1) was positive in 3 calves, Bovine viral diarrhea virus (BVDV) was positive in 3 calves, Bovine respiratory syncytial virus (BRSV) was positive in 1 calf, and Bovine parainfluenza virus type 3 (BPI3) was positive in 1 calf. Adenovirus infection was detected in 5 animals together with other viruses. Mixed infections were observed in 12 calves. Haptoglobin, serum amyloid A and hepcidin values were measured in sera. According to the results, statistically significant differences were observed between the two groups for the haptoglobin (P<0.01), serum amyloid A (P<0.05) and hepcidin (P<0.05). It was concluded that these parameters can be used to support the diagnosis of respiratory infections of the calves.

Keywords: Calf, haptoglobin, hepcidin, serum amyloid A, viral pneumonia.

# Yaygın viral pnömonili buzağılarda serum amiloid A, haptoglobin ve hepsidin düzeylerinin belirlenmesi

Özet: Bu çalışmanın amacı, buzağılarda çeşitli viral etkenlerinin neden olduğu solunum yolu enfeksiyonlarında serum amiloid A, haptoglobin ve hepcidin düzeylerini araştırmaktır. Pnömoni, tüm hayvanların solunum sistemlerini etkileyen, enfektif veya nonenfektif sebeplere bağlı olarak gelişen bir hastalık olup, yangı sırasında akut faz proteinlerinin kandaki değerleri hızla artmaktadır. Hepsidin; birçok fonksiyona sahip olan bir hormon olup aynı zamanda tip II akut faz reaktant olduğu bilinmektedir. Araştırmanın çalışma grubunu oluşturmak için, toplam 20 buzağıdan kan örnekleri alındı. Kontrol grubu ise, sağlıklı 10 hayvandan oluşturuldu. Kan serumları, antijen tespiti için ELISA testi ile tarandı. Elde edilen sonuçlara göre 3 hastada Bovine herpes virüs tip 1 (BHV-1), 3 hastada Bovine viral diarrhea virus (BVDV), 1 hastada Bovine respiratory syncytial virüs (BRSV) ve 1 hastada Bovine parainfluenza virüs tip 3 (BPI3) pozitifti. Adenovirüs enfeksiyonu 5 hayvanda diğer virüslerle birlikte tespit edildi. Oniki hastada miks enfeksiyon saptandı. Haptoglobin, serum amiloid A ve hepsidin değerleri toplanan serum örneklerinde ölçüldü. Sonuç olarak; iki grup arasında, haptoglobin (P<0,01), serum amiloid A (P<0,05) ve hepsidin (P<0,05) farkları istatistiksel açıdan önemli bulunarak, buzağıların solunum sistemi enfeksiyonlarının teşhisini desteklemede, belirtilen parametrelerin kullanabileceği saptandı.

Anahtar sözcükler: Buzağı, haptoglobin, hepsidin, serum amiloid A, viral pnömoni.

#### Introduction

Pneumonia is an infectious and non-infectious originated disease affecting lungs of all animals. There are many factors that lead the disease. It is a disease which causes serious financial losses especially in cattle farms. Pneumonia is the inflammation of the lung parenchyma. It is usually associated with inflammation of bronchioles and pleuritis. Clinically; it is characterized by fever, nasal discharge, increased respiratory rate, changes in depth and characteristics of breaths, cough, abnormal breathing sounds in auscultation, lacrimation, depression and partial anorexia (2, 3). In dairy calf pneumonia; in acute outbreaks, many animals have been reported to be depressed and have a high fever. In some calves; there are visible signs in the respiratory system (22).

In the acute phase of the inflammation, proteins that show significant changes in blood levels are called acute phase protein (APP). Acute phase proteins are used to assess the response of the body's immune system to inflammation or trauma (15, 19). Their secretion is regulated by the proinflamatory cytokines, in particular interleukin 6 (IL-6) (15). In addition to diarrhea, respiratory diseases are one of the major health problems affecting young dairy calves. In order to prevent outbreaks of disease, early detection, isolation and treatment of sick animals are important. The results obtained in several different studies have shown that APPs are useful for the detection and monitoring of respiratory diseases. In calves with respiratory tract diseases, the following APPs were determined: haptoglobib (Hp), fibrinogen (Fb), serum amyloid A (SAA), transferrin (Tf), lipopolysaccharide binding protein (LBP), α1-acid glycoprotein (AGP), α1antitrypsin (a1-AT), seromucoid (Sm), ceruloplasmin (Cp), albumin and  $\alpha$ 2-macroglobulin (21).

Hepcidin is a hormone that has multiple functions and a peptide structure (14). Hepcidin has been studied in human and many animal species (mice, rats, pigs, fish, dogs) to date. In a study on healthy canine tissues, hepcidin was found to be secreted mostly in the liver and less in the lungs and kidneys but not in other tissues (9). Hepatic hepcidin production is under the influence of many stimulants such as low levels of iron and erythropoietic activation. Also some cytokines, especially IL-6, increase the level of hepcidin (13). Hepcidin also assists host defense due to its direct antimicrobial properties (8, 23). Inflammation states stimulate hepcidin production and increase its release, leading to a decrease in iron secretion from macrophages and a decrease in plasma iron levels (5, 16).

The aim of this study was to investigate serum amyloid A, haptoglobin and hepcidin levels in respiratory infections caused by various viral agents in calves.

#### **Material and Methods**

This research was carried out on the basis of the permission of Mehmet Akif Ersoy University Local Animal Ethics Committee dated 05.10.2016 and numbered 214. The research material consisted of Holstein breed calves from the farms in Burdur Province that were brought to Veterinary Hospital of Mehmet Akif Ersoy University. The owners of the animals in the study were informed about the applications. The blood samples were collected from the animals of the study group with high respiratory rate, wheezy respiratory, cough and seroprulant nasal discharge.

Blood samples were collected from a total of 20 calves according to the criteria mentioned above. These samples were gathered out of Holstein calves with 8 females and 12 males (aged 2 - 6 months). The control group consisted of 10 healthy calves (aged 2-6 months) from another farm. Complete blood counts of the calves

were performed in both groups (Diatron Abacus Junior Vet Hematology Analyzer, S / N 130702 model).

The blood samples were coagulated, and their sera were separated in the cooled centrifuge at 4000 rpm for 5 min. The resulting serum samples were transferred to Eppendorf tubes (1.5 ml) evenly using micropipette. The tubes were recorded with their sample numbers, and stored at -20°C until used.

Twenty blood serum collected from suspected animals with clinical signs of pneumonia were analyzed with ELISA test for the detection of causative agents antigens (Bio-X Diagnostics S.A ELISA kit for serodiagnosis of Bovine herpesvirus type-1 (BHV-1)), Bovine viral diarrhea virus (BVDV), Bovine respiratory syncytial virus (BRSV), Bovine parainfluenza virus type 3 (BPI3) and Adenovirus 3. Sera were diluted 1/100 before inoculation. The samples were analyzed with ELISA according to the manufacture's recommendations. In the ELISA reader, the percentages of the agents in the blood were determined numerically. The results were evaluated according to the evaluation of the agent positivity given in the kit procedure. Bovine specific haptoglobin, serum amyloid A and hepcidin ELISA (Enzyme-Linked Immunosorbent Assay) kits (MyBioSource, San Diego (USA)) were used for the measurement of their levels in sera of study and control group animals. In order to increase the reliability of the study, serum samples were run in duplicate.

*Statistical analysis:* The obtained data were evaluated by the 2-Sample t test method between the study and control groups. Data were given as mean and standard error. P value of <0.05 was considered significant.

#### Results

Agent positivity evaluation intervals are shown in Table 1. According to the results, BHV-1 was positive in 3 calves, BVDV was positive in 3 calves, BRSV was positive in 1, and BPI3 was positive in 1 calf. Mixed infections were determined in 12 calves (Table 2).

The common symptoms of calves were cough, wheezing, seroprulent respiratory nasal discharge and depression. The clinical findings of calves in study and control groups are given in Table 3. While there was no difference in the body temperature between the two groups, the number of respiratory (P<0.01) and pulse rate (P<0.05) of the calves in study group were found to be higher than in those in control group. When the hematological parameters of the control and study groups were evaluated, the difference between the values of leukocyte (P<0.01), monocyte (P<0.01), neutrophils (P<0.01), and hematocrit (P<0.05) were found to be significant. No significant difference was found between lymphocyte, erythrocyte, hemoglobin and platelets values (P>0.05) (Table 4).

	0	+	++	+++	++++	+++++
BHV-1	V≤30	<v≤67< td=""><td><v≤104< td=""><td><v≤141< td=""><td><v≤178< td=""><td>178&lt;</td></v≤178<></td></v≤141<></td></v≤104<></td></v≤67<>	<v≤104< td=""><td><v≤141< td=""><td><v≤178< td=""><td>178&lt;</td></v≤178<></td></v≤141<></td></v≤104<>	<v≤141< td=""><td><v≤178< td=""><td>178&lt;</td></v≤178<></td></v≤141<>	<v≤178< td=""><td>178&lt;</td></v≤178<>	178<
BVDV	V≤20	<v≤40< td=""><td><v≤60< td=""><td><v≤80< td=""><td><v≤100< td=""><td>100&lt;</td></v≤100<></td></v≤80<></td></v≤60<></td></v≤40<>	<v≤60< td=""><td><v≤80< td=""><td><v≤100< td=""><td>100&lt;</td></v≤100<></td></v≤80<></td></v≤60<>	<v≤80< td=""><td><v≤100< td=""><td>100&lt;</td></v≤100<></td></v≤80<>	<v≤100< td=""><td>100&lt;</td></v≤100<>	100<
BRSV	V≤20	<v≤40< td=""><td><v≤60< td=""><td><v≤80< td=""><td><v≤100< td=""><td>100&lt;</td></v≤100<></td></v≤80<></td></v≤60<></td></v≤40<>	<v≤60< td=""><td><v≤80< td=""><td><v≤100< td=""><td>100&lt;</td></v≤100<></td></v≤80<></td></v≤60<>	<v≤80< td=""><td><v≤100< td=""><td>100&lt;</td></v≤100<></td></v≤80<>	<v≤100< td=""><td>100&lt;</td></v≤100<>	100<
BPI3	V≤20	<v≤40< td=""><td><v≤60< td=""><td><v≤80< td=""><td><v≤100< td=""><td>100&lt;</td></v≤100<></td></v≤80<></td></v≤60<></td></v≤40<>	<v≤60< td=""><td><v≤80< td=""><td><v≤100< td=""><td>100&lt;</td></v≤100<></td></v≤80<></td></v≤60<>	<v≤80< td=""><td><v≤100< td=""><td>100&lt;</td></v≤100<></td></v≤80<>	<v≤100< td=""><td>100&lt;</td></v≤100<>	100<
Adeno 3	V≤20	<v≤33< td=""><td><v≤56< td=""><td><v≤79< td=""><td><v≤102< td=""><td>102&lt;</td></v≤102<></td></v≤79<></td></v≤56<></td></v≤33<>	<v≤56< td=""><td><v≤79< td=""><td><v≤102< td=""><td>102&lt;</td></v≤102<></td></v≤79<></td></v≤56<>	<v≤79< td=""><td><v≤102< td=""><td>102&lt;</td></v≤102<></td></v≤79<>	<v≤102< td=""><td>102&lt;</td></v≤102<>	102<

**Table 1.** Agent positivity evaluation intervals.

V: Value, +: Strength of positivity.

BHV-1: Bovine herpesvirus type 1; BVDV: Bovine viral diarrhea virus; BRSV: Bovine respiratory syncytial virus; BPI3: Bovine parainfluenza virus type 3; Adeno 3: Adebovirus type-3.

Table 2. Percentages of	pneumonia agents in ca	lves of study group.
-------------------------	------------------------	----------------------

Calves (No)	BHV-1	BVDV	BRSV	BPI3	Adeno3
	(% agent)	(% agent)	(% agent)	(% agent)	(% agent)
1	17.2	22.9	36.77+	65.62+++	15.34
2	30.5	36.3	59.16++	10.61	15.6
3	119.0	170.2+++++	101.1+++++	108.7+++++	81.6++++
4	20.7	16.4	58.38++	55.3++	52.1++
5	96.7++	9.09	16.44	15.3	10.5
6	22.9	161.3+++++	70.94+++	69.1+++	16.3
7	15.6	11.7	9.5	31.97+	13.9
8	17.6	32.88+	$23.05^{+}$	19.07	15.9
9	100.7++	13.54	15.11	10.6	138.3+++++
10	31.0	84.319++++	85.72++++	103.3+++++	56.7++
11	32.0	156.25+++++	98.61++++	110.6+++++	108+++++
12	26.3	86.45++++	21.22	14.5	10.9
13	18.8	167.6+++++	67.27+++	14.5	18.9
14	36.9+	7.74	18.8	20.1	7.4
15	82.1++	18.8	17.27	17.3	11.9
16	129.6++++	168.3+++++	68.61+++	80.7+++	15.8
17	20.6	45.6++	17.2	18.87	12.9
18	11.1	58.08++	13.38	20.16	9.22
19	15.5	30.75++	25.33+	33.8++	19.5
20	19.7	85.11++++	54.83++	77.7+++	13.7

+ : Strength of positivity.

Table 3. Comparison of clinical findings of calves in control and study groups.

Parameters	Study group (n= 20)	Control group (n= 10)	Р
	Mean±SD	Mean±SD	
Respiratory rate/ min	72.8±4.6	34.8±1.6	< 0.001
Pulse rate/ min	101.9±4.4	84.1±3.3	0.005
Body temperature/ °C	38.49±0.21	38.64±0.10	0.504

Table 4. Mean values of some hematological parameters of calves in control and study groups.

Parameters	Study group	Control group	Р
	( <b>n=20</b> )	( <b>n=10</b> )	
WBC (10^9/l)	18.42±2.07	9.185±0.691	< 0.001
LYM (10^9/I)	$6.22 \pm 0.70$	$6.60{\pm}0.56$	0.674
MON (10^9/l)	$0.963 \pm 0.2$	$0.092{\pm}0.011$	< 0.001
NEU <sup>(10^9/L)</sup>	11.24±1.6	2.73±0.39	< 0.001
RBC (10^12/l)	$8.38{\pm}0.5$	9.17±0.32	0.192
HGB(g/dl)	8.35±0.43	8.77±0.39	0.480
HCT (%)	26.17±1.7	30.0±1.2	0.036
MCV(fl)	31.75±1.2	33.60±1.0	0.250
PLT (10^9/I)	923±393	707±51	0.593

WBC: Leukocyte, LYM: Lymphocyte, MON: Monocytes, NEU: Neutrophil, RBC: Erythrocyte, HGB: Hemoglobin, HCT: Hematocrit, MCV: Mean cell volume, PLT: Platelets.

Table 5. Statistical evaluation of haptoglobin, hepcidin and serum amyloid A values in both groups.

Parameters	Study group (n=20)	Control group (n=10)	Р
Haptoglobin (ng/ml)	197±20	1.00±1.6	< 0.001
Serum amyloid A (ng/ml)	4.89±1.2	$0.66{\pm}0.73$	0.005
Hepcidin (ng/ml)	$0.833 \pm 0.055$	$0.073 \pm 0.800$	0.005

A moderate difference was found between hepcidin and serum amyloid A values of the control and study groups (P<0.05). The most significant difference was observed in the haptoglobin (P<0.01). The values of haptoglobin, hepcidin and serum amyloid A were showed in Table 5.

#### **Discussion and Conclusion**

In this study, while there was no difference in body temperature between the two groups, the number of breath and pulse rates of the calves in study group per minute was found to be higher than in those in control group (P<0.01, P<0.05, respectively). Our results were similar to the results of Y1lmaz and Gökçe (24). The lack of fever in the animals in the study group is thought to be due to the passing of the viremia period.

When the hematological parameters of the animals in the study and control groups were evaluated, it was determined that leukocytosis was related to acute respiratoric inflammation due to infection and it was found to be similar to the other study findings (24). There was a difference in neutrophil counts (P<0.01) and hematocrit values (P=0.036), but no significant difference was found in other parameters. Since no bacterial agents were investigated in the present study, secondary bacterial infection is highly possible.

Acute phase proteins have been accepted as a valuable indicator in determining the importance of pathological conditions in humans and animals. In cattle, SAA and Hp take place at the forefront of important acute-phase proteins, but their responses differ from individual animals.

In the case of viral and bacterial infections, acute phase response can be differ. For example, acute phase response to bacterial infections is reported to be more severe than viral infections in bovine mastitis (6).

Alsemgeet et al. (1) reported that haptoglobin was not detected in healthy cows and newborn calves. In the same study serum amyloid A level was found to be lower in healthy calves compared to adult cows, whereas in calves with inflammatory diseases; it has been reported to be significantly high. Niine et al. (17) reported that SAA and Hp levels were significantly increased in calves with Giardia and Cryptosporidium infections compared to healthy calves. Kabu et al. (12) reported a statistically significant increase in the concentrations of serum Hp and SAA in water buffaloes with dermatosis compared to the control group. Sahinduran et al. (20) reported that serum Hp and SAA concentrations, in cattle with BVD and BHDV-1 in single and dual infection, have shown statistically significant increase compared to healthy cattle. In the other study, significant increases were observed in serum Hp and SAA concentrations in calves suspected from Bovine respiratory disease (10). Also significant increase in the SAA concentration had been observed in calves with enteritis and pneumonia compared to the control group (11). Coşkun et al. (4) reported that SAA and Hp levels in both of the blood serum and the bronchoalveolar lavage were higher in calves with bronchopneumonia than in healthy calves. In our study, SAA and Hp levels were significantly higher in calves with pneumonia than in healthy calves (P<0.05 and P<0.00, respectively). It was thought that the increase in the values in these parameters was the result of acute phase response. These results are in agreement with above mentioned studies. Especially in cattle, SAA and Hp levels were found to be a suitable marker for site conditions in early detection of the diseases with viral pneumonia. In addition, the levels of these parameters were higher in patients with BVDV and in animals with mixed infections in this study.

Hepcidin plays a fundamental role in the regulation of iron metabolism. Hepcidin is an acute phase protein that is synthesized to restrict the body's iron stores and to being requisitioned by invading prevent iron microorganisms (18) and inflammation states stimulate hepcidin production (5). Hepcidin also helps host defense due to its direct antimicrobial properties (8, 23). Erkilic et al. (7), reported that serum hepcidin levels were very high in calves suspected of septicemia, and these levels decrease drastically after treatment. In our previous study on 56 cattle with single and dual infection (BHV-1, BVDV), hepcidin levels were compared. It was found that there was a significant difference between the serum hepcidin concentrations of cattle with single and dual infection and the control group of cattle (20). The results of this study were similar to the above mentioned study and the difference between serum hepcidin concentration of calves in study group and serum concentrations of calves in control group was significant (P<0.05).

Considering all three parameters (SAA, Hp and hepcidin) in our study, haptoglobin level was higher than

the other two parameters. Many studies have indicated the significance of Hp as a clinically useful parameter for measuring the occurrence and severity of inflammatory responses in cattle with mastitis, pneumonia, enteritis, peritonitis, endocarditis, abscesses, endometritis and other natural or experimental infectious conditions (6). Accordingly, haptoglobin which is important acute phase protein in bovine, can have more importance in respiratory infections in calves.

As a conclusion, hepcidin, serum amyloid A and especially haptoglobin were found to be significantly increased in calves with viral pnemonia.

#### Acknowledgements

It was summarised from a first author's master's thesis.

#### **Financial Support**

This study was supported by Mehmet Akif Ersoy University Scientific Research Projects Commission (Project No: 0421-YL-17).

#### **Conflict of Interest**

The authors declare that there have no conflict of interests.

#### References

- 1. Alsemgeest SPM, Kalsbeek HC, Wensing T, et al (1994): Concentrations of serum amyloid-a (SAA) and haptoglobin (Hp) as parameters of inflammatory diseases in cattle. Vet Quart, 16, 21-23.
- Ames TR (1997): Dairy Calf Pneumonia. The disease and its impact. Vet Clin North Am Food Anim Pract, 13, 379-391.
- **3.** Bryson DG, McNulty MS, McCracken RM (1983): Ultrastructural features of experimental parainfluenza type 3 virus pneumonia in calves. J Comp Pathol, **93**, 397-414.
- 4. Coskun A, Guzelbektes H, Simsek A, et al (2012): Haptoglobin and SAA concentrations and enzyme activities in bronchoalveolar lavage fluids from calves with bronchopneumonia. Revue Méd Vét, 163, 615-620.
- 5. Coyne DW (2011): *Hepcidin: clinical utility as a diagnostic tool and therapeutic target.* Kidney Int, **80**, 240-244.
- 6. Eckersall PD, Bell R (2010): Acute phase proteins: Biomarkers of infection and inflammation in veterinary medicine. Vet J, 185, 23-27.
- 7. Erkilic EE, Erdogan HM, Ogun M, et al (2016): Relationship between hepcidin and oxidant/antioxidant status in calves with suspected neonatal septicemia Vet World, 9, 1238-1241.
- 8. Falzacappa MVV, Muckenthaler MU (2005): Hepcidin: iron-hormone and anti-microbial peptide. Gene, 364, 37– 44.

- Fry MM, Liggett JL, Baek SJ (2004): Molecular cloning and expression of canine hepcidin. Vet Clin Path, 33, 223-227.
- Joshi V, Gupta VK, Bhanuprakash AG, et al (2018): Haptoglobin and serum amyloid A as putative biomarker candidates of naturally occurring bovine respiratory disease in dairy calves. Microb Pathogenesis, 116, 33-37.
- 11. Kabu M, Elitok B, Kucukkurt I (2016): Decetion of serum amyloid-A concentration in the calf clinically diagnosed with pneumonia, enteritis and pneumoenterites. Ciência Rural, 46, 293-299.
- Kabu M, Sayın Z (2016): Concentrations of serum amyloid A, haptoglobin, tumour necrosis factor and interleukin-1 and -6 in Anatolian buffaloes naturally infected with dermatophytosis. Vet Med-Czech, 61, 133-135.
- **13.** Kemna E, Pickkers P, Nemeth E, et al (2005): *Timecourse analysis of hepcidin, serum iron, and plasma cytokine levels in humans injected with LPS.* Blood, **106**, 1864-1866.
- 14. Krause A, Neitz S, Magert HJ, et al (2000): LEAP-1, a novel highly disulfide-bonded human peptide, exhibits antimicrobial activity. FEBS Lett, 480, 147–150.
- **15.** Murata H, Shimada N, Yoshioka M (2004): Current research on acute phase proteins in veterinary diagnosis. Vet J, **168**, 28-40.
- **16.** Nemeth E, Valore EV, Territo M, et al (2003): *Hepcidin, a putative mediator of anemia of inflammation, is a type II acute-phase protein.* Blood, **101**, 2461-2463.
- 17. Niine T, Peetsalu K, Nieminen M, et al (2017): Giardia and Cryptosporidium infections in neonatal reindeer calves: Relation to the acute phase response. Comp Immunol Microb, 54, 45-50.
- **18.** Orro T, Jacobsen S, LePage JP, et al (2008): Temporal changes in serum concentrations of acute phase proteins in newborn dairy calves. Vet J, **176**, 182-187.
- **19.** Petersen HH, Nielsen JP, Heegaard PMH (2004): Application of acute phase protein measurements in veterinary clinical chemistry. Vet Res, **35**, 163-187.
- 20. Sahinduran Ş, Kale M, Kıyıcı R, et al (2017): Some acute phase proteins and hepcidin levels in single and dual infection with BVD and BHV-1. MAKÜ Sag Bil Enst Derg, 5, 115-123.
- **21.** Svensson C, Lundborg K, Emanuelson U, et al (2003): Morbidity in Swedish dairy calves from birth to 90 days of age and individual calf-level risk factors for infectious diseases. Prev Vet Med, **58**, 179–197.
- 22. Waltner-Toews D, Martin SW, Meek AH (1978): Dairy calf management, morbidity and mortality in Ontario Holstein herds. II: Age and seasonal patterns. Prev Vet Med, 4, 125-135.
- **23.** Wessling-Resnick M (2010): Iron Homeostasis and the Inflammatory Response. Annu Rev Nutr, **30**, 105–122.
- 24. Yılmaz O, Gökçe G (2017): Sığırlarda enfeksiyöz solunum sistemi hastalıkları kompleksinde (brdc) klinik, hematoloji, biyokimya, oksidatif stres, akut faz proteinler üzerinde araştırmalar. Ataturk Universitesi Vet Bil Derg, 12, 34-44.

Ankara Üniv Vet Fak Derg, **67**, 133-136, 2020 DOI: 10.33988/auvfd.523745

### Antioxidant activities, total phenolic and flavonoid contents of honey collected from different botanical origins

#### Deniz AKER<sup>a</sup>, Cevat NİSBET<sup>b,⊠</sup>

Ondokuz Mayıs University, Faculty of Veterinary Medicine, Department of Biochemistry, Samsun, Turkey. <sup>a</sup>ORCID:0000-0003-3959-6713; <sup>b</sup>ORCID: 0000-0002-7042-4605

<sup>⊠</sup>Corresponding author: cnisbet@omu.edu.tr Received date: 07.02.2019- Accepted date: 21.12.2019

**Abstract:** In this study, it is aimed to determine the presence of antioxidant capacity, total phenolic and flavonoid contents in six different types (multiflora, pine, chestnut, sunflower, acacia, citrus) and eventually 65 samples of honey from different parts of Turkey. Pollen analysis of all honey samples in the laboratory was carried out to determine the purity (> 65-70%) of the plant source. Total phenolic content determined in honey samples was found the highest value in pine honey 166.46 ± 5.80 (mgGAE / 100 g honey) while the lowest value was found in flower honey with (106.04 ± 9.55). The level of flavonoid contents of the groups was lowest on the flower and citrus honey ( $1.3 \pm 0.2$  and  $1.6 \pm 0.1$ ) and the highest value were on chestnut and pine ( $2.7\pm0.4$  and  $2.8 \pm 0.2$ ) were detected. Comparing the radical scavenger activity in honey groups, the activity of chestnut honey was the highest (100.54 ± 22.72). The results of this study show that the phytochemical structure and biological activity of honey are completely different from each other depending on the plant source. On the other hand, it is possible to say that the antioxidant, phenolic, and flavonoid values are high, which is a good indicator of the quality and naturalness of honey.

Keywords: Antioxidant, flavonoid, honey, honey bee, phenolic acid.

#### Farklı botanik kaynaklardan elde edilen balların antioksidan aktiviteleri, toplam fenolikleri ve flavonoid içerikleri

**Özet:** Bu çalışmada, Türkiye'nin çeşitli bölgelerinde üretilmiş 6 farklı çeşit (yayla, çam, kestane, ayçiçeği, akasya, narenciye) toplam 65 bal numunesinde toplam fenolik asit, flavonoid düzeyi ve total antioksidan aktivitesi incelenmiştir. Laboratuvarda tüm bal numunelerinin polen analizi yapılarak bitki kaynağının saflık derecesi (>%65-70) tespit edilmiştir. Bal örneklerinde belirlenen total fenolik içeriği 166,46 ± 5,80 (mgGAE/100g bal) ile en yüksek değer çam balına ait iken, en az 106,04 ± 9,55 ile çiçek balında bulunmuştur. Gruplar arası flavonoid içeriği en düşük düzey çiçek ve narenciye balında sırasıyla (1,3 ± 0,2 ve 1,6 ± 0,1), en yüksek değer ise kestane ve çam balında (2,7±0,4 ve 2,8 ± 0.2) tespit edilmiştir. Bal gruplarında yapılan radikal süpürücü aktivitesinin karşılaştırılmasında ise kestane balın aktivitesinin en yüksek olduğu (100,54 ± 22,72) ortaya konulmuştur. Çalışmanın sonuçları, balın fitokimyasal yapısı ve biyolojik aktivitesinin bitki kaynağına bağlı olarak birbirinden tamamen farklı olduğunu göstermektedir. Diğer taraftan bu değerlerin yüksek olması balın kalitesinin ve doğallığının iyi bir göstergesi olacağını söylemek mümkündür.

Anahtar sözcükler: Antioksidan, bal, balarısı, fenolik asit, flavonoid.

#### Introduction

For centuries, bees and bee products have been used for therapeutical purposes. Today, they maintain their actuality in the field of supportive therapy and show a fast development under the name of apitherapy. Honey, which is one of the apitherapy products and one of the oldest nutritional sources of mankind, is a sweet product which bees collect from flower nectars or from secretions of some insects living on plants and store in honeycombs by exposing them to enzymatic change (3, 6, 22). The formation and composition of honey produced naturally differ significantly according to regions and sources of the plant (23). The composition of plant nectar differs according to the geography the plant is grown in, soil fertility, rainfall, light, altitude and a great number of other environmental factors (14, 24). In other words, the quality and biochemical characteristics of honey differ according to the source of nectar (8, 13, 22). Studies conducted show that the antioxidant activity of natural foods is higher than food products with synthetic structure (18). For this reason, daily intake of antioxidant food is important and required for free radicals which form as a metabolism product in the organism. In addition to being a food product and source of energy, honey is also important for human health as a natural antioxidant source due to various phytochemicals it includes (3, 12, 17).

Antioxidant compositions of honey are enzymatic (glucose oxidase, catalase, peroxidase) or non-enzymatic (phenolic acids, flavonoids, ascorbic acid, tocopherol, carotenoids) (18, 19). Phenolic acids contribute significantly to the antioxidant activity of honey (1, 17). Flavonoids have a significant antioxidant and antiinflammatory function by providing free-radical scavenging, inhibiting cyclooxygenase and lipooxygenase enzymes, chelating transition metals such as iron and copper, protecting  $\alpha$ -tocopherol at LDL and providing oxidizable ascorbic acid regeneration (16). The rates of these compositions in honey differ significantly depending on the source flora honey is collected from and the structure of the geographical areas (8, 12, 23). For this reason, every honey has a different apitherapic value. The present study researches the biological activities of honey obtained from different regions and plant sources of Turkey.

#### **Materials and Methods**

*Sampling:* The study was conducted with a total of 65 honey samples from different regions of Turkey (11 sample meadow honey from the provinces of Erzurum and Sivas, 11 sample pine honey from Muğla, 11 sample chestnut honey from Sinop, 11 sample Acacia honey from Trabzon, 10 sample from citrus tree honey from Antalya and 11 sample sunflower honey from Samsun). Pollen analyses of all samples were made in the laboratory and purity degree of plant sources was found (>65-70%).



Biochemical analysis: Calculation of the total antioxidant effect in the samples was made according to the free radical scavenging effects of the prepared extracts on DPPH (1,1-diphenyl-2-picrylhydrazyl) in honey by using an indirect method. As a result of DPPH radical scavenging activity, IC50 values were assessed as mg/ml. The measurement was made according to Meda and Dimins method modified at 520 nm with a spectrophotometer (9, 21). AA [%] = (Abs cont-Abssample)/ Abs cont.X100. Total phenolic concentration level was read at 750 nm in spectrophotometer with Folin-Ciocalteau method and the total phenol amount was calculated as equivalent to mg gallic acid in 100 gram extract (4, 9). The Dowd method was used in total flavonoid analysis. In this method, honey solutions prepared based on quercetin (mgQE/100gr) concentration in mg in a kilogram of honey were read at 415 nm with a spectrophotometer (9, 21).

*Statistical analysis:* The data obtained from the study were evaluated with the variance analysis (ANOVA) technique in factorial order and the differences between the means were determined by Duncan multiple comparison test. Statistical evaluations were made using the SPSS statistical program (26).

#### **Results**

Average values of total phenolic, flavonoid levels and antioxidant activity found in honey samples of groups are given in (Table 1, Figure 1).

Figure 1. Means of total phenolic, flavonoid and antioxidant analysis results in honey samples

**Table 1.** Means and standard errors of total phenolic (mg GAE/100 g), flavonoid QE/100 g) and antioxidant (mg/ml) in honeys samples produced from different botanical origins.

Samples	Total flavonoid	(Min-Max)	Total phenolic	(Min-Max)	Antioxidant, DPPH-sc50	(Min-Max)
Citrus tree honey	$1.6\pm0.1$	1.2 -1.9	$113.77\pm4.27$	100.21-120.62	$152.65\pm4.96$	140.40 -163.91
Chestnut honey	$2.7\pm0.4$	1.4 -4.0	$138.27\pm19.31$	97.64 -206.22	$100.54\pm22.72$	41.05 -165.03
Sunflower honey	$1.8\pm0.5$	1.7 -2.0	$127.27\pm13.54$	88.03 -127.33	$167.33\pm9.69$	135.40 -198.81
Pine honey	$2.8\pm0.2$	2.1 -3.6	$166.46\pm5.80$	149.37-186.11	$145.48 \pm 16.49$	93.61 -206.38
Meadow honey	$1.3\pm0.2$	0.5 -1.9	$106.04\pm9.55$	68.85 -130.94	$270.86 \pm 51.22$	136.17 -506.38
Acacia honey	$2.1 \pm 0.4$	1.3 -3.5	$143.71 \pm 16.99$	71.16 -183.06	$108.91\pm54.00$	69.34 -142.55

The highest total phenolic composition was found in pine honey with 166.46  $\pm$  5.80 (mgGAE/100g honey), while the lowest value was found in flower honey with 106.04  $\pm$  9.55. While the intergroup flavonoid composition lowest value was found in flower and citrus honey (1.30  $\pm$  0.2 and 1.6  $\pm$  0.1), the highest value was found in chestnut and pine honey (2.7  $\pm$  0.4 and 2.8  $\pm$  0.2). Radical scavenging activity comparison of honey groups showed that chestnut honey had the highest activity (100.54  $\pm$  22.72) As a result of the analysis of variance, there was a significant difference between the honey samples (P <0.01).

#### **Discussion and Conclusion**

Phenolic acids and flavonoids, which are components of honey, are important due to their antimicrobial, antioxidant, anticancer and antioxidative effects and their being associated with human and animal health and on the other hand due to being a criterion for honey's being refined (1, 5, 6, 22). The concentration of these compounds which significantly affects honey's therapeutic activity differs according to plant flora which constitutes the sources of honey (11).

In the present study, it was found that total phenolic composition differed between 106 - 166 mgGAE/100 g in all honey samples. While the lowest concentration was found in flower honey, the highest concentration was found in pine honey (Figure1). Similarities and differences are seen between our study results and the results of the studies previously conducted. While total phenolic acid concentration was reported as 32.59-114.75 mgGAE/100 gr and an average of 74.38  $\pm$  20.54 in South African honey samples (21), phenolic acid concentration was reported as 36.26-102.80 mgGAE/100g and an average of 65.31±19.50 in a study conducted with Nigeria honey (5). Vit et al., (28) reported in their study that phenolic matter amount differed between 38.15 and 182.10 mgGAE/100 g in Venezuela honey. Total phenolic amount in Yemeni honey obtained from different regions and different plants differed between 56.32 and 246.21 mg/100g (1). In a study conducted on the honey in the Indian Bengal region, it was reported that gallic acid rates per 100 g honey differed between 9.9  $\pm$  0.6 and 44.7  $\pm$  2 mg (7). In a study conducted with different honey from Slovenia, the total phenolic composition was found as 241.4 mg/kg in fir tree honey, as 233.9 mg/kg in chestnut honey and as 44.8 mg/kg in acacia honey (4). In studies conducted on acacia honey, total phenolic compositions differ significantly among countries. While Iranian acacia honey total phenolic content was found as 22.9-65.5 mg gallic acid/g (15), it was found as  $627 \pm 44.03$  in Germany acacia honey (2). In a study conducted on the honey of Black Sea flora, average phenolic content was found as 0.224 mg as gallic acid equivalent for 1 g honey sample (11).

In the study, while the lowest average total flavonoid (mgQE/100g honey) value of honey was found in flower honey with an average of  $1.3 \pm 0.2$  the highest value was found in chestnut honey with an average of  $2.7 \pm 0.4$ . In a study conducted on Indian honey, quercetin (mg/100g) amount was found to be between 5.12  $\pm$  0.23 and 19.4  $\pm$ 1.38 (Das et al., 2013). In Brazilian multiflora and citrus honey, quercetin (mg/100g) composition was reported as  $1.96 \pm 1.53$  and  $0.17 \pm 0.15$ , respectively (20). In a study conducted on Malesian honey, flavonoid average value was reported as  $5.42 \pm 0.62$  (19). In China unifloral honey, flavonoid average value was reported to differ between 9.41 and 102.1 (10). In a study conducted in South Africa, the total flavonoid composition average was reported as  $2.57 \pm 2.09$  in 27 honey samples (21). In a study conducted on pine honey in our country, total flavonoid level was found as  $22.80 \pm 2.45$  on average in QE/kg honey equivalent (25). In the present study, this value was found as  $2.8 \pm 0.2$  in the same honey type.

In the study, total antioxidant activity was found as  $100.54 \pm 22.72$  in chestnut honey and as  $270.86 \pm 51.22$  in multiflora flower honey. When studies conducted in many countries were examined in terms of their similarities and differences of these values, it was found that radical scavenger DPPH average IC50 =  $23.92 \pm 1.12$  mg/mL in Indian honey (7), the same value was found as  $10.0 \pm 1.8$ and  $10.7 \pm 2.2$ , respectively in Slovenian chestnut and flower honey (4); average antioxidant activity was found as  $31.96 \pm 18.07$  in chestnut honey in Brazilian multifloral honey and as  $15.22 \pm 10.75$  in citrus honey (20), while antioxidant activity was reported as 35-122 in China unifloral honey (10). Ertürk et al. (11) found IC50 values as between 29.388 and 458.450 mg/mL as a result of the DPPH radical scavenging activity test in Black Sea flora honey in our country.

Some studies show a correlation between the phenolic compound level and antioxidant activity in some types of honey; however, since flavonoids have hydroxyl, they are oxidized very quickly. For this reason, despite structural similarities, the difference between antioxidant activities depends on hydroxylation and methylation degree (1). In our study, no correlation was found between groups. It is possible to say that this is because in addition to flavonoid and phenolic acid, the presence of Vitamin E and C and carotenoids may have influenced total antioxidant activity (27).

Consequently, our observations that phytochemical structure and biological activity of honey differ completely depending on the origins of the plant.

#### Acknowledgments

The authors thank Dr. Serhat Arslan for editing the content of this manuscript related to the statistical analysis. This article is produced from master's thesis of the first author.

#### **Financial Support**

This work was supported by Ondokuz Mayıs University, Scientific Research Projects Unit (BAP) under project number (PYO.VET.1904.13.006).

#### **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### References

- **1.** Al-Mamary M, Al-Meeri A, Al-Habori M (2002): Antioxidant activities and total phenolics of different types of honey. Nutr Res, **22**, 1041-47.
- Alzahrani H, Boukraa L, Bellik Y, et al (2012): Evaluation of the antioxidant activity of three varieties of honey from different botanical and geographical origins. Global J Health Sci, 4, 191-196.
- **3.** Atrott J, Henle T (2009): *Methylglyoxal in Manuka honey* – correlation with antibacterial properties. Czech J Food Sci, **27**, 163-165.
- **4.** Bertoncelj J, Dobersek U, Jamnik M, et al (2007): Evaluation of the phenolic content, antioxidant activity and colour of Slovenian honey. Food Chem, **105**, 822–28.
- Buba F, Gidado A, Shugaba A (2013): Analysis of biochemical composition of honey samples from North-East Nigeria. Biochem Anal Biochem, 2, 139-40.
- Ching H, Hou YC, Hsiu SL, et al (2002): Influnce of honey on the gastrointestinal metabolism and disposition of glycyrhizm and glycyrhetic acid in rabbits. Biol Pharm Bull, 25, 87-91.
- 7. Das A, Mukherjee A, Dhar P (2013): Characterization of antioxidants and antioxidative properties of various unifloral honeys procured from West Bengal, India. IOSR-JESTFT, 7, 56-638.
- Demirezen D, Aksoy A (2010): Determination of heavy metals in bee honey using by inductively coupled plasma optical emission spectrometry (ICP-OES). GU J Sci, 18, 569-575.
- Dimins F, Kuka P, Augspole I (2010): Characterisation of honey antioxidative properties. International Conference of Food Innova, 28-29- Oct. Latvia.
- Dong R, Zheng Y, Xu B (2013): Phenolic profiles and antioxidant capacities of Chinese unifloral honeys from different botanical and geographical sources. Food Bioprocess Technol, 6, 762–770.
- Ertürk Ö, Şahin H, Kolaylı S, et al (2014): Antioxidant and antimicrobial activity of East Black Sea. Turk J Biochem, 39, 99–106.

- Gheldof N, Wang XH, Engeseth NJ (2002): Identification and quantification of antioxidant components of honeys from various floral sources. J Agric Food Chem, 50, 5870-5877.
- **13.** Giorgi A, Madeo M, Baumgartner J, et al (2011): The Relationships between phenolic content, pollen diversity, physicochemical information and radical scavenging activity in honey. Molecules, **16**, 336-342.
- 14. Güler A, Bakan A, Nisbet C, et al (2007): Determination of important biochemical proreties of honey to discriminate pure and adulterated honey with sucrose (Saccharum officinarum L.) syrup. Food Chem, 105, 1119–1125.
- **15. Javanmardia J, Stushnoff C, Locke E, et al** (2003): Antioxidant activity and total phenolic content of Iranian Ocimum accessions. Food Chem, **83**, 547–550.
- Kahraman A, Serters M, kokenT (2002): Flavonoids, Kocatepe Med J, 3, 1-8.
- **17.** Kesic A, Mazalovic M, Crnkic A, et al (2009): *The Influence of L-Ascorbic Acid Content on Total Antioxidant Activity of Bee-Honey.* Eur J Sci Res, **32**, 95-101.
- Kesic A, Crnkic A, Hodžic Z, et al (2012): Influence of Polyphenol Content on total Antioxidant Activity of Honey from Different Botanical and Geographical Origin. Eur J Sci Res, 89, 500-511.
- **19.** Khalil MI, Sulaiman SA, Boukraa L (2010): Antioxidant properties of honey and its role in preventing health disorder. Open Nutraceuticals J, **3**, 6-16.
- Lianda RLP, Sant'ana LD, Echevarria A, et al (2012): Antioxidant activity and phenolic composition of Brazilian honeys and their extracts. J Braz Chem Soc, 23, 618-627.
- **21. Meda A, Lamien CE, Romito M, et al** (2005): Determination of the total phenolic, flavonoid and proline contents in Burkina Fasan honey, as well at their radical scavenging activity. Food Chem, **91**, 571–577.
- 22. Nisbet C, Kazak F, Yuksel A (2018): Determination of quality criteria that allow differentiation between honey adulterated with sugar and pure honey. Biol Trace Elem Res, 186, 288-293.
- 23. Nisbet C, Guler A, Bıyık S (2019): Effects of different environmental conditions on the cognitive function of honeybee (Apis mellifera L.) and mineral content of honey. Ankara Üniv Vet Fak Derg, 66, 95-101.
- 24. Oddo LP, Bogdanov S (2004): Determination of honey botanical origin: problems and issues. Apidologie, 35, 2-3.
- Ozkok A, Darcy B, Sorkun K (2010): Total total phenolic acid and total flavonoid content of Turkish pine honeydew honey. JAAS, 2, 65-71.
- **26.** SPSS. Statistical package for the social sciences User's guide. SPSS Inc. 2004. Chicago IL 60606–6412, USA.
- 27. Vinson JA, Hontz BA (1995): Phenol antioxidant index: comparative antioxidant effectiveness of red and whitewines. J Agric Food Chem, 43, 401–403.
- 28. Vit P, Rodríguez-Malaver A, et al. (2009): Expanded parameters to assess the quality of honey from Venezuelan bees (Apis mellifera). JAAS, 1, 72-81.

Ankara Üniv Vet Fak Derg, **67**, 137-142, 2020 DOI: 10.33988/auvfd. 561824

# Investigation of enrofloxacin and ciprofloxacin residues by LC-MS/MS in chicken liver marketed in Afyonkarahisar

#### Damla ARSLAN-ACARÖZ<sup>1,a,⊠</sup>, Nalan BAYŞU SÖZBİLİR<sup>2,b</sup>

<sup>1</sup>Afyon Kocatepe University, Bayat Vocational School, Afyonkarahisar; <sup>2</sup>Afyon Kocatepe University, Faculty of Veterinary Medicine, Department of Biochemistry, Afyonkarahisar, Turkey. <sup>a</sup>ORCID: 0000-0001-9230-6725; <sup>b</sup>ORCID: 0000-0002-2270-8312

<sup>™</sup> Corresponding author: darslan@aku.edu.tr
Received date: 08.04.2019- Accepted date: 23.11.2019

**Abstract:** Antibiotic residues can remain in animal source foods, mainly in the liver. Enrofloxacin (ENR) is a synthetic fluoroquinolone antimicrobial agent which is extensively used to treat bacterial infections in poultry. This study aimed to evaluate ENR and its main metabolite ciprofloxacin (CIP) occurrence in 100 chicken livers marketed Afyonkarahisar, Turkey. Chicken liver samples were analyzed by LC-MS/MS method with limits of detection for  $1.17 \,\mu$ g/kg (ENR) and  $1.24 \,\mu$ g/kg (CIP). ENR and CIP were detected in 39% and in 31% of samples, respectively. However, the total sum of both antibiotic residue levels was found to be far below the established maximum residue level (200  $\mu$ g/kg) in all of chicken liver samples. In conclusion, according to the results of this study, chicken livers marketed in Afyonkarahisar city are safe for public health regarding ENR and CIP residues. Nonetheless, periodical control of chicken liver is advised due to the high incidence of these antibiotics.

Keywords: Antibiotic residue, chicken liver, ciprofloxacin, enrofloxacin, LC-MS/MS.

#### Afyonkarahisar'da satışa sunulan tavuk karaciğerlerinde enrofloksasin ve siprofloksasin kalıntılarının LC-MS/MS ile araştırılması

Özet: Antibiyotik kalıntıları hayvansal orjinli gıdalarda özellikle karaciğerde bulunabilir. Enrofloksasin (ENR) sentetik bir florokinolon antimikrobiyal ajan olup kanatlılarda bakteriyel enfeksiyonların tedavisinde oldukça yaygın olarak kullanılmaktadır. Bu çalışmada Afyonkarahisar, Türkiye'de satışa sunulan 100 tavuk karaciğerinde ENR ve onun ana metaboliti olan siprofloksasin (CIP) varlığının araştırılması amaçlanmıştır. Tavuk karaciğer örnekleri LC-MS/MS metodu ile analiz edilerek, saptama limitleri ENR için 1,17 µg/kg ve CIP için 1,24 µg/kg olarak belirlenmiştir. ENR ve CIP sırasıyla örneklerin %39 ve %31'inde saptanmıştır. Ancak, tavuk karaciğer numunelerinde her iki antibiyotik kalıntısının toplamı belirlenen maksimum kalıntı düzeyinin (200 µg/kg) oldukça altındadır. Sonuç olarak, Afyonkarahisar ilinde satışa sunulan tavuk karaciğerleri ENR ve CIP kalıntısı bakımından değerlendirildiğinde halk sağlığına risk oluşturmamaktadır. Bununla birlikte, bu antibiyotiklerin yüksek insidensi nedeniyle tavuk karaciğerinin periyodik olarak kontrol edilmesi tavsiye edilmektedir.

Anahtar sözcükler: Antibiyotik kalıntısı, enrofloksasin, LC-MS/MS, siprofloksasin, tavuk karaciğeri.

#### Introduction

Fluoroquinolones, essential drugs for human and animal health, are widely used for the treatment of several diseases in veterinary medicine (21, 24). Enrofloxacin (ENR), a common fluoroquinolone, has a broad spectrum of action and exhibits high efficacy against infectious diseases that are caused by mycoplasma and gram-positive and gram-negative bacteria (14, 21). Furthermore, this antibiotic is used in the treatment of common poultry infections, such as mycoplasmal infections, colibacillosis and pasteurellosis, due to its rapid oral absorption and long elimination half-life (10, 22). ENR shows its effect by interfering in the activities of bacterial DNA-gyrase and topoisomerase IV enzymes and, consequently, it inhibits bacterial DNA replication and transcription (5). Ciprofloxacin (CIP) is the main metabolite of ENR and occurs at different levels in foods of animal origin after the administration of ENR (12). Currently, the presence of contaminants in animal origin products is a major concern regarding food safety (16). Health problems can occur as a result of the excessive use of veterinary drugs in foodproducing animals because most of these substances may produce important toxic effects such as genotoxicity, carcinogenicity and immunotoxicity, as well as endocrine effects, on humans (1, 28). In addition, the improper use of ENR in food-producing animals and not respecting the necessary withdrawal time of this antibiotic may lead to residues in edible animal tissues. The presence of ENR residues in foodstuffs may cause allergic reactions in hypersensitive individuals and could lead to an increased pathogen resistance to clinical drugs in humans; therefore, they may represent important consequences for public health (8,18). To protect food safety and consumer health, Maximum Residue Limits (MRL) have been established by the European Union for pharmacologically active compounds, including ENR. In chicken liver, the MRL is the total sum of ENR and CIP (200  $\mu$ g/kg) (7). Therefore, if the residue levels in the target tissues are lower than the established MRL, the related food is considered safe regarding the presence of these antimicrobials (6).

The liver takes its place in the human diet owing to its fundamental nutritive value in providing dietary iron. However, it should be considered that the liver is a very complex organ due to its role in many drug metabolisms. Therefore, edible animal liver tissue must be monitored regularly regarding the presence of residues (11,16). Chicken liver is one of the favorite foods of animal origin due to its high nutritional value and cheap price.

This study aimed to determine the levels of ENR and its metabolite CIP in chicken liver based on LC-MS/MS, with respect to the European Commission MRL.

#### **Material and Methods**

*Chemicals and reagents:* Methanol, formic acid, ENR and CIP were purchased from Sigma-Aldrich (St. Louis, MO, USA). All other reagents and chemicals were analytical grade and obtained from commercial sources.

*Sample collection:* A total of 100 chicken liver samples were obtained from Afyonkarahisar city, Turkey from September 2017-January 2018. Liver samples were purchased by supermarkets and local markets. The transportation of samples to the laboratory was performed under cold conditions (+4°C) immediately after sampling and the collected samples were stored at -20°C in a deep freezer until the analysis.

*Standard solutions:* ENR and CIP stock solutions were prepared at a concentration of 1 mg/ml in methanol. The working solutions were prepared from stock solutions to generate a calibration curve and to perform recovery studies of the chicken liver.

Sample preparation: Each liver sample was homogenized by a food blender and 5 g of sample was transferred into a polypropylene centrifuge tube (50 ml) followed by addition of 10 ml methanol and then the mixture was vigorously shaken for 5 min. After the addition of 1% formic acid (4ml) and 0.01M EDTA (400  $\mu$ l), the centrifuge tube was vortexed for 3 and 2 min, respectively. Subsequently, centrifugation of the mixture was performed at 2000g for 10 min at 4°C. After centrifugation, the supernatant of each sample was filtered through a membrane filter (0.45  $\mu$ m) and the obtained filtrate was transferred to a High Performance Liquid Chromatography (HPLC) vial for injection.

Instrumental conditions: The LC method described here was developed using an LC system of Agilent Technologies 1200 series (Waldbronn, Germany), combined to a binary high-pressure gradient pump. LC separation was performed by using Agilent Zorbax Eclipse XDB C18 column (4.6 x 30 mm x 3.5 µm). Mobile phase A consisted of water containing 0.1% formic acid while mobile phase B consisted of methanol containing 0.1% formic acid. The flow rate and injection volume of the mobile phases, and column temperature were 0.4 ml/min, 10µl, and 40°C, respectively. LC separation gradient was set to A/B (80/20) at 0.0 min; it was changed to A/B (10/90) and kept unchanged until 4.10 min, A/B (80/20). MS analysis was conducted on Agilent 6460 LC/MS Triple Quadrupole instrument equipped with an ESI (Waldbronn, Germany) in positive ion mode. MS parameters of drying gas temperature, nebulizer gas, sheath gas flow, capillary voltage and sheath gas temperature were set to 350°C, 40 p.s.i., 10 l/min, 4000 V, and 400 °C, respectively. Precursor ion and productions of ENR were 360, 342, and 316, respectively while precursor ion and productions of CIP were 332, 231 and 314, respectively.

**Ouality parameters of method:** The validation of this method was performed by spiking chicken liver samples and limit of detection (LOD), the limit of quantification (LOQ), linearity range, intra- and inter-day precision and recovery were used as quality parameters. The concentrations of the samples were calculated according to the calibration curve method for which a series of external standard solutions are prepared and measured. For the calibration equation, a line is fitted to these data and the obtained equation is employed to calculate the concentration unknown samples. In addition, the strength of the linear regression is expressed with a correlation coefficient  $(r^2)$ . These parameters were calculated by the software (Agilent MassHunter Workstation Software Version 7) of the analytical instrument. The lowest concentration of ENR or CIP that the analytical process can reliably differentiate from background levels (signal-to-noise ratio≥3) and lowest concentration of ENR or CIP that can be quantified (signal-to-noise ratio  $\geq 10$ ) were defined as LOD and LOQ, respectively.

#### Results

ENR and CIP in chicken liver samples were quantified by LC-MS/MS. The typical chromatogram and standard calibration curves of ENR and CIP are shown in Figure 1 and Figure 2. LOD, LOQ, calibration curve parameters and retention times are shown in Table 1. Relative standard deviations used for precision and intraand inter-day recoveries used for the accuracy of the method are given in Tables 2 and 3. The results of the present study show that 39% of all chicken liver samples (n=100) were contaminated with ENR while 31% of

chicken liver samples contained CIP (Table 4). Additionally, the highest concentrations of ENR and CIP were determined to be 30.23 and  $6.92 \mu g/kg$ , respectively.



Figure 1. Chromatograms of enrofloxacin (A) and ciprofloxacin (B).



Figure 2. Standard calibration curves of enrofloxacin (A) and ciprofloxacin (B).

Table 1. Analytical parameters of enrofloxacin and ciprofloxacin in chicken livers.

Antibiotic	LOD (µg/kg)	LOQ (µg/kg)	<b>Correlation</b> <b>coefficient</b> (r <sup>2</sup> )	Calibration equation	Data point	Linear range (µg/kg)	RT (min)
Enrofloxacin	1.17	3.90	0.995	y=0.039058x +0.014185	5	1-20	2.57
Ciprofloxacin	1.24	4.14	0.998	y=0.018435x +7.606705	5	1-20	2.45

Enrofloxacin	Intra-day assays (n=10)		Inter-day assays (n=14)		
Spiked (µg/kg)	Percentage	RSD	Percentage	RSD	
	<b>Recovery</b> ±CV	(%)	<b>Recovery</b> ±CV	(%)	
40	97.86±1.87	1.91	99.30±5.23	5.27	
80	90.70±2.86	3.15	90.21±4.51	4.51	
160	92.87±2.55	2.75	90.71±2.38	2.62	

Table 2. Intra- and inter-day precisions for enrofloxacin in chicken liver samples.

Table 3. Intra- and inter-day precisions for ciprofloxacin in chicken liver samples.

Ciprofloxacin	Intra-day assays (n=10)		Inter-day assays (n=14)		
Spiked (µg/kg)	Percentage	centage RSD Percen		RSD	
	<b>Recovery</b> ±CV	(%)	<b>Recovery</b> ± <b>CV</b>	(%)	
40	93.78±3.07	3.28	94.48±4.66	4.94	
80	89.19±4.49	5.03	88.13±4.87	4.87	
160	94.00±2.44	2.6	91.47±1.63	1.78	

Table 4. The enrofloxacin and ciprofloxacin levels of chicken liver samples (n=100).

Enrofloxacin	Range (µg/kg)	<1.17	1.17-9.99	10-19.99	20-35
	Number of samples	61	30	6	3
Ciprofloxacin	Range (µg/kg)	<1.24	1.24-1.99	2-3.99	4-7
	Number of samples	69	13	11	7

<b>Table 5.</b> Previous studies investigating the	presence of quinolone-residues in chicken livers.

References	Country	Analyze	Samples size	Positive (%)	Range (µg/kg)	Method
Attari et al. (5)	Iran	ENR	20	17 (85%)	5.30-90.7	ELISA
Salehzadeh et al. (23)	Iran	ENR	90	90 (100%)	4.3-66.2	HPLC
Rezaee et al. (22)	Iran	ENR	50	26 (52%)	16.7-296.7	HPLC
		CIP		15 (30%)	9.8-93.3	
Aslam et al. (4)	Pakistan	ENR	75	59 (78.7%)	527 <sup>b</sup>	HPLC
Naeem et al. (19)	Pakistan	ENR	40	37 (92%)	3.02-364	HPLC
		CIP		34 (85%)	2.08-245	
Younus et al. (30)	Pakistan	ENR	45	40 (88.8%)	1.409 <sup>a</sup>	ELISA
Sattar et al. (26)	Bangladesh	ENR	50	20 (40%)	-	TLC
		CIP		22 (44%)	-	
Sarker et al. (25)	Bangladesh	ENR	160	57 (36%)	-	TLC
Faten et al. (9)	Egypt	CIP	15	13 (86.7%)	96.33-300.27	HPLC
Sultan (27)	Iraq	ENR	30	10 (33.31%)	0.01-10.69	ELISA
Nizamlıoğlu and Aydın (20)	Turkey	CIP, DAN, DIF, ENR, FLU, MAR, OA, SAR	50	17 (34%)	18.5-147.88	ELISA
Metli et al. (17)	Turkey	CIP, DAN, DIF, ENR, FLU, MAR, NA, NOR, OA, SAR	34	-	-	LC- MS/MS
This Study	Turkey	ENR	100	39 (39%)	1.42-30.23	LC- MS/MS
		CIP		31 (31%)	1.25-6.92	

<sup>a</sup>Mean concentration, Ciprofloxacin (CIP), Danofloxacin (DAN), Difloxacin (DIF), Enrofloxacin (ENR), Flumequine (FLU), Marbofloxacin (MAR), Nalidixic Acid (NA), Norfloxacin (NOR), Oxolinic Acid (OA), Sarafloxacin (SAR)

#### **Discussion and Conclusion**

Poultry meat and giblets, including liver, are widely consumed throughout the world. However, the liver is considered a highly risky poultry product due to its role in the detoxification of toxic substances, including antibiotics (2, 13, 15). Quinolones are powerful options among the antimicrobial agents employed in the treatment of various bacterial-caused poultry diseases (29). In addition, the presence of quinolones in poultry products causes public health concerns (21). Local and international regulatory and public health agencies give attention to screening of antibiotic residues in foodproducing animals. In Turkey, National Residue Monitoring Plan is established to determine the presence of these substances including quinolone residues in foods of animal origin (3). Several monitoring studies conducted in different countries to investigate the presence of quinolone residues in chicken liver based on various methods (Table 5). Some of them reported very high incidence for quinolone residues in Pakistan (4, 19, 30), Iran (5, 23) and Egypt (9). Attari et al. (5) reported that in 85 % of 20 chicken liver samples collected from the Northwestern part of Iran, the ENR residue level ranged from 5.30 to 90.70  $\mu$ g/kg, with a mean concentration of 39.54 µg/kg. In another study conducted in Iran using HPLC, a high contamination level (100%) was reported for 90 chicken liver samples in a range from 4.3 to 66.2 µg/kg (23). Similarly, Faten et al. (9) found, using HPLC, that 86.7% of 15 chicken liver samples marketed in Egypt tested positive for CIP at concentrations between 96.33 and  $300.27 \,\mu g/kg$ . Our results are lower than these studies. However, the findings of our study are in consistent with other studies for those incidence of quinolone-residues ranged from 33.31% to 44% (20, 26, 27). Sattar et al. (26) performed a monitoring study in Bangladesh based on thin-layer chromatography (TLC) and found contamination levels in chicken liver samples of 40% and 44% for ENR and CIP, respectively. In another study conducted in Iraq, ENR incidence reported as 33.31 % while ENR level ranged from 0.01  $\mu$ g/kg to 10.69  $\mu$ g/kg. Nizamlıoğlu and Aydın (20) evaluated a total of 50 chicken liver samples regarding the presence of quinolone-residues based on ELISA method and reported that 34 % samples contained quinolones. Additionally, in a study conducted in Antakya province of Turkey, chicken liver samples were collected from 34 different markets and analyzed regarding 38 antibiotic residues and none of the samples have quinolone residues (17). Our results are not compatible with this study. In the present study, the ENR contamination level was found to be between 1.42 and 30.23µg/kg (39%), and the CIP contamination level was found to be between 1.25-6.92  $\mu$ g/kg (31%). According to the results of the present study, the total sum of ENR and CIP did not exceed the established MRL in any of the chicken samples. Taken together, different incidences of

quinolone-residues may be due to misuse or overuse of these antibiotics or disrespect of withdrawal time during poultry breeding.

In conclusion, poultry meat and meat products are extensively consumed throughout the world owing to their nutritive value and suitable price. However, antibiotic residues in foods of animal origin are an important public health issue. Therefore, monitoring programs for veterinary drug residues in these types of foods have been performed by national authorities. This study contributes to those residue monitoring programs by providing a highly sensitive and accurate LC-MS/MS method for chicken liver samples.

#### **Financial Support**

The authors thanks to the Scientific Research Council of Afyon Kocatepe University due to financial support to this study (Project no: 17.C.140).

#### **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### References

- **1.** Acaroz U, Arslan-Acaroz D, Gurler Z (2016): *immunoassay development for the detection of antibiotic residues in foods.* Kocatepe Vet J, **9**, 122–126.
- 2. Acaroz U, Gurler Z, Kara R, et al (2018): Presence of Salmonella spp. in chicken meat and giblets marketed in Afyonkarahisar province. Kocatepe Vet J, 11, 1–5.
- Anonymus (2019): Kalıntı izleme planı. Available at https://www.tarimorman.gov.tr/Konular/ Gida-Ve-Yem-Hizmetleri/Gida-Hizmetleri/Kalinti-Izleme. (Accessed Sept 09, 2019).
- Aslam B, Kousar N, Javed I, et al (2016): Determination of enrofloxacin residues in commercial broilers using high performance liquid chromatography. Int J Food Prop, 19, 2463–2470.
- Attari VE, Abbasi MM, Abedimanesh N, et al (2014): Investigation of enrofloxacin and chloramphenicol residues in broiler chickens carcasses collected from local markets of Tabriz, Northwestern Iran. Heal Promot Perspect, 4, 151– 157.
- 6. Bonassa KPD, Miragliotta MY, Simas RC, et al (2017): Tissue depletion study of enrofloxacin and its metabolite ciprofloxacin in broiler chickens after oral administration of a new veterinary pharmaceutical formulation containing enrofloxacin. Food Chem Toxicol, **105**, 8-13.
- 7. EU COMMISSION REGULATION (2010): No 37/2010 of 22 December 2009 on pharmacologically active substances and their classification regarding maximum residue limits in foodstuffs of animal origin.
- 8. Fàbrega A, Sánchez-Céspedes J, Soto S, et al (2008): *Quinolone resistance in the food chain.* Int J Antimicrob Agents, **31**, 307-315.
- **9.** Faten SH, Mousa MM, Mahomud AH, et al (2016): *Ciprofloxacin residues in chicken and turkey carcasses*. Benha Vet Med J, **31**, 136-143.

- **10.** Gbylik-Sikorska M, Posyniak A, Sniegocki T, et al (2016): Influence of enrofloxacin traces in drinking water to doxycycline tissue pharmacokinetics in healthy and infected by Mycoplasma gallisepticum broiler chickens. Food Chem Toxicol, **90**, 123-129.
- 11. Kaufmann A, Butcher P, Maden K (2012): Determination of aminoglycoside residues by liquid chromatography and tandem mass spectrometry in a variety of matrices. Anal Chim Acta, **711**, 46-53.
- Kirbis A, Marinsek J, Flajs VC (2005): Introduction of the HPLC method for the determination of quinolone residues in various muscle tissues. Biomed Chromatogr, 19, 259-265.
- Kirrella GAK, Deeb AMM, Abdallah RMI (2017): Safety of frozen liver for human consumption. J Food Drug Anal, 25, 520-524.
- 14. Lolo M, Pedreira S, Miranda JM, et al (2006): Effect of cooking on enrofloxacin residues in chicken tissue. Food Addit Contam, 23, 988–993.
- Magdelaine P, Spiess MP, Valceschini E (2008): Poultry meat consumption trends in Europe. Worlds Poult Sci J, 64, 53–63.
- **16.** Martins MT, Barreto F, Hoff RB, et al (2015): Determination of quinolones and fluoroquinolones, tetracyclines and sulfonamides in bovine, swine and poultry liver using LC-MS/MS. Food Addit Contam - Part A Chem Anal Control Expo Risk Assess, **32**, 333–341.
- 17. Metli M, Yakar Y, Tekeli Y (2015): Determination of antibiotic residues in chicken liver by liquid chromatography-tandem mass spectrometry. Adıyaman Uni J Sci, 5, 120-131.
- Moema D, Nindi MM, Dube S (2012): Development of a dispersive liquid-liquid microextraction method for the determination of fluoroquinolones in chicken liver by high performance liquid chromatography. Anal Chim Acta, 730, 80-86.
- **19.** Nacem M, Khan K, Rafiq S (2006): Determination of residues of quinolones in poultry products by high pressure liquid chromatography. J Appl Sci, **6**, 373-379.

- 20. Nizamlioğlu F, Aydın H (2012): Quinolone antibiotic residues in raw milk and chicken liver in Konya. Eurasian J Vet Sci, 28, 154-158.
- 21. Panzenhagen PHN, Aguiar WS, Gouvêa R, et al (2016): Investigation of enrofloxacin residues in broiler tissues using ELISA and LC-MS/MS. Food Addit Contam - Part A Chem Anal Control Expo Risk Assess, 33, 639–643.
- Rezaee N, Rafie S, Javadi A, et al (2018): Determination of enrofloxacin and ciprofloxacin residues in five different kinds of chicken tissues by dispersive liquid – liquid microextraction coupled with hplc. Iran J Pharm Res, 17, 1182–1190.
- 23. Salehzadeh F, Salehzadeh A, Rokni N, et al (2007): Enrofloxacin residue in chicken tissues from Tehran slaughterhouses in Iran. Pakistan J Nutr, 6, 409-413.
- 24. San Martin B, Cornejo J, Lapierre L, et al (2010): Withdrawal time of four pharmaceutical formulations of enrofloxacin in poultry according to different maximum residues limits. J Vet Pharmacol Ther, 33, 246–251.
- **25.** Sarker YA, Hasan MM, Paul TK, et al (2018): Screening of antibiotic residues in chicken meat in Bangladesh by thin layer chromatography. J Adv Vet Anim Res, **5**, 140.
- **26.** Sattar S, Hassan MM, Islam SKMA, et al (2014): Antibiotic residues in broiler and layer meat in Chittagong district of Bangladesh. Vet World, **7**, 738-743.
- **27.** Sultan IA (2014): Detection of Enrofloxacin Residue in Livers of Livestock Animals Obtained from a Slaughterhouse in Mosul City. J Vet Sci Technol, 5, 2-4.
- Toldrá F, Reig M (2006): Methods for rapid detection of chemical and veterinary drug residues in animal foods. Trends Food Sci Technol, 17, 482–489.
- **29.** Trouchon T, Lefebvre S (2017): A Review of enrofloxacin for veterinary use. Open J Vet Med, 6, 40–58.
- 30. Younus M, Abbas T, Zafar M, et al (2017): Quantification of enrofloxacin residues in broiler chicken tissues using competitive enzyme-linked immunosorbent assay. Indian J Anim Sci, 87, 659–661.

Ankara Üniv Vet Fak Derg, **67**, 143-152, 2020 DOI: 10.33988/auvfd.570790

### Honey price estimation for the future in Turkey; example of 2019-2020

#### Mustafa Bahadır ÇEVRİMLİ<sup>1,a,⊠</sup>, Mehmet Saltuk ARIKAN<sup>2,b</sup>, Mustafa Agah TEKİNDAL<sup>3,c</sup>

<sup>1</sup>Selçuk University, Faculty of Veterinary Medicine, Department of Livestock Economics and Management, Konya; <sup>2</sup>Fırat University, Faculty of Veterinary Medicine, Department of Animal Health Economics and Management, Elazığ; <sup>3</sup>Selçuk University, Faculty of Veterinary Medicine, Department of Biostatistic, Konya, Turkey.

<sup>a</sup>ORCID: 0000-0001-5888-242X; <sup>b</sup>ORCID: 0000-0003-4862-1706; <sup>c</sup>ORCID: 0000-0002-4060-7048.

<sup>⊠</sup>Corresponding author: bahadir.cevrimli@gmail.com Received date: 27.05.2019 - Accepted date: 01.11.2019

**Abstract:** The purpose of the present study is to develop models for the time series data of honey prices in Turkey between 1998 and 2018 using Box-Jenkins and Winter's Exponential Smoothing methods and to evaluate the TRY- and US\$-denominated forecasts made using such models for 2019-2020. The method employed to analyse the time series data is the Seasonal Auto Regressive Integrated Moving Average (SARIMA). The most suitable model to forecast the honey prices was found to be SARIMA (2,1,2)(1,0,1) for TRY-denominated prices and SARIMA (0,1,3)(0,0,0) for US\$-denominated prices. It was forecast that the average price of 1 kg of honey would be 49.08 TRY/6.15 US\$ at the end of 2020. Since the study will provide beekeepers with information on future prices, it will help them better plan their production activities. The price forecasts in this study will be useful for producers and those involved in sale of honey with respect to prediction of how the honey prices will change in the years to come.

Keywords: Forecasting, honey, time series, Turkey.

#### Türkiye'de bal fiyatlarının geleceğe yönelik kestirimi; 2019-2020 örneği

Özet: Bu çalışmada, Türkiye'de 1998-2018 yıllarına ait aylık bal fiyatları zaman serisinin Box-Jenkins ve Winter's Üstel Düzgünleştirme yöntemleri ile modellenerek ve bu modellerin 2019-2020 yılları arasında TL ve dolar bazında öngörülerinin değerlendirilmesi amaçlanmıştır. Zaman serilerinin analizlerinde mevsimsel otoregresif hareketli ortalama (SARIMA- Seasonal Autoregressive Integrated Moving Average) yöntemi kullanılmıştır. Bal fiyatları için belirlenen en uygun tahmin modeli TL bazında SARIMA(2,1,2)(1,0,1) modeli ve dolar bazında SARIMA(0,1,3)(0,0,0) modeli olduğu tespit edilmiştir. Çalışma sonucunda 2020 yılı sonunda bal fiyatlarının doğrusal artış ile ortalama 49.08 TL/kg ve 6.15 US\$ olacağı tahmin edilmiştir. Sonuç olarak arıcılıkla uğraşanların gelecekteki fiyatlar hakkında bilgi sahibi olmaları onlara üretim faaliyetinin planlanmasında faydalı bilgiler sağlayacaktır. Çalışmada yapılan fiyat tahminleri balın satış aşamasında faaliyet gösteren kişiler ve üreticiler için ürün fiyatlarının nasıl şekilleneceği konusunda öngörü sahibi olmalarını sağlayacaktır.

Anahtar sözcükler: Bal, tahminleme, Türkiye, zaman serisi.

#### Introduction

Today, the beekeeping industry has become a significant rural production sector with its increasing economic returns and product diversity (11). Turkey ranks second following China in both the total number of hives and total honey production (12). In 2018, Turkey produced 107 thousand tons of honey with a total of 8.1 million honey-producing hives (35).

While the marketing infrastructure of honey in Turkey is conventional, the marketing channels are divided into three groups, namely, producers, wholesalers and retailers (10). In addition to the marketing channels, the beekeeping enterprises are able to find markets for their products through direct sales to processing companies and via cooperatives. However, marketing via cooperatives is a last resort for producers due to its low margin of profit (25). The major marketing-related problems of beekeeping enterprises are that they cannot sell their products in a timely manner and cannot charge the price they prefer, and that a quality-price relationship does not exist for honey in the domestic market (11, 29).

Previous studies found that honey production costs varied depending on the total number of hives belonging to the enterprises (7, 22), and that the enterprises could reduce their production costs through sale of by-products and migratory beekeeping (11). In 2018, the average retail

price of strained honey in Turkey was 30.48 TRY (Turkish Lira)/kg (minimum-maximum: 28.22-33.72, standard deviation: 2.10) or 6.43 US\$/kg (minimum-maximum: 5.0-7.5, standard deviation: 0.82) (36).

The future of honey production and prices is important for both producers and consumers. Honey consumption per capita in Turkey was 0.84 kg in 2003 and rose to 1.22 kg as of 2013, the most recent year for which latest data are available (13). It is noted that honey price is the most important factor in the countries' honey export and competitiveness in the international market. Taking this into account, estimations and inferences are made regarding the competitiveness of countries in the international market using the past price data (20). Today, honey price affects the consumer demand for honey, along with a number of factors such as quality, production technique and origin of the honey (19, 21).

Time series is a series of observations made at certain time intervals, which allows development of an appropriate model using statistical methods to make forecasts (2). Time series analysis has been recently found wide use in studies focusing on honey production, specifically to predict the number of colonies and honey export and import (23, 28), to determine the competitive power in honey export (34), to forecast honey production using Autoregressive integrated moving average (ARIMA) models (6, 9, 18) and to determine the supply of and demand for honey (26).

The main objective of our study is to examine the fluctuations in TRY- and USD-denominated honey prices. Furthermore, we aim to develop models for the time series data of honey prices without taking into account the exchange rate parity and to make price forecasts. What we intend to do is to determine and predict the fluctuations in TRY- and USD-denominated prices.

The purpose of the present study is to use known models for the time series data of honey prices in Turkey between 1998 and 2018 using Box-Jenkins and Winter's Exponential Smoothing methods and to evaluate the TRY- and US\$-denominated forecasts made using such models for 2019-2020.

#### **Material and Methods**

**Data set:** The study consisted of data sets retail honey prices on a monthly basis belonging to the 1998-2018 year in Turkey (36). In the analysis of the time series, the predictions obtained by using the seasonal autoregressive moving average (SARIMA) method were evaluated. For analysis of time series, the SPSS program version 25.0 was used (15).

*The Box-Jenkins method (ARIMA):* Autoregressive Integrated Moving Average (ARIMA) method used in forecasting time series events was developed by Box and Jenkins (5). ARIMA modeling approach is limited to the assumption that there is linearity between the variables. Apart from that, the researchers developed alternative modeling perspectives for forecasting the time series events not fulfilling the linearity assumption.

ARIMA or Box-Jenkins models are the combinations of AR and MA models administered to the series differenced at degree d. The essence of the Box-Jenkins method is the choice of an ARIMA model that is the most suitable one among various models based on the structure of the current data but contains limited number of parameters. As a whole, these models are represented as ARIMA (p, d, q).

In the models (38),

p: degree of autoregressive model, q: order of moving average model, d: degree of non-seasonal differencing.

The expression of ARIMA (p, d, q) model can be defined as indicated in equation 1

$$Z_{t} = \phi_{1}Z_{t-1} + \phi_{2}Z_{t-2} + \dots + \phi_{P}Z_{t-P} + a_{t} - \theta_{1}a_{t-1} - a_{t} - \theta_{2}a_{t-2} - \dots - a_{q} - \theta_{q}a_{t-q}$$
[1]

Here:

 $\phi_P$ : parameter values for autoregressive operator,  $a_t$ : error term coefficients,  $\theta_q$ : parameter values for moving average operator,  $Z_t$ : time series of the original series differenced at degree d. In other words,

$$b_P \quad W_t = Y_t - Y_{t-1}, t = 1, 2, \dots, t$$
 [2]

The first differences series is defined as given in equation 2. Here:

 $W_t$  = The first differences series,  $Y_t$  = the random variables subset of the original time series. If the first differences series is not stationary, stationary is checked by differencing the first time series again. This is modeled as given in equation 3.

$$Z_t = W_t W_{t-1} \ t = 1, 2, \dots, t$$
[3]

When the degree of difference is d = 0 (that means that the original series is stationary), the ARIMA model will be AR, MA, or ARMA model. Due to this feature, it can be said that ARIMA models incorporate all of the Box Jenkins models.

As done by ARIMA (p, d, q) model, seasonal ARIMA(P,D,Q)s models only distinguish the interval between the observations that effect one another. These are (seasonal) periods. They do not indicate period involving a single interval.

For example: ARIMA(1,0,0)12 model is [monthly AR(1)]].

$$Z_t = \alpha + \phi_1 Z_t - 12 + A_t$$
 [4]

Thus, autocorrelation decrease exponential lags may be at 12, 24, 36, etc.

Partial autocorrelation lag is  $12 = \phi_1$ 

If lag is after 1, it becomes equal to 0.

Seasonal ARIMA(P,D,Q)s Models ARIMA(p, d, q) Models relationship can be expressed as SARIMA models:

$$\Phi P(B) \ \phi \ p(B) \ [1-B^s]^D \ [1-B]^d \ Z_t = \alpha + \Theta Q(B) \ \theta q(B) \ A_t$$
[5]

This model is used if we want to understand what is meant by autocorrelation design through two autoregressive polynomials and two moving average polynomials, and this can be referred to as difference operator as well (27). The model establishment process involves certain repetitive steps (5). These steps are indicated in the flow chart given in Figure 1.

The establishment of Box-Jenkins ARIMA models involves four main steps. In the first step, the class of the general model is determined. In selecting the general model, the graphs of autocorrelation and partial autocorrelation functions are used. The features of theoretical functions concerning ARIMA models are used based on autocorrelation and partial autocorrelation functions in Figure 1 (5). In the second step, a transient model compliant with the structure of the data is determined. To this end, autocorrelation and partial correlation functions are used. In determining the model, a model is selected from model classes such as AR, MA, ARMA, ARIMA, and SARIMA (32). In the third step, the parameters of the transient model are forecasted by use of efficient statistical techniques, and the standard errors of coefficients are calculated to test whether or not they are significant. In the last stage, compliance of the model is checked for forecasting. To this end, the autocorrelation function of the model is examined by drawing the graph of the autocorrelation coefficients of the errors of the transient model that is assumed to be compliant. If this function displays a particular shape, it is concluded that errors are not random. This kind of finding means that the determined transient model is not compliant. Therefore, one turns to the second step again, and this process is repeated until the compliant model is determined through a new transient model. The model passing the compliance check is now ready to be used for forecasting (3, 17, 31, 39).

Moving averages and exponential smoothing techniques were used in forecasting. Simple exponential smoothing is a method derived from the moving averages technique, and its equation is as follows:

$$Z_{t} = \alpha Z_{t} + (1 - \alpha) Z_{t-1}$$

$$\hat{Z}_{t+1} = Z_{t}$$
[6]

where  $Z_{t+1}$  is the forecast value for the next period,  $\alpha$  is the smoothing factor in the range of  $0 < \alpha < 1$ ,  $Z_t$  is the actual value of the new observation at time t, and  $Z_t$  is the previous smoothed value. Note that  $\alpha$  is set such that it minimises the mean squared errors.

t is the value observed at  $Y_t$ . Then, t is a seasonal component,  $b_t$  is the smoothing components of the trend t, L is the number of periods in a season,  $F_{t+m}$  is one forecast ahead of m periods, m is the number of forecast periods,  $\alpha$  (Alpha) smoothes the parameter,  $\beta$  (Beta) is the seasonal smoothing parameter, and  $\gamma$  (Gamma) is the smoothing parameter of trend (8).



Figure 1. Model establishment process.

- Forecast Error (e):  $\mathbf{e}_t = \mathbf{A}_t \mathbf{F}_t$  [7] • Mean Forecast Error (MFE or Bias):  $MFE = \frac{\sum_{i=1}^{n} (e_i)}{n}$
- ٠

• Mean Absolute Deviation (MAD): 
$$MAD = \frac{\sum_{i=1}^{n} |e_i|}{n}$$
[9]

• Mean Absolute Percentage Error (MAPE):  $M = \frac{1}{n} \sum_{t=1}^{n} \left| \frac{A_t - F_t}{A_t} \right|$ [10]

• Mean Square Error (MSE): 
$$MSE = \frac{\sum_{t=1}^{N} E_t^2}{n}$$
 [11]

#### Results

A time series analysis of the honey prices (in TRY and US\$) between 1998 and 2018 was conducted to forecast the future prices of honey. Time series graph of honey prices is given in Figure 2.

Figure 2 shows that the series of TRY prices has an increasing trend, along with some fluctuations. Seasonal effects and trends in the time series prevent the series from

40

being stationary. Autocorrelation function (ACF) and partial autocorrelation function (PACF) graphs are presented in Figure-3 for understanding the stability in the series. It is understood that the series is not stationary because there are more than one delays outside the confidence limits. In order for the series to be stationary and to decrease the difference between the values, the logarithm of the series was taken and the difference was continued until the series was stationary. It is concluded that after the difference for the trend, the series becomes stationary. The time series graph of honey prices obtained after taking its first difference is given in Figure 4, and the ACF and PACF graphs are given in Figure 5.

When the ACF and PACF graphs are examined in Figure 5, it is seen that the two delays exceed the confidence limit and the other two delays are close to the confidence limit. It can be said that the series has become static. In addition, the stability analysis of the series was performed by Augmented Dickey Fuller (ADF) unit root test (14). According to the ADF test result, the series was not stationary (t = -0,258; P = 0.111) before the difference was obtained, whereas after the difference-taking process the series became stable (t = -14,789; P = 0.001).

Several different models were tested and the most suitable model was found to be SARIMA (2,1,2) (1,0,1) for TRY-denominated prices and SARIMA (0,1,3) (0,0,0) for US\$-denominated prices.

Honey Price TRY

Honey Price US\$

30 RY-US\$/Kg 20 10 0 **JAN 1998** NOV 1998 SEP 1999 Ē MAY 200 SEP 2004 Ē MAY 200 MAR 2007 NOV 2008 SEP 2009 MAY 201 MAY 201 MAR 2017 MAR 2003 JAN 2003 NOV 200; **JAN 2008** JUL 2010 MAR 201: NOV 201: SEP 2014 JUL 2015 NOV 2018 JAN 201: JAN 2018 2000 2005 Date

Figure 2. Time series graph of honey prices (TRY-US\$).

[8]

146



Figure 3. Autocorrelation function (ACF) and partial autocorrelation function (PACF) graph of honey series.



Transforms: natural logarithm, difference(1)

Figure 4. Graph of time series taken from the difference of honey prices.


Figure 5. Autocorrelation function (ACF) and partial autocorrelation function (PACF) graph of the difference series of honey prices.

Table 1 illustrates the forecast values of honey prices until 2020. Honey prices (TRY/kg) are expected to follow an increasing trend between 2019 and 2020, similarly to that in the previous years. Accordingly, honey price is forecast to range between 30.07 and 75.81 TRY/kg as of December 2020 with an increase of 44% (average forecast value 49.08 TRY/kg). As is seen in Figure 6 where the results of the model SARIMA(2,1,2)(1,0,1) are shown, honey prices have been in a drastic increasing trend after 2014. The graph of honey prices in US\$ indicates that the prices have a constant trend similar to the previous years. Results of the model SARIMA(0,1,3)(0,0,0) indicate that honey prices peaked in 2009 after which they followed a decreasing trend (Figure 6). The average value of honey prices as of December 2020 was forecast to be 6.15 US\$/kg (3.87-8.79 US\$).

The goodness of fit of the obtained models is examined in comparison with a criterion known as  $R^2$ (determination coefficient). The values are between 0-1 and the values are close to 1. The stationary  $R^2$  is a measure that compares the stationary part of the model with the basic model. It is preferred in case of trend or seasonal structure in the series. RMSE is the square root of the mean squares error. It refers to how different the model is from the level estimated by the model in the dependent series. Smaller values indicate better model estimates. MAPE is the mean absolute percentage error. It is independent of the units of the series and can therefore be used to compare different series. MAE stands for mean absolute error and is expressed in the original unit of the series. MaxAPE is maximum absolute percentage error. It shows the maximum error among the forecast values and is expressed as a percentage. Therefore, it is independent of the unit used. It is useful for imagining a worst-case scenario for the forecasts. MaxAE measures maximum absolute error, expressed in the same unit as that of the dependent series. Normalized BIC (Bayesian information criteria) is a general measure of the overall fit of a model. It is used to make a comparison between different models for a series, with the low values suggesting a better model (1).

From Table 2, it is clear that the Box-Jenkins models developed for honey prices in TRY and US\$ are statistically significant (P=0.045 and P=0.001). MAPE value indicates that the series have quite usable forecasts.

Date month/year	Predicted honey price (TRY/kg) SARIMA (2,1,2) (1,0,1)	Lower confidence limit	Upper confidence limit	Predicted honey price (US\$/kg) SARIMA (0,1,3)(0,0,0)	Lower confidence limit	Upper confidence limit
Jan 2019	34.07	32.12	36.10	6.17	5.55	6.81
Feb 2019	34.43	31.91	37.10	6.08	5.22	6.99
Mar 2019	34.91	31.60	38.47	6.10	5.04	7.22
Apr 2019	35.35	31.50	39.53	6.10	4.95	7.33
May 2019	35.79	31.28	40.78	6.10	4.87	7.43
Jun 2019	36.29	31.22	41.95	6.10	4.79	7.53
Jul 2019	36.96	31.22	43.46	6.11	4.72	7.62
Aug 2019	37.82	31.43	45.13	6.11	4.65	7.71
Sep 2019	38.64	31.54	46.86	6.11	4.58	7.79
Oct 2019	39.33	31.58	48.42	6.11	4.52	7.87
Nov 2019	39.88	31.44	49.88	6.12	4.46	7.95
Dec 2019	40.42	31.34	51.34	6.12	4.41	8.03
Jan 2020	40.88	31.10	52.78	6.12	4.36	8.10
Feb 2020	41.47	30.98	54.38	6.12	4.30	8.17
Mar 2020	41.97	30.76	55.95	6.13	4.25	8.24
Apr 2020	42.62	30.66	57.73	6.13	4.21	8.3
May 2020	43.12	30.43	59.38	6.13	4.16	8.37
Jun 2020	43.83	30.34	61.33	6.13	4.12	8.43
Jul 2020	44.62	30.28	63.47	6.13	4.07	8.50
Aug 2020	45.74	30.43	66.14	6.14	4.03	8.56
Sep 2020	46.73	30.47	68.70	6.14	3.99	8.62
Oct 2020	47.66	30.44	71.21	6.14	3.95	8.68
Nov 2020	48.34	30.24	73.43	6.14	3.91	8.74
Dec 2020	49.08	30.07	75.81	6.15	3.87	8.79

Table 1. Monthly honey prices forecast values according to Box Jenkins models

\*The calculations made in Turkish Lira (TRY) were converted into US\$ using the exchange rate in the relevant period (Avarage exchange rate for 2018: US\$1= ±4.74).



**Figure 6.** The price of honey Box Jenkins model forecast chart.

Model		Ljung-Box Q(18)									
	Stationary R- squared	R-squared	RMSE	MAPE	MAE	MaxAPE	MaxAE	Normalized BIC	Statistics	DF	Р
Honey price TRY SARIMA (2,1,2)(1,0,1)	0.244	0.997	0.392	1.751	0.191	22.392	3.518	-1.741	10.582	12	0.045
Honey price US\$ SARIMA (0,1,3)(0,0,0)	0.040	0.979	0.357	3.690	0.252	21.128	1.971	-2.041	45.487	17	0.001

Table 2. Box jenkins model fit criteria

#### **Discussion and Conclusion**

In Turkey, the production of apicultural products is one of the branches of production that has a high competitiveness among animal products (24). In order for businesses in this sector to plan their production and manage their risks, they need to have knowledge of apicultural practices as well as forecasts for the future of the sector. The models developed by statistical methods enable such forecasts. Since the data are a series of observations made at certain time intervals, they are modelled by time series analysis (16).

A study forecasting the supply of and demand for honey in Turkey using the Box-Jenkins model predicted that the supply of and demand for honey would increase from 1.54 kg/person to 1.40 kg/person in 2020 and 2023, respectively (26).

Using the time series analysis, it was forecast that honey production would increase to 115 thousand tons and the number of colonies would reach 10 million in 2020 (28). Using the ARIMA model, it was forecast that in 2020 the honey production, number of hives and beeswax production would reach 14.6 thousand tons, 1,226 thousand and 552.9 tons, respectively (18).

In a study conducted to forecast honey production in Turkey and to identify the type of model that gives the most successful results, the most suitable forecast model was found to be ARIMA(0,1,1), and using this model it was forecast that honey production in Turkey would constantly increase, reaching 106.410 tons in 2019 and 107.887 tons in 2020 (9). In a study conducted to forecast honey production using the ARIMA model and time series data, the most suitable forecast model was found to be ARIMA(0,1,1), and using this model it was forecast that honey production in Turkey would constantly increase between 2017 and 2023, reaching 121.216 tons in 2023 (6). Another study conducted in the Czech Republic predicted using the ARIMA model that the number of honey-producing hives would be 529.730, the number of beekeepers 47.373 and the price of honey 132 CZK/kg in 2011-2012 (30).

The present study used the honey prices in TRY and US\$ between 1998 and 2018. Honey prices until 2020 were forecast using the SARIMA models selected in the study. According to the results of the analysis, average honey prices in Turkey were forecast to reach 37.82 TRY/kg and 6.11 US\$/kg in August 2019 and further increase to 49.08 TRY/kg ve 6.15 US\$/kg in December 2020 (Table 1).

Among the model fit criteria in Table 2, MAPE value suggests that the forecast model developed for honey prices has a percentage error of 1.751% for prices in TRY and 3.690% for prices in US\$. The main reason for the difference between TRY and dollar MAPE value is that TRY is depreciating against the dollar. Forecasts with an MAPE value below 10% are considered good forecasts (33). So, it may be said that the forecast values obtained from both models developed for honey prices in TRY and US\$ have a sufficient level of accuracy.

The top three exporters of honey in the international market are China, Argentina and Mexico. The main reason why they are leaders in this market is that they export at a price level lower than the average world price. While average honey export price in the world was \$1.9/kg in 2006, the export prices of China, Argentina and Mexico were \$1.3/kg, \$1.4/kg and \$1.9/kg, respectively (20). In the same year, Turkey exported honey at an average price of \$2.3/kg. The price of honey exported by Turkey in 2016 is reported to be \$4.11/kg (13).

Although the major factor affecting honey export is reported to be its price (20), increasing concerns, including over food reliability and residues, currently cause trade restrictions to be imposed on many countries in international trade. China is one of the major exporters on which restrictions are imposed (37). The restrictions on China are mostly imposed by European Union countries (20). Additionally, it is noted that consumers are willing to pay higher prices for quality honey (19). Particularly in EU countries, the origin, value and production technique of honey are reported to be the major factors affecting the consumers' decision to purchase honey (4, 21). Although Turkey seems to have disadvantages in honey export from the perspective of honey price compared to China, Argentina and Mexico, it may gain an advantageous position, particularly in the EU market, as the honey produced in Turkey is of higher quality and the price forecasts in this study suggest that the USD-denominated price of Turkish honey may remain constant.

In conclusion, the forward forecasting of honey prices using time series data and the sharing of the up-todate data with producers and producer associations at certain intervals will make a significant contribution to the development of a production and marketing strategy in Turkey, which is a leading country in the beekeeping sector. With timely and accurate price forecasts, producers can have the opportunity to develop their migratory beekeeping plans more precisely one year in advance. Additionally, they can increase their total sale revenues by developing a strategy regarding how much of the honey produced they will market via which marketing channel. If the beekeeping registration system is improved, price forecasts for various types of honey as well as other beekeeping products such as pollen and propolis can provide producers with motivating information for the next year with regard to which nectar flow they will go for and what kind of a migratory beekeeping route they will follow. Using the forecasts made with producer associations and public authorities, producers can be prompted to develop a production pattern and plan that is aimed at first meeting the domestic demand and then increasing export of high-quality beekeeping products.

#### **Financial Support**

This research received no grant from any funding agency/sector.

#### **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### References

- 1. Akaike HA (1974): A new look at the statistical model identification. 215-222. In: Selected Papers of Hirotugu Akaike, Springer, New York.
- **2. Akdi Y** (2012): Zaman Serileri Analizi (Birim Kökler ve Kointegrasyon). Gazi Kitabevi, 2. Baskı, Ankara.
- **3.** Arıkan MS, Çevrimli MB, Mat B, et al (2018): Price forecast for farmed and captured trout using box-jenkins method and 2009-2017. 79-90. In: Academic Studies in Health Sciences. Gece Publishing, Ankara.
- 4. Batt PJ, Liu A (2012): Consumer behaviour towards honey products in Western Australia. BFJ, 114, 285-297.
- **5.** Box GEP, Jenkins GM, Reinsel GC, et al (2015): Time series analysis: forecasting and control. 5th Edition, John Wiley & Sons, USA.
- Burucu V, Gülse Bal HS (2017): Türkiye'de arıcılığın mevcut durumu ve bal üretim öngörüsü. Tar Eko Der, 3, 28-37.

- 7. Cejvanovic F, Grgic Z, Maksimovic A, et al (2017): Assumptions of economic model for sustainable productions of beekeeping in the Bosnia and Hercegovina. JASTA, 5, 481-485, 2011.
- 8. Chatfield C, Yar M (1998): *Holt-Winters forecasting: some practical issues*. The Statistician, **37**, 129-140.
- **9.** Çelik § (2015): Modelling of honey production by using time series in Turkey. SAUJS, **19**, 377-382.
- Çevrimli MB, Sakarya E (2018): Türkiye arıcılık sektöründe mevcut durum, sorunlar ve çözüm önerileri. Erciyes Üniv Vet Fak Derg, 15, 58-67.
- Çevrimli MB, Sakarya E (2019): Economic analysis of beekeeping enterprises in Aegean Region, Turkey. Ankara Univ Vet Fak Derg, 66, 109-115.
- FAOSTAT (2018): Food and Agriculture Organization of the United Nations Statistics Division. Beekeping statictics. Available at http://faostat3.fao.org/download/Q/\*/E. (Accessed Sept 07, 2018).
- FAOSTAT (2019): Food and Agriculture Organization of the United Nations Statistics Division. Crops and Livestock Statictics. Available at http://faostat3.fao.org/download /Q/\*/E. (Accessed Aug 29, 2019)
- **14.** Gujarati DN (2009): Temel ekonometri. 6.Baskı, Literatür Yayıncılık, İstanbul.
- **15. IBM Corp. Released** (2013): IBM SPSS statistics for windows, Version 22.0. Armonk, NY: IBM Corp.
- **16. Kaymaz Ö** (2018): Forecasting of commercial egg production in Turkey with box-jenkins and winter's exponential smoothing methods. Eurasian J Vet Sci, **34**, 142-149.
- **17.** Kaymaz Ö, Tekindal MA, Ateş C, et al (2018): Power analysis and sample size determination in log-rank (lakatos) test. Biostat Biometrics Open Acc J, 6, 1-4.
- Koç B, Terin M, Ceylan M, et al (2010): General situation of beekeeping in the Eastern Anatolian Region of Turkey and ARIMA model with the help of long-term analysis. AJAVA, 5, 537-546.
- Kos Skubic M, Erjavec K, Klopčič M (2018): Consumer preferences regarding national and EU quality labels for cheese, ham and honey: The case of Slovenia. BFJ, 120, 650-664.
- Ma L (2009): International comparison of the export competitiveness of Chinese honey. Asian Agric Res, 1, 17-20.
- **21.** Menozzi D, Mora C, Chryssochoidis G, et al (2010): *Traceability, quality and food safety in consumer perception,* ECAG, **12**, 137-158.
- 22. Ören MN, Alemdar T, Parlakay O, et al (2011): Adana ilinde arıcılık faaliyetinin ekonomik analizi. Ankara: Tarımsal Ekonomi Araştırma Enstitüsü (TEAE) Publication number:178.
- 23. Parlakay O, Yılmaz H, Yaşar B, et al (2018): The situation of beekeeping in Turkey and the future expectations by the trend analysis method. Bursa Uludağ Üniv Ziraat Fak Derg, 22, 17-24.
- 24. RTMD (2013): Republic of Turkey Ministry of Development, 10<sup>th</sup> Development Plan 2014-2018. Special report on livestock commission, Ankara, Turkey. Available at http://www.sbb.gov.tr/wp-content/uploads/2018/10/10 \_Hayvancilik.pdf. (Accessed June 07, 2019)

- 25. Saner G, Engindeniz S, Çukur F et al (2005): İzmir ve Muğla illerinde faaliyet gösteren arıcılık işletmelerinin teknik ve ekonomik yapısı ile sorunların üzerine bir araştırma. T.C. Tarım ve Köyişleri Bakanlığı Tarımsal Ekonomi Araştırma Enstitüsü, Yayın No:126, Ankara.
- **26.** Saner G, Adanacioğlu H, Naseri Z (2018): Forecasting honey supply and demand in Turkey. TJAE, **24**, 43-51.
- 27. Schwert GW (1989): Tests for Unit Roots: A Monte Carlo Investigation. J Bus Econ Stat, 7, 147-160.
- **28.** Semerci A (2017): Overall situation of beekeeping in *Turkey and future prospects*. MKUTBD, **22**, 107-118.
- SERKA (2012): Serhat Kalkınma Ajansı. Ardahan Kafkas Arı Irkı ve Arıcılık Çalıştayı Sektör Raporu; 58-59 Kars, Türkiye.
- **30.** Šimpach O (2012): Statistical view of the current situation of beekeeping in the Czech Republic. In: The 6<sup>th</sup> International Days of Statistics and Economics, Sep 13-15, Czech Republic.
- **31.** Tekin ME, Tekindal MA (2019): Optimum sample size in group comparisons in animal breeding researches with simulation study. JSTR, 5, 279-286.
- **32.** Tekindal MA, Güllü Ö, Yazıcı AC, et al (2016): The modelling of time-series and the evaluation of forecasts for the future: the case of the number of persons per physician in Turkey between 1928 and 2010. Biomed Res, 27, 965-971.

- Temuçin T, Temiz İ (2016): Türkiye dış ticaret ihracat hacminin projeksiyonu: holt-winters ve box, jenkins modellerinin bir kıyaslaması. SDÜ İİBF Dergisi, 21, 937-960.
- **34.** Terin M, Yıldırım İ, Aksoy A, et al (2018): Competition power of Turkey's honey export and comparison with Balkan Countries. Bulg J Agric Sci, 24, 17-22.
- TURKSTAT (2019): Turkey Statistical Institute Livestock statistics database. Available at https://biruni.tuik.gov.tr/ hayvancili-kapp/hayvancilik.zul. (Accessed Jan 05, 2019).
- 36. TURKSTAT (2019): Turkey Statistical Institute. Agricultural price statistics database. Available at https://biruni.tuik.gov.tr/medas/?kn=110&locale=tr. (Accessed Feb 22, 2019).
- **37.** Wei GX, Huang JK, Jun Y (2012): Honey safety standards and its impacts on China's honey export. JIA, **11**, 684-693.
- 38. Wickramarachchi AR, Herath HMLK, Jayasinghe-Mudalige UK, et al (2017): An analysis of price behavior of major poultry products in Sri Lanka. JAS Sri Lanka, 12, 138-148.
- **39. Yenice S, Tekindal MA** (2015): Forecasting the stock indexes of fragile five countries through box-jenkins methods. IJBSS, **6**,180-191.

Ankara Üniv Vet Fak Derg, **67**, 153-160, 2020 DOI: 10.33988/auvfd.591296

## Knowledge and attitudes in food safety and the occurrence of indicator bacteria on hands of food handlers at the point of pastrami sale

#### Yeliz YILDIRIM<sup>1,a,⊠</sup>, Nurhan ERTAŞ ONMAZ<sup>1,b</sup>, Zafer GÖNÜLALAN<sup>1,c</sup>, Harun HIZLISOY<sup>2,d</sup>, Serhat AL<sup>1,e</sup>, Fulden KARADAL<sup>3,f</sup>, Erhan KUM<sup>4,g</sup>, Aytaç AKÇAY<sup>5,h</sup>

<sup>1</sup>Erciyes University, Faculty of Veterinary Medicine, Department of Food Hygiene and Technology, Kayseri; <sup>2</sup>Erciyes University, Faculty of Veterinary Medicine, Department of Veterinary Public Health, Kayseri; <sup>3</sup>Niğde University, Bor Vocational School, Niğde;
 <sup>4</sup>Municipality of Melikgazi, Directorate of Environmental Protection and Control, Kayseri; <sup>5</sup>Erciyes University, Faculty of Veterinary Medicine, Department of Biometry, Kayseri, Turkey

<sup>a</sup>ORCID: 0000-0001-8783-3889; <sup>b</sup>ORCID: 0000-0002-4679-6548; <sup>c</sup>ORCID: 0000-0002-3935-6296; <sup>d</sup>ORCID: 0000-0003-3391-0185; <sup>e</sup>ORCID:0000-0003-2721-9275; <sup>f</sup>ORCID: 0000-0002-5113-5883; <sup>g</sup>ORCID: 0000-0002-1704-5286; <sup>b</sup>ORCID: 0000-0001-6263-5181

Corresponding author: yyildirim@erciyes.edu.tr Received date: 12.07.2019 - Accepted date: 03.11.2019

**Abstract:** The aim this study was to evaluate food safety knowledge and attitudes of food handlers and to determine the presence of indicator bacteria on the hands of food handlers in the pastrami retail points in Kayseri, Turkey. Food handlers were interviewed and the samples were taken from their hands by touching the petri dishes. Food handlers' knowledge and attitudes were evaluated by questionnaires and checklists. Conventional methods were followed for microbiological analyses. The presence of total coliforms, *E. coli*, members of the family Enterobacteriaceae and *S. aureus* on the hands of food handlers were analyzed. Results indicated that coliforms were present in 37.3% of food handlers hands and 32% exceeded the limit when compared to the literature suggesting a target value of <2.5 cfu/cm<sup>2</sup>. *E. coli*, Enterobacteriaceae and *S. aureus* were present in 10.6%, 44.6% and 34% of handler's hands, respectively. Food safety knowledge evaluation received high scores with mean score of 69.06%. Only one handler was below the score of 50%. Despite the high scores on food safety knowledge, high numbers of hand samples were found to be contaminated with pathogens. Data obtained from this study reveal that despite their adequate food safety knowledge, food handlers on pastrami retail points were found insufficient for implementation. Therefore, it is not only sufficient to provide food safety trainings but also the sources of problems, such as lack of hygiene infrastructure, in the practice should be identified and resolved in retail points.

Keywords: Food handler, food safety knowledge, indicator bacteria, pastrami.

# Pastırma satış noktalarındaki gıda çalışanlarının gıda güvenliği bilgi ve tutumları ve ellerindeki indikatör bakteri varlığının belirlenmesi

Özet: Bu çalışmanın amacı, Kayseri'de bulunan pastırma satış noktalarındaki gıda çalışanlarının gıda güvenliği bilgi ve tutumlarını değerlendirmek ve ellerindeki indikatör bakterileri varlığını tespit etmektir. Gıda çalışanlarıyla görüşülmüş ve çalışanların ellerinden petrilere dokundurularak örnek alınmıştır. Gıda çalışanlarının gıda güvenliği bilgi ve tutumları, anketler ve kontrol listeleri ile değerlendirilmiştir. Mikrobiyolojik analizler için geleneksel yöntemler takip edilmiştir. Gıda çalışanlarının ellerinde total koliform, *E. coli*, Enterobacteriaceae türleri ve *S. aureus* analiz edilmiştir. Elde edilen sonuçlara göre gıda çalışanlarının %37,3'ünün el örneklerinde koliform bulunmuş ve %32'sinin literatürde koliformlar için limit değer olarak belirtilen <2,5 cfu/cm<sup>2</sup>'yi aştığı belirlenmiştir. *E. coli*, Enterobacteriaceae türleri ve *S. aureus*, gıda çalışanlarına ait el örneklerinin sırasıyla % 10,6, % 44,6 ve % 34'ünde bulunmuştur. Gıda güvenliği bilgi değerlendirme anket çalışması, ortalama % 69,06 skoru ve % 50 puanın altında tek gıda çalışanı ile yüksek puanlar almıştır. Gıda güvenliği bilgisine dair yüksek puanlara rağmen, gıda çalışanlarına ait el örneklerinin patojenlerle kontaminasyon düzeyleri oldukça yüksek bulunmuştur. Bu çalışmadan elde edilen veriler, Türkiye'de pastırma satış noktalarında çalışanların gıda güvenliği hakkında yeterli bilgiye sahip olmalarına rağmen uygulama noktasında yetersiz kaldıklarını ortaya koymaktadır. Dolayısıyla sadece gıda güvenliği eğitimi vermek yeterli olmamakta, satış yerlerindeki hijyen altyapı yetersizliği gibi uygulamadaki aksaklık nedenlerinin da belirlenerek çözülmesi de gerekmektedir.

Anahtar sözcükler: Gıda çalışanı, gıda güvenliği bilgisi, indikatör bakteri, pastırma.

154 Yeliz Yıldırım - Nurhan Ertaş Onmaz - Zafer Gönülalan - Harun Hızlısoy - Serhat Al - Fulden Karadal - Erhan Kum -Aytaç Akçay

#### Introduction

Pastrami is a raw, cured, edible paste coated and commonly consumed ready-to-eat (RTE) meat product in Turkey. It is often sliced now of sale by food handlers. RTE products that have been associated with several outbreaks are recognized to be contaminated during slicing (32).

Food borne diseases remain the significant public health problem although improvements in health systems and the increase in food hygiene training programs in developed countries. A report from the USA indicates annually 76 million food borne diseases with 5200 cases of mortality (6).

Food handlers should have the necessary knowledge and skills to enable them to apply hygiene practices that are obligatory in EU (27). According to EU regulation no: 852/2004 on the hygiene of food staff, it is required for food handlers to be trained on food hygiene according to their work activities. Besides, food business operators are required to ensure the hygiene requirements at all stages of food production (34). Hence, great emphasis on managers was underlined to provide food hygiene training program for food handlers and to monitor their performance at work place (19). Previous data on risk factors for food borne disease suggest that, most of the outbreaks occur through undesirable food handling practices of food handlers that cannot be easily overturned, even by the training programs (9,16,37,38).

Presences of indicator organisms are commonly used to evaluate the effectiveness of the sanitation programs (17,30). Some pathogens including *S. aureus* and *E. coli* have shown to survive on hands and surfaces even for days after the contact with the organism (20,22).

The safety of pastrami should be guaranteed in every stage of food chain to protect public health. Lack of knowledge or negligence of food handlers are predisposing factors in increasing prevalence of food borne disease. Assessment of food handler's knowledge and practices are essential to provide required data for further action that are previously studied by some other reporters (4,19,21,27,28,40). There is need to understand all factors contributing to poor hygiene practices at the point of sale. Therefore, this study is designed to evaluate the food handling attitudes and food safety knowledge of pastrami handlers and to highlight the importance of hand hygiene in pastrami contamination in Kayseri, Turkey.

#### **Materials and Methods**

This study was designed among pastrami handlers at retail points where pastrami is often sliced at moment of sale in Kayseri, Turkey. Total of 150 retail points located in city center were visited. Samples were collected from 150 food handler's forefingers and thumps during working hours without previous notification and were transported to laboratory in a cool box.

Rodac plates containing Baird Parker Agar (Merck 1.05406) with Egg-yolk Tellurite emulsion were used to sample forefingers for *S. aureus* enumeration. Agar plates were incubated at 35°C for 24-48 h and black colonies with clear zones around were evaluated after being confirmed with coagulase test (3). *S. aureus* ATCC 25923 was used as positive control.

Rodac plates containing Chromocult Coliform Agar (Merck 1.10426) were used to sample thumbs; to enumerate total coliforms, *E. coli* and members of Enterobacteriaceae family. Plates were incubated at 35-37 °C for 24 h and colonies were differentiated presumptively on the basis of their appearance and color. In this context, colonies salmon to red in color were reported as total coliforms, dark blue to violet as *E. coli* and colorless ones as members of Enterobacteriaceae family (5,11).

A checklist was formed to evaluate food hygiene level in the establishments. Fourteen food construct items were selected including food handlers attitudes (4 items) and retail store conditions (10 items). The checklist was aiming to determine the food safety attitude of food handlers depending on the observations of the interviewer. Additionally, two questionnaires, modified from previous studies, were designed to determine the food safety knowledge and demographic characteristics of food handlers (14,28,40). The emphasis was put on hygiene practices, personal hygiene, food borne disease and temperature control. Handlers with the correct answers lower than 10 for 16 questions ( $\geq 62.5\%$  accuracy) were considered to have "insufficient" knowledge and those with scores greater than 10 points were reported to have "Good" knowledge.

The population of the study consisted of 468 active markets and delicatessens operating in the central districts of Kayseri. Previsit observations revealed that almost half of the markets were retailing pastrami (N = 234). In the study, it was calculated that at least 145 retail points should be taken by using stratified sampling method according to districts with 95% confidence interval. The sample size of the study was determined as 150 pastrami retail points throughout Kayseri (41).

Calculation of sample size (n0) is given in the following equation;

$$n_0 = \frac{Nt^2 pq}{d^2(N-1) + t^2 pq} = \frac{234(1,96)^2 0.5 * 0.5}{0.05^2(234-1) + (1.96)^2 * 0.5 * 0.5} \cong 145$$

#### Representation power: 150/234=0,64

N = Population size; t-table value for t = 95% confidence interval = 1.96; p, q = The frequency of occurrence of the event of concern, agent being (+) and (-) p = 0.5, q = 0.5; d = Deviation according to the frequency of occurrence.

Microorganisms	n	No. of positive samples (%)	No. of positive samples exceeding the target value (cfu/cm <sup>2</sup> )
Enterobacteriaceae*		67 (44.6)	-
Coliforms	150	56 (37.3)	48 (32)≥2.5
E. coli	150	16 (10.6)	16 (10.6) >1
S. aureus*		51 (34.0)	-

Table 1. Hand hygiene profiles of food handlers.

n: Total number of hand samples from different pastrami store.

\*: No target value

#### **Results**

No target values were found in Turkish Food Codex for bacteriological counts on food handlers hands. Therefore, only coliforms and *E. coli* positive samples were compared to the literature target values of  $\geq 2.5$ cfu/cm<sup>2</sup> and  $\geq 1$  cfu/cm<sup>2</sup>, respectively. Table 1 summarizes the microbial contaminations on hands of pastrami food handlers.

When demographic characteristics of food handlers were considered, all food handlers (100%) analyzed were male. Approximately two third of the participants (65.2%) were aged between 25 and 45 years. Most of them were married (82.6%) and the education level of largest group (47.3%) was primary school. Two third of the participants had the experience more than 10 years on their work. The demographic characteristics of pastrami food handlers were presented in Table 2.

 Table 2. Demographic characteristics of pastrami food handlers.

Characteristics	Ν	%
Gender		
Male	150	100
Age		
17-25	14	9.3
25-35	46	30.6
35-45	52	34.6
45-55	24	16
>55	14	9.3
Marital status		
Married	124	82.6
Education		
Primary school	71	47.3
12th year	63	42
University	16	10.6
Experience (years)		
0-10	50	33.3
10-25	58	38.6
>25	42	28
Position		
Owner	60	40.1
Staff	90	59.9

N: Number of retail points.

#### Table 3. Food safety construct items.

	n	%
Food Handler Attitude		
1. Glove use	14	9.3
2. Apron use	72	48
3. Nail length		
Trimmed short	134	89.3
Long	16	10.6
4. Wearing jewelry	22	14.6
Retail store conditions		
1. Type of retail store		
Producer	4	2.6
Market	20	13.3
Franchise	126	84
2. Retail store age		
≥10	56	37.3
<10	94	62.6
3. Number of Employees		
>5	12	8
2-5	90	60
1	48	32
4. Floor material		
Tile	126	84
Other	24	16
5. Wall material		
Tile	80	53.3
Other	70	46.6
6. Product variety to be sailed		
Only pastrami	124	82,6
Market (All meat and meat products)	26	17,3
7. Cutting board material		
Wood	116	77.3
Acrylic	32	21.3
Other	2	1.3
8. Knife haft material		
Wood	136	90.6
Acrylic	12	8
Other	2	1.3
9. Cleaning agent		
Detergent	118	78.6
Bleach	4	2.6
Detergent + Bleach	20	13.3
Other	8	5.3
10.Pastrami storage conditions		
Refrigerator temperature	96	64
Room temperature	12	8
Both	42	28

#### 156 Yeliz Yıldırım - Nurhan Ertaş Onmaz - Zafer Gönülalan - Harun Hızlısoy - Serhat Al - Fulden Karadal - Erhan Kum -Aytaç Akçay

Table 4.	Food	safety	knowle	dge	of foc	d h	andlers	in	pastrami reta	il	points.
				- 0 -					F		r

Statement	Correct Answer (%)
1. Hand washing before handling reduces food contamination risk	100
2. Food borne infections could be reduced if the food handlers properly wash their hands where necessary	88
3. Disposable gloves should be worn before handling pastrami	96
4. Proper cleaning and sanitation of utensils reduces the food contamination risk	96
5. Food and beverage consumption by handlers in the work place increases the food contamination risk	44
6. Previous preparing of food prevents food contamination	36
7. Repetitive heating of cooked food can increase contamination risk	80
8. Foods must be stored at 5 °C	80
9. Periodical check of refrigerator temperatures could reduce the microbial growth in food	85
10. Freezing could kill all the bacteria in food	40
11. Contaminated food always changes in color, odor and taste	8
12. In case of infectious wounds on skin, take a break from work is necessary	92
13. Is it possible that your diarrheal infection threatens consumers health	44
14. Healthy people could also carry pathogen microorganisms on their skin, nose and mouth	84
15. Healthy adults and the people at risk group (children, pregnant woman, elderly) can be considered at equal risk for food borne disease	64
16. Food-borne diseases could be the cause of abortion in pregnant woman	68

The majority of food handlers were recorded to not to use gloves (90.7%) and not to wear (52%) aprons during handling pastrami. In addition, 10.6% of them had long nails and 14.6% wore jewelry. Checklist of food safety construct items (food handler attitude and retail store conditions) and their corresponding scores were summarized in Table 3. Within the scope of the current study, 60% of attendants declared that they received formal food hygiene training (Data not shown).

The average of correct answers for the questionnaire was 11, corresponding to 69.06% of questions answered correctly. The maximum score was 15 (93.75%) and the minimum was 6 (37.5%). Table 4 shows the percentage of correct answers obtained for each question. Except for questions 5, 6, 10, 11 and 13 (given bold in Table 4), all other questions received high percentage of correct answers (above 50%). The highest scores achieved by food handlers in this study focused on general cleaning procedures.

Almost all of the food handlers (92%) in our study believed that contaminated food could be distinguished by sensory properties. Half of them (56%) did not believe that food and beverage consumption in work place and their diarrheal infections threaten consumer's health. The lowest level of knowledge was obtained from the topics closely related to food safety issues especially source of food contamination (questions 5, 13) and sensory properties of contaminated food (question 11).

Questions, emphasizing hand washing before handling had highest percentage of correct answers

followed by questions related to wearing gloves and cleaning of utensils, respectively.

#### **Discussion and Conclusion**

RTE food handlers has critical role to ensure food safety and to prevent food borne diseases. As stated by Codex Alimentarius (10), all individuals in contact with food must be qualified and must recognize their role and responsibility in not to contaminate food. Likewise, every food producing point should provide a training program.

*S. aureus* is the very common pathogen reaching RTE food by food handlers who carry this agent on their nasal cavity or skin (1,33). Nose flora often contaminates fingers and back side of hands (15). Being the permanent flora of human skin, *S. aureus* could not be removed even after proper hand washing practices therefore no fixed acceptable contamination level is available for this agent (2). As shown in Table 1, *S. aureus* were isolated from 34% of the sampled population's hands ranging from 1 to 4.0x10<sup>1</sup> cfu/cm<sup>2</sup>. Higher results were obtained by Lues et al. (24) and Soares et al. (40) who reported 88% and 53.3% of food handler's hands to be contaminated with *S. aureus* and coagulase-positive staphylococci, respectively.

In contrast to *S. aureus*, Enterobacteriaceae numbers could be reduced by proper hand washing. Therefore, its presence points out to the possible presence of *Salmonella*, *Shigella*, *Yersinia*, *Proteus*, *Klebsiella* spp. and others (31). In this study, Enterobacteriaceae was detected from 44.6% of hands ranging from 5 to 20 cfu/cm<sup>2</sup>. Higher results were obtained in a comparable study, reporting

Enterobacteriaceae to be isolated from 56% of food handler's hands in South Africa (24). Enterobacteriaceae presence on hands is reported to not to be a good indicator of personal and toilet hygiene as hands might be contaminated with Enterobacteriaceae irrelevant to toilet use and could be reduced by hand washing (12).

Coliforms are often used to evaluate the efficiency of sanitation programs as their presence indicates the increased risk of pathogen existence. For different kind of surfaces, target value was suggested as <2.5 cfu/cm<sup>2</sup>(30). In this study, 32% of hands were found to exceed the target value for coliforms, which reveals the improper personal hygiene. Lues et al. (24) and Campos et al. (8) also reported 40% and 55.6% of retail food handler's hands to be contaminated with coliforms which are higher than our results.

Being normally absent on hands, *E. coli* is regarded as the better indicator of the fecal contamination and enteric pathogens compared to all Enterobacteriaceae family members (12). *E. coli* was detected from 16% of the food handler's thumbs in this study. *E. coli* presence on the handler's hands is worthy of consideration as these isolates could involve Enterohaemorrhagic *E. coli* O1111 or *E. coli* O157:H7. In the view of the literature, the specified limit for *E. coli* on surfaces is 1 cfu/cm<sup>2</sup> (23). High prevalence rate of the indicator bacteria in hands and some inappropriate attitudes observed in this study could be sourced from poor working conditions, and lack of periodical training. Most of the retail points were observed to have no hand washing sink to be used where needed.

All of the participants were male in this study. On the contrary Santos et al. (35) and Martins et al. (27) report 85% and 96% of participants to be female in Portugal, respectively. In the present study, about two third of the participants (65.2%) were aged between 25 and 45 years. Most of them were married (82.6%) and the education level of largest group (47.3%) was primary school. Two third of the participants had the experience more than 10 years on their work. Similarly, Marais et al. (25) reported more than 60% of managers to have >5 years of experience in food industry. Hand hygiene profiles of experience food handlers were reported to be more satisfactory than the hands of inexperienced staff (23). It is indicated that experienced staff are less likely to participate training activities (43).

Most of the food handlers were observed to not to use gloves (90.7%) and more than half of the participants were not wearing aprons during handling pastrami. Although food handlers knew when gloves needed to be worn, in practice this remains a problem, as they state that wearing gloves during pastrami slicing does not allow to slice thinner as preferred by consumers. In this regard hand hygiene and proper glove and apron use by food handlers need to be emphasized within the frame of the training program. On the contrary, Eren et al. (13) indicated that most of the food handlers had proper attitudes during food preparation including wearing aprons and gloves, using different cutting boards and knifes for raw meat and salads.

In this study, 10.6% of them had long nails and 14.6% wore jewelry. These results on long nails are lower than that reported by Miranda et al. (29) and Campos et al. (8) reporting 41.7% and 81.5% of long nailed food handlers respectively.

In this study, 60% of attendants declared that they received formal food hygiene training (Data not shown). Such training was done as once-off activity without refreshing training. Similarly, Baş et al. (4) reported that almost 52% of participants received food safety training in Turkey and the food safety knowledge score was 43.4. Campos et al. (8) in Brazil and Silva et al. (39) in Spain stated that only 35.9% and 43.3% of food handlers received periodical training, respectively. In this study, 79.3% (119/150) of the participants had the score above 62.5% and were reported to have "good knowledge" on food safety. Only one food handler had the score below 50% (Data not shown).

Food handlers who participated in the study answered 69.06% of questions correctly. This is higher than that obtained by Gomes-Neves et al. (14) and Martins et al. (27) who reported 62% and 56.5% of handlers had correct answers in Portugal respectively. Similarly, Marais et al. (25) obtained 46% of correct answers from their food hygiene knowledge study on food handlers in South Africa which is similar to Baş et al. (4) with the mean score of 43.4% in Turkey. On the other hand, Yardımcı et al. (44) reported that food staff achieved 76.5% of the total score from personal hygiene, 73.3% of food hygiene, and 76.6% of kitchen and equipment hygiene knowledge tests.

In this study the lowest level of knowledge was obtained from the topics closely related to food safety issues especially source of food contamination (questions 5, 13) and sensory properties of contaminated food (question 11) which is different from those obtained by Jevsnik et al. (19), Santos et al. (35), Marais et al. (25) and Martins et al. (27) reporting that food handlers have significantly lower level of knowledge on the role of temperature in maintaining food safety. These results suggest significantly lower level of knowledge and understanding probably due to lack of appropriate hygiene training on these topics. Low level of knowledge may result in decreased level of concern during food handling and promote wrong practices (27).

In our study, 92% of attendants believe that contaminated food could be distinguished by visual, olfactory or taste properties. These results are in same line with Gomes-Neves et al. (14), Walker et al. (42) and 158 Yeliz Yıldırım - Nurhan Ertaş Onmaz - Zafer Gönülalan - Harun Hızlısoy - Serhat Al - Fulden Karadal - Erhan Kum -Aytaç Akçay

Jevsnik et al. (19) with 55.7%, 57% and 52.5% of incorrect results for the similar questions respectively.

In this study, 56% of food handlers did not believe that food and beverage consumption in work place and their diarrheal infections threaten consumer's health which implies that they do not have perceptions on the sources of food contaminations. According to Codex Alimentarius (10), food handlers should not eat, handle money or engage in any act that could contaminate the food during performance of their activities. Lack of knowledge on contamination sources might increase improper implementations in work place which in turn could promote microbial hazards and food borne diseases.

Other topics having low scores from food handlers were; freezing and in advance preparing of food. More than half of respondents (60%) believed that freezing process could kill all the bacteria in the food. Baş et al. (4) also reported deficit knowledge on the critical temperature of food preservation among food handlers.

In our study, questions emphasizing hand washing had highest score followed by questions related to wearing gloves and cleaning of utensils, respectively. Our results are compatible to the works of Martins et al. (27) and Marais et al. (25) reporting the best results from the issues of cleaning and sanitation. Although 96% of participants had the correct answers on glove use, their glove use practice does not reach this score. Seaman and Eves (36) underline that training could only be effective if the knowledge leads to required changes in behavior during food handling.

In this study, food handler's awareness on not handling food in case of infectious wounds on their skin was evident (92%). Most of food handlers (64-68%) gave correct answers to the questions referring to high risk population group.

Result of this study revealed that although the food handlers have the satisfactory knowledge on food safety, hand hygiene profiles do not seem to be parallel to their questionnaire results. A possible explanation for these findings may be related to low educational level of food handlers. This finding corroborates with the results of Baş et al. (4) who indicate that training may cause an increase in knowledge on food safety however may not always give rise to a positive change in behavior. Similarly, no significant relation was reported between the hygiene training of food handlers and hygiene standards of retail points (42). It is a great concern in food industry to encourage food handlers implement the principles they learnt on food safety (26). However, Cakiroglu and Ucar (7) reported inadequate knowledge of hygiene perception among food handlers and they underlined the immediate need for training on the subject.

The reality of the retail points exhibited that despite the fact of high level of food safety knowledge, food handler's hand hygiene profiles were not reflecting the same results. The main problems identified within the frame of this study were poor working conditions, lack of hand washing infrastructure in the retail points and lack of specific, periodic and formal training among food handlers although it is legal requirement in Turkey. Evaluation of the effectiveness of training is an important part of food safety, as stated by ISO 22000:2005 (18).

Data obtained from this study reveal that despite their adequate food safety knowledge, food handlers on pastrami retail points were lack of food safety practices. Therefore, it is not only sufficient to provide food safety trainings, but also the sources of failures in the practice should be identified and resolved.

#### Acknowledgements

The abstract of this study was submitted at the International Congress on Food of Animal Origin held in of North Cyprus, on 10-13 November 2016.

#### **Financial Support**

This research was funded by "Erciyes University, Scientific Research Project Coordination Unit" (TCD-2013-4396).

#### **Conflict of Interest**

All authors declare that they have no conflicts of interest.

#### References

- Angelillo IF, Viggiani NMA, Rizzo L, et al (2000): Food handlers and food borne diseases: Knowledge, attitudes, and reported behavior in Italy. J Food Protect, 63, 381-385.
- Ayçiçek H, Aydoğan H, Küçükkaraaslan A, et al (2004): Assessment of the bacterial contamination on hands of hospital food handlers. Food Control, 15, 253-259.
- **3.** Baird RM, Lee WH (1995): Media used in the detection and enumeration of Staphylococcus aureus. Int J Food Microbiol, **26**, 15-24.
- Baş M, Ersun AS, Kıvanç G (2006): The evaluation of food hygiene knowledge, attitudes, and practices of food handlers in food businesses in Turkey. Food Control, 17, 317-322.
- Blood RM, Curtis GDW (1995): Media for 'total' enterobacteriaceae, coliforms and Escherichia coli. Int J Food Microbiol, 26, 93-115.
- Buzby JC, Roberts T (2009): The economics of enteric infections: human food borne disease costs. Gastroenterology, 136, 1851-1862.
- 7. Cakiroglu FP, Ucar A (2008): Employees' perception of hygiene in the catering industry in Ankara (Turkey). Food Control 19, 9-15.

- 8. Campos AKC, Cardonha ÂMS, Pinheiro LBG, et al (2009): Assessment of personal hygiene and practices of food handlers in municipal public schools of Natal, Brazil. Food Control, **20**, 807-810.
- **9.** Clayton D, Griffith CJ, Price P, et al (2002): Food Handlers' beliefs and self-reported practices. Int J Environ Health Res, **12**, 25-39.
- **10.** Codex Alimentarius (2003): Recommended international code of practice general principles of foodhygiene. CAC/RCP 1-1969, Rev. 4.
- **11.** De Boer E (1998): Update on media for isolation of enterobacteriaceae from foods. Int J Food Microbiol, **45**, 43-53.
- De Wit JC, Rombouts FM (1992): Faecal microorganisms on the hands of carriers: Escherichia coli as model for Salmonella. Zentralbl Hyg Umweltmed, 193, 230-236.
- **13.** Eren R, Nebioğlu O, Şık A (2017): Knowledge Levels on Food Safety of Employees Working in Hotel Enterprises' kitchen: The example of Alanya. JOMAT, **1**, 47 - 64
- Gomes-Neves E, Araújo AC, Ramos E, et al (2007): Food handling: comparative analysis of general knowledge and practice in three relevant groups in Portugal. Food Control, 18, 707-712.
- **15.** Gorman R, Bloomfield S, Aley CC (2002): A study of cross-contamination of food-borne pathogens in the domestic kitchen in the Republic of Ireland. Int J Food Microbiol, **76**, 143-150.
- 16. Greig JD, Todd EC, Bartleson CA, et al (2007): Outbreaks where food workers have been implicated in the spread of food borne disease. Part 1. Description of the problem, methods, and agents involved. J Food Protect, 70, 1752-1761.
- **17. Ingham SC, Reyes JC, Schoeller NP, et al** (2000): Potential use of presumptive enterococci and staphylococci as indicators of sanitary conditions in plants making hard Italian-type cheese. J Food Protect, **63**, 1697-1701.
- **18.** International Standard Organization (2005): ISO/IEC22000-Food safety management systems. Requirements for any organization in the food chain. Geneva, Switzerland.
- **19.** Jevsnik M, Hlebec V, Raspor P (2008): Food safety knowledge and practices among food handlers in Slovenia. Food Control, **19**, 1107-1118.
- Jiang XP, Doyle MP (1999): Fate of Escherichia coli 0157:H7 and Salmonella Enteritidis on currency. J Food Protect, 62, 805-807.
- **21.** Jianu C, Chis C (2012): Study on the hygiene knowledge of food handlers working in small and medium-sized companies in western Romania. Food Control, **26**, 151-156.
- 22. Kusumaningrum HD, Van Putten MM, Rombouts FM, et al (2002): Effects of antibacterial dish washing liquid on food borne pathogens and competitive microorganisms in kitchen sponges. J Food Protect, **65**, 61-65.
- 23. Legnani P, Leoni E, Berveglieri M, et al (2004): Hygienic control of mass catering establishments, microbial monitoring of food and equipment. Food Control, 15, 205-211.
- 24. Lues JFR, Van Tonder I (2007): The occurrence of indicator bacteria on hands and aprons of food handlers in

the delicatessen sections of a retail Group. Food Control, **18**, 326-332.

- 25. Marais M, Conradie N, Labadarios D (2007): Small and micro enterprises–aspects of knowledge, attitudes and practices of managers' and food handlers' knowledge of food safety in the proximity of Tygerberg Academic Hospital, Western Cape. South Afr J Clin Nutr, 20, 50-61.
- **26.** Martinez-Tome M, Vera AM, Murcia MA (2000): Improving the control of food production in catering establishments with particular reference to the safety of salads. Food Control, **11**, 437-445.
- 27. Martins RB, Hogg T, Otero JG (2012): Food handlers' knowledge on food hygiene: the case of a catering company in Portugal. Food Control, 23, 184-190.
- McIntyre L, Vallaster L, Wilcott L, et al (2013): Evaluation of food safety knowledge, attitudes and selfreported hand washing practices in FOODSAFE trained and untrained food handlers in British Columbia, Canada. Food Control, 30, 150-156.
- Miranda LK, Damasceno KSFSC, Cardonha AMS (2002): Panos de prato e mãos de manipuladores: avaliação das condições higiênico-sanitárias. Higiene Alimentar, 16, 51-58.
- **30.** Moore G, Griffith CA (2002): Comparison of surface sampling methods for detecting coliforms on food contact surfaces. Food Microbiol, **19**, 65-73.
- **31.** Nel S, Lues JFR, Buys EM, et al (2004): Bacterial populations associated with meat from the deboning room of a high throughput red meat abattoir. Meat Sci, 66, 667-674.
- **32.** Perez-Rodriguez F, Castro R, et al (2010): Evaluation of hygiene practices and microbiological quality of cooked meat products during slicing and handling at retail. Meat Sci, **86**, 479-485.
- **33.** Portocarrero SM, Newman M, Mikel B (2002): Staphylococcus aureus survival, staphylococcal enterotoxin production and shelf stability of country-cured hams manufactured under different processing procedures. Meat Sci 62, 267-273.
- **34. Regulation EC** (2004): *No* 852/2004 of European *Parliament and of the Council of 29 April 2004 on Hygiene of Foodstuffs.* OJEC, 18 p.
- **35.** Santos MJ, Nogueira JR, Patarata L, et al (2008): Knowledge levels of food handlers in Portuguese school canteens and their self-reported behavior towards food safety. Int J Environ Health Res, **18**, 387-401.
- **36.** Seaman P, Eves A (2006): *The management of food safety* - *The role of food hygiene training in the UK service sector*. Int J Hosp Manag, **25**, 278-296.
- Seaman P, Eves A (2010): Perceptions of hygiene training amongst food handlers, managers and training providers -A qualitative study. Food Control, 21, 1037-1041.
- **38.** Sheppard J, Kipps M, Thomson J (1990): *Hygiene and hazard analysis* 192-226. In: Cooper C (Ed), Progress in tourism, recreation and hospitality management, Belhavel Press, London.
- **39. Silva CS, Germano MIS, Germano PML** (2003): Condições Higiênico– Sanitárias dos Locais de Preparação da Merenda Escolar, da Rede Estadual de Ensino em São Paulo, SP. Higiene Alimentar, **17**, 49-55.

160 Yeliz Yıldırım - Nurhan Ertaş Onmaz - Zafer Gönülalan - Harun Hızlısoy - Serhat Al - Fulden Karadal - Erhan Kum -Aytaç Akçay

- **40.** Soares LS, Almeida RCC, Cerqueira ES, et al (2012): Knowledge, attitudes and practices in food safety and the presence of coagulase-positive staphylococci on hands of food handlers in the schools of Camaçari, Brazil. Food Control, **27**, 206-213.
- **41. Sumbuloğlu V, Sumbuloğlu K** (2005): *Klinik ve Saha Araştırmalarında Örnekleme Yöntemleri ve Örneklem Büyüklüğü*, Hatiboğlu Yayınevi, 1. Baskı, Ankara.
- 42. Walker E, Pritchard C, Forsythe S (2003): Food handlers' hygiene knowledge in small food business. Food Control, 14, 339-343.
- **43.** Yapp C, Fairman R (2006): Factors affecting food safety compliance within small and medium-sized enterprises: Implications for regulatory and enforcement strategies. Food Control, **17**, 42-51.
- **44.** Yardımcı H, Hakli G, Cakıroglu FP, et al (2015): Hygiene Knowledge of Food Staff in Catering Industry: A Sample From Turkey. SAGE Open 1–7.

Ankara Üniv Vet Fak Derg, **67**, 161-168, 2020 DOI: 10.33988/auvfd.594391

# Three-dimensional tomographic reconstruction and morphometric analysis of skull in gazelles (*Gazella subgutturosa*)

#### Bestami YILMAZ<sup>1,a,⊠</sup>, İsmail DEMİRCİOĞLU<sup>1,b</sup>, Faruk BOZKAYA<sup>2,c</sup>, Nazan GEZER İNCE<sup>3,d</sup>

<sup>1</sup>Harran University, Faculty of Veterinary Medicine, Department of Anatomy, Şanlıurfa; <sup>2</sup>Harran University, Faculty of Veterinary Medicine, Department of Genetics, Şanlıurfa; <sup>3</sup>İstanbul Cerrahpaşa University, Faculty of Veterinary Medicine, Department of Anatomy, İstanbul, Turkey.

> <sup>a</sup>ORCID: 0000-0002-0901-3129, <sup>b</sup>ORCID: 0000-0002-0724-3019, <sup>c</sup>ORCID: 0000-0001-6423-8067, <sup>d</sup>ORCID: 0000-0003-1627-5757

<sup>™</sup>Corresponding author: byilmaz@harran.edu.tr Received date: 19.07.2019- Accepted date: 14.11.2019

**Abstract:** This study was carried out to determine the osteometric features of the skull by using three dimensional computed tomography images in gazelles (*Gazelle subgutturosa*). In the study, nine skull samples of adult gazelles (*Gazella subgutturosa*) were used. Images of the skull sections of 0.625 mm thickness were acquired by using a computer tomography device with 64 detectors applying 80 kV, 200 mA and 639 mGY. Three-dimensional images of the skull samples were reconstructed and morphometric measurements (39 linear, 1 volumetric and 1 surface area) were performed by using the software program MIMICS 12.1. Mean skull volumes in males and females were found to be  $115.74\pm2.43$  cm<sup>3</sup> and  $87.69\pm1.09$  cm<sup>3</sup> while the mean skull surface areas in males and females were 79.62 $\pm$ 8.56 cm<sup>2</sup> and 77.34 $\pm$ 1.18 cm<sup>2</sup>, respectively. Significant differences between males and females for median frontal length (MFL), frontal length (FRL), upper neurocranium length (UNCL), greatest length of the lacrimal bone (GLLB), oral palatal length (OPL), length of the upper molar row (LUMR) and the greatest neurocranium breadth (GNCB) were observed. The difference in the cranial index between males and females was statistically significant (P<0.01). The data obtained in this study will contribute to detect differences between the gazelles and other species with respect to skull morphometry.

Keywords: Computed tomography, gazelle, morphometry, reconstruction, skull.

#### Ceylanlarda (*Gazella subgutturosa*) kafatasının üç boyutlu tomografik rekonstruksiyonu ve morfometrik analizi

Özet: Bu çalışma; ceylan kafatasının bilgisayarlı tomografi görüntülerini kullanılarak kafatası kemiklerinin osteometrik özelliklerini belirlemek amacıyla yapılmıştır. Çalışmada 9 adet erişkin ceylan (*Gazella subgutturosa*) kafatası kullanıldı. Kafataslarının 64 dedektörlü CT cihazı ile 80 kv, 200 MA, 639 mGY ve 0,625 mm kesit kalınlığında görüntüleri alındı. Bu görüntüler MIMICS 12.1 programı yardımıyla üç boyutlu yapıya dönüştürülerek morfometrik ölçümleri (39 linear, 1 hacim ve 1 yüzey alanı) yapıldı. Erkeklerde kafatasının ortalama hacim değeri 115,74±2,43 cm<sup>3</sup>, dişilerde 87,69±1,09 cm<sup>3</sup> olarak tespit edilirken cranium'un ortalama yüzey alanı erkeklerde 79,62±8,56 cm<sup>2</sup>, dişilerde 77,34±1,18 cm<sup>2</sup> olarak bulundu. Çalışmada, median frontal uzunluk (MFL), frontal uzunluk (FRL), üst neurocranium uzunluğu (UNCL), lacrimal kemiğin maximum uzunluğu (GLLB), oral palatal uzunluk (OPL), üst molar diş sırası uzunluğu (LUMR) ve en büyük neurocranium genişliği (GNCB) parametrelerinde dişi ve erkekler arasındaki farklar istatistiksel olarak anlamlı bulundu. Cranial index değeri açısından dişi ve erkekler arasındaki fark istatistiksel olarak anlamlıydı (P<0,01). Çalışmada elde edilen bilgilerin ceylan türlerinin tipolojisi ile diğer türlerle arasındaki farklılıkların tespitine katkı sunacağı düşünülmektedir.

Anahtar sözcükler: Bilgisayarlı tomografi, ceylan, kafatası, morfometri, rekonstruksiyon.

#### Introduction

Even among the closely related species, there are apparent differences in the skeletal systems. These differences are crucial for taxonomic classification of species and for evaluation of the archaeological or forensic findings (26). Skull is the most studied bone for reconstructing the evolutional taxonomy. However, the assignment of the species based on skull characteristics is difficult due to variation within species (1). Knowledge of cranial morphometry is also important for the diagnosis of cranial or dental deformities for designing implants or dental instruments (26, 27).

Three different techniques have been used for obtaining osteometric parameters. The first is the measurement of bones obtained from archaeological excavations or after maceration by using a compass (23). The second is the evaluation of the radiological images from the target region (16). The third one is the measurement of the images obtained by using computer tomography (CT), which is a recently more frequently used technique (27). Images of two-dimensional sections from CT are compiled to reconstruct a three dimensional (3D) image using special software programs (10, 22). The 3D modeling technique is widely employed in plastic surgery, orthopedic surgery, neurosurgery, traumatology and medical education (17).

Gazella is one of the most species-rich genus comprising numerous species within Bovidae (1). Gazelles in Sanliurfa belongs to *Gazella subgutturosa*, which has a wide distribution area ranging from China to North Africa. Since the second half of 20<sup>th</sup> - century number of the gazelles have rapidly declined due to human activities including habitat destruction, expansion of the agricultural areas, hunting, etc. (19).

Several morphometric studies have been performed for establishing a comprehensive and reliable database in gazelles (1, 9, 31). The objective of this study was to morphometrically analyze the skulls of gazelles by using the CT images in order to provide species specific data that can be used by veterinary clinicians for managing pathological formations on the skull.

#### **Material and Methods**

Animal material: In the study nine cadavers (5 females and 4 males) of adult gazelles were used. Body weights of the cadavers were among 11.4 - 18.1 kg. The cadavers were submitted to the clinics of Harran University Animal Hospital in Sanliurfa province of Turkey for treatment yet died for various reasons. The animals had no clinical or pathological skull problems. The use of the cadavers was approved by the General Directorate of Nature Conservation and National Parks-Turkey (Approval no: 2017/209842) and Harran University Animal Experimentation Local Ethics Committee (Approval no: 2018/006-11).

CT-Imaging, reconstruction and morphometric analysis: For obtaining the CT images the gazelle

cadavers were placed on a sternal position into a CT device with 64 detectors (GE Company, USA). Images of the skull sections of 0.625 mm thickness were acquired by applying 80 kV, 200 mA, and 639 mGY. The CT images were stored in DICOM format and the 3D skull images were reconstructed using the basic module of the 3D modeling program MIMICS 20.1 (The Materialise Group, Leuven, Belgium). Osteometric measurements on the digital images were performed for 39 different parameters according to the measurement points reported in the literature (25, 29). Definitions and the abbreviations of the studied parameters were shown in Table 1. After morphometric measurements, volume and surface area of the skulls were estimated by excluding the horns and mandible. Further 6 different indices were calculated based on the craniometric measurements (Table 2). The definitions were based on Nomina Anatomica Veterinaria (20).

Statistical analysis: All morphometric parameters were expressed as Mean  $\pm$  Standard Error (SE). The presence of significant differences between sexes was examined by using the Mann-Whitney U test. For statistical analyses SPSS, 17.0 was used.

#### Results

In this study, 39 linear parameters of the skull were measured (Figure 1-4). The mean  $\pm$  standard error values for each parameter in males and females were shown in Table 3. Statistically significant differences (P<0.05) between males and females for MFL (median frontal length), FRL (frontal length), UNCL (upper neurocranium length), GLLB (greatest length of the lacrimal bone), OPL (oral palatal length), LUMR (length of the upper molar row) and GNCB (greatest neuro-cranium breadth) were observed.

Furthermore, cranial volume values in males and females were detected to be  $115.74\pm2.43$  cm<sup>3</sup> and  $87.69\pm1.09$  cm<sup>3</sup>, respectively. The cranial surface area in males and females was  $79.62\pm8.56$  cm<sup>2</sup> and  $77.34\pm1.18$  cm<sup>2</sup>, respectively (Table 4). The difference in mean cranial volume between males and females was significant while there was no difference in cranial surface area between sexes. Data on the skull indices have been shown in (Table 5). A statistically significant difference between males and females was observed only for cranial index values.

Parameter	Abbreviation	Definition
1	TLS	Total length of the skull: the distance between akrokranion-prosthion
2	CBL	Condylobasal length: caudal border of occipital condyles-prosthion
3	TLCB	Total length of the cranial base: basion-prosthion
4	SSL	Short skull length: basion-premolare
5	PPL	Premolare-prosthion length
6	NCL	Neurocranium length: basion-nasion
7	ULVC	Upper length of the viscerocranium: nasion-prosthion
8	MFL	Median frontal length: akrokranion-nasion
9	ACBL	Akrokranion-bregma length
10	FRL	Frontal length: bregma-nasion
11	UNCL	Upper neurocranium length: akrokranion-supraorbitale
12	FCL	Facial length: supraorbitale-prosthion
13	ACIO	Akrokranion-infraorbitale length
14	GLLB	Greatest length of the lacrimal bone
15	GLNB	Greatest length of the nasal bone: nasion-rhinion
16	EOPL	Entorbitale-prosthion length
17	DOCI	Distance between the caudal border of occipital condyle and the infraorbitale
18	DTL	Dental length: postdentale-prosthion
19	OPL	Oral palatal length: palatinoorale-prosthion
20	LLPM	Lateral length of the premaxilla: nasointermaxillare-prosthion
21	LMTR	Length of the maxillary tooth row
22	LUMR	Length of the upper molar row
23	LUPR	Length of the upper premolar row
24	GIWO	Greatest inner width of the orbit: ectorbitale-entorbitale
25	GIHO	Greatest inner height of the orbit
26	GMB	Greatest mastoid breadth: otion-otion
27	GBOC	Greatest breadth of the occipital condyles
28	GBPP	Greatest breadth at the bases of the paracondylar processes
29	GBFM	Greatest breadth of the foramen magnum
30	HFM	Heigth of the foramen magnum: basion-opisthion
31	LBP	Least breadth of parietal
32	GBLH	Greatest breadth between the lateral borders of the horncore base
33	GNCB	Greatest neurocranium breadth: euryon-euryon
34	GFB	Greatest frontal breadth: ectorbitale-ectorbitale
35	LBO	Least breadth between the orbits: entorbitale-entorbitale
36	FCB	Facial breadth: between facial tuberosities
37	GBAN	Greatest breadth across the nasal bones
38	GBAP	Greatest breadth across the premaxilla
39	GPB	Greatest palatal breadth

Table 1. Studied cranial parameters (according to Von den Driesch (30)).ParameterAbbreviationDefinition

Table 2. Indices and formulas of the skulls (According to Parés-Casanova (26)).

Studied indexes	Formulas
Skull index	greatest frontal breadth (var. 34) / total length of the skull (var. 1) x 100
Cranial index	greatest neurocranium breadth (var. 33) / median frontal length (var. 8) x 100
For. magnum index	height of the for. magnum (var. 30) / greatest breadth of the for. magnum (var. 29) x 100.
Orbital index	orbital inner width (var. 24) / orbital inner height (var. 25) x 100
Facial index	facial width (var. 36) / facial length (var. 12) x 100.
Nasal index	greatest breadth across the nasals (var. 37) / greatest length of the nasals (var. 15) $\times$ 100.

		Gen	eral statistics		Females	Males	n	
Para	ameter	Mean±SEM	Mean±SEM Min. Max.			Mean±SEM	Р	
1.	TLS	165.59±2.78	155.29	178.92	164.40±4.89	$167.08{\pm}2.28$	0.730	
2.	CBL	163.87±2.85	156.03	180.27	162.70±4.43	$165.34 \pm 3.84$	0.556	
3.	TLCB	153.65±2.34	146.88	169.08	153.29±4.00	$154.10 \pm 2.43$	0.556	
4.	SSL	113.10±2.12	104.46	125.48	111.51±3.75	$115.10 \pm 1.04$	0.190	
5.	PPL	40.02±0.93	34.40	44.49	$40.86 \pm 1.15$	$38.97 \pm 1.53$	0.556	
6.	NCL	103.56±1.52	100.01	113.93	$105.51 \pm 2.41$	$101.13 \pm 0.80$	0.190	
7.	ULVC	83.71±1.88	73.97	92.27	$84.01 \pm 2.07$	$83.33 \pm 3.75$	0.905	
8.	MFL	98.45±2.19	89.29	112.52	$102.37 \pm 2.66$	$93.55 \pm 1.56$	0.016	
9.	ACBL	31.92±2.07	22.97	40.71	$33.32 \pm 3.27$	$30.17 \pm 2.45$	0.556	
10.	FRL	82.41±2.91	71.92	97.85	$88.27{\pm}3.09$	$75.08 \pm 1.58$	0.032	
11.	UNCL	72.83±3.29	55.76	90.14	$79.14 \pm 3.26$	$64.94 \pm 3.15$	0.016	
12.	FCL	121.28±4.94	93.00	141.39	$111.89 \pm 5.02$	$133.03 \pm 4.68$	0.016	
13.	ACIO	118.75±1.79	111.49	129.16	$118.72 \pm 2.73$	$118.78 \pm 2.62$	0.730	
14.	GLLB	21.09±0.79	16.51	23.53	$19.80 \pm 1.11$	$22.71 \pm 0.38$	0.05	
15.	GLNB	49.95±2.93	36.75	59.93	$46.27{\pm}4.49$	$54.55 \pm 2.26$	0.286	
16.	EOPL	80.65±1.55	73.64	87.64	$79.35 \pm 2.30$	$82.28 \pm 1.99$	0.556	
17.	DOCI	118.53±1.89	112.62	130.91	$120.05 \pm 3.20$	$116.63 \pm 1.48$	0.730	
18.	DTL	94.40±1.95	88.18	105.11	$96.98 \pm 2.93$	$91.18 \pm 1.51$	0.111	
19.	OPL	77.97±3.60	64.86	94.76	$85.47 \pm 3.72$	$68.60 \pm 1.34$	0.016	
20.	LLPM	50.02±1.42	43.98	57.90	$48.08 \pm 1.48$	$52.45 \pm 2.22$	0.190	
21.	LMTR	53.22±1.28	47.67	59.51	$51.78 \pm 1.70$	$55.02 \pm 1.73$	0.413	
22.	LUMR	30.15±1.42	24.39	36.17	$27.83 \pm 1.70$	$33.04 \pm 1.52$	0.05	
23.	LUPR	22.32±0.69	18.70	25.06	$23.44 \pm 0.60$	$20.91 \pm 1.03$	0.111	
24.	GIWO	33.46±0.32	32.19	34.86	$33.16 \pm 0.52$	$33.83 \pm 0.28$	0.413	
25.	GIHO	34.88±0.57	32.59	37.23	$34.10 \pm 0.54$	$35.84{\pm}0.94$	0.111	
26.	GMB	51.95±1.04	46.46	55.96	$52.76 \pm 1.76$	$50.94{\pm}0.83$	0.413	
27.	GBOC	33.92±1.57	27.31	40.84	$34.13 \pm 2.31$	$33.65 \pm 2.41$	1.000	
28.	GBPP	48.28±1.67	39.79	54.17	$48.12 \pm 2.83$	$48.47{\pm}1.83$	1.000	
29.	GBFM	16.28±0.36	14.57	18.35	$15.87{\pm}0.41$	$16.80 \pm 0.57$	0.190	
30.	HFM	15.03±0.32	13.44	16.27	$15.53 \pm 0.27$	$14.42 \pm 0.52$	0.190	
31.	LBP	35.74±0.96	30.88	40.14	$36.42 \pm 1.52$	$34.89 \pm 1.11$	0.413	
32.	GBLH	55.75±2.25	49.95	65.15	-	$55.75 \pm 3.37$	-	
33.	GNCB	$56.84 \pm 0.84$	52.98	60.25	$55.21{\pm}0.75$	$58.88{\pm}0.89$	0.032	
34.	GFB	69.16±2.24	57.12	75.93	$68.95{\pm}3.29$	$69.44 \pm 3.47$	0.905	
35.	LBO	82.07±1.11	78.56	88.03	$80.80 \pm 1.83$	$83.66 \pm 0.55$	0.190	
36.	FCB	56.47±1.12	51.96	63.40	$56.79{\pm}1.94$	$56.09 \pm 1.10$	1.000	
37.	GBAN	24.60±1.19	19.32	28.82	$23.30{\pm}1.58$	$26.24{\pm}1.64$	0.286	
38.	GBAP	28.96±1.17	21.45	33.91	$27.84{\pm}1.64$	$30.35 \pm 1.61$	0.730	
39.	GPB	47.39±0.87	44.46	51.77	$47.39 \pm 1.24$	$47.40 \pm 1.40$	0.905	

Table 3. The mean and standard deviations of the skull measurements (mm).

S.E.: Standard error of mean.

Table 4. The mean and standard deviations of the skull volume and surface area.

Parameter -	Ger	neral statistics		Females	Males	Р
	Mean±SEM	Min.	Max.	Mean±SEM	Mean±SEM	P
Volume (cm <sup>3</sup> )	101.71±2.31	69.14	151.67	87.69±1.09	115.74±2.43	0.008
Area (cm <sup>2</sup> )	$78.48 \pm 9.80$	61.31	91.17	77.34±1.18	79.62±8.56	NS

SEM: Standard error of mean, NS: Non significant.

I. J.	Gei	neral statistic	s	Females	Males	р
Index	Mean±SEM	Min.	Max.	Mean±SEM	Mean±SEM	r
Skull	41.86±1.50	32.94	46.36	42.12±2.45	41.54±1.86	0.730
Cranial	58.01±1.74	49.87	65.60	$54.03 \pm 1.08$	$62.98 \pm 1.18$	0.016
For. magnum	92.84±3.35	73.24	108.92	98.16±3.53	86.18±4.49	0.111
Orbital	96.09±1.49	90.59	104.02	97.27±1.12	94.61±3.15	0.286
Facial	47.45±2.92	37.97	68.17	51.51±4.36	42.36±2.04	0.063
Nasal	50.30±3.17	38.44	64.27	51.67±4.70	48.60±4.67	0.556

Table 5. The mean and standard deviations of the craniofacial indices.

SEM: Standard error of mean.



Figure 1. Measurement points of craniometric variables in the gazelle skull (lateral view).

A: Akrokranion, **Br:** Bregma, **Ect:** Ectorbitale, **Ent:** Entorbitale, **Ni:** Nasointermaxillare **If:** Infraorbitale, **N:** Nasion, **P:** Prosthion, **6:** Neurocranium length (NCL), **7:** Upper length of the viscerocranium (ULVC), **14:** Greatest length of the lacrimal bone (GLLB), **17:** Distance between the caudal border of one occipital condyle and the infraorbitale of the same side (DOCI), **20:** Lateral length of the premaxilla (LLPM), **24:** Greatest inner width of the orbit (GIWO), **25:** Greatest inner height of the orbit (GIHO).



Figure 2. Measurement points of craniometric variables in the gazelle skull (dorsal view).

A: Akrokranion, **Br:** Bregma, **Ect:** Ectorbitale, **Ent:** Entorbitale, **Eu:** Euryon, **If:** Infraorbitale, **N:** Nasion, **P:** Prosthion, **Rh:** Rhinion, **Sp:** Supraorbitale, **1:** Total length of the skull (TLS), **8:** Median frontal length (MFL), **9:** Akrokranion-bregma length (ACBL), **10:** Frontal length (FRL), **11:** Upper neurocranium length (UNCL), **12:** Facial length (FCL), **13:** Akrokranion-infraorbitale length (ACIO), **15:** Greatest length of the nasal bone (GLNB), **16:** Entorbitale-prosthion length (EOPL), **31:** Least breadth of parietal (LBP), **33:** Greatest neurocranium breadth (GNCB), **34:** Greatest frontal breadth (GFB), **35:** Least breadth between the orbits (LBO), **36:** Facial breadth (FCB), **37:** Greatest breadth across the nasal bones (GBAN), **38:** Greatest breadth across the premaxilla (GBAP).



Figure 3. Measurement points of craniometric variables in the gazelle skull (ventral view).

**B:** Basion, **P:** Prosthion, **Pd:** Postdentale, **Pm:** Premolare, **Po:** Palatinoorale, **2:** Condylobasal length (CBL), **3:** Total length of the cranial base (TLCB), **4:** Short skull length (SSL), **5:** Premolare-prosthion length (PPL), **18:** Dental length (DTL), **19:** Oral palatal length (OPL), **21:** Length of the maxillary tooth row (LMTR), **22:** Length of the upper molar row (LUMR), **23:** Length of the upper premolar row (LUPR), **39:** Greatest palatal breadth (GPB).



Figure 4. Measurement points of craniometric variables in the gazelle skull (occipital view). A: Akrokranion, B: Basion, O: Opisthion, Ot: Otion, 26: Greatest mastoid breadth (GMB), 27: Greatest breadth of the occipital condyles (GBOC), 28: Greatest breadth at the bases of the paracondylar processes (GBBPP), 29: Greatest breadth of the foramen magnum (GBFM), 30: Heigth of the foramen magnum (HFM).

#### **Discussion and Conclusion**

Craniometric analyses have been used to differentiate species within the same genus and to investigate morphological variations within species. Several reports on craniometric measurements using traditional methods (the help of scale and digital calipers) in gazelles are found in the literature (7, 31). This study presents for the first time morphometric and volumetric measurements of the skull in gazelles by using threedimensional CT images. Due to the lack of data on CT based measurements in gazelles, data obtained from different gazelle species by traditional methods or data obtained from sheep and goats were used for comparison.

Due to remarkable morphological variations both among gazelle species and among individuals within the same species, assigning an individual to a certain species might be difficult (28). Therefore, more data are required for assessing the morphometric variation within the species. On the other hand, craniofacial index parameters are also necessary for examining craniofacial deformities and investigating brain development (13). Zhu (31) has reported the skull index by examining the craniometrics values of Tibetan gazelle as  $43.22\pm0.44$  mm, cranial index as  $58.37\pm0.80$  mm and facial index as  $116.37\pm1.24$  mm. The facial index value found in the present study ( $47.45\pm2.92$  mm) was lower than that reported by Zhu (31). The difference might be attributed to the use of different species and methods.

The orbital region plays an important role in craniofacial measurements, forensic processes and differential diagnosis (8). A tubular shape of orbita was observed in gazelles in the present study. The orbita can have a different shapes depending on the species and the breed of the same species. It has been reported that orbita has the shape of almond in Spanish Xisqueta sheep (24) while it has an oval shape in Mehreban sheep of Iran (14). Even a bilateral variation between the right and left orbitas in Kagani goats (Capra hircus) has been reported (12). In accordance with the present study Leslie (18), has reported a similar shape of orbita in Procapra picticaudata. Similar to our findings Parés-Casanova et al. (24) have reported an orbital index value of 97.27±1.12 mm and 94.61±3.15 mm in female and male Spanish Xisqueta sheep respectively.

Mean breadth and height of foramen magnum in the gazelles were measured as  $16.28\pm0.36$  mm and  $15.03\pm0.32$  mm respectively and foramen magnum index was  $92.84\pm3.35$  mm. These values were lower than those found in sheep (21) and goats (15). Similar to those reported in sheep and goats (15, 21) the horizontal diameter of the foramen magnum was longer than its vertical diameter in the gazelles.

Sexual dimorphism is common among mammals and has been an important evolutionary factor in social ecology (5). The effect of sex on bone morphology has been intensively studied in humans (2) goats (6) and wild sheep (11). However, the limited number of studies on the effect of sex on bone morphology in gazelles have been conducted (30). In the present study, significant differences between males and females were observed for MFL (median frontal length), FRL (frontal length), UNCL (upper neurocranium length), GLLB (greatest length of the lacrimal bone), OPL (oral palatal length), LUMR (length of the upper molar row) and GNCB (greatest neurocranium breadth).

Conventional radiological methods used for assessing the skull volume employ two-dimensional measurements. Computer tomography based methods present a more precise and noninvasive way for estimating in vivo skull volume (3). Mean skull volumes in females and males were detected as  $87.69\pm1.09$  cm<sup>3</sup> and  $115.74\pm2.43$  cm<sup>3</sup>, respectively. In contrast to the findings in this study, Chanpanitkitchote et al. (4) have reported a skull volume of Grant's gazelles (*Nanger granti*) as  $1016\pm11$  cm<sup>3</sup>. The differences in the morphometric values

between the species have been attributed to inclusion or exclusion of mandible, horn status of the animal, measurement methods used or live weight of the animal.

In conclusion, new technologies like CT presents opportunities for obtaining comprehensive data on skull morphometry in animals. This study was the first reporting the use of CT for morphometric analysis of the skull in goitered gazelle (*Gazella subgutturosa*). The data obtained in this study will be useful for not only the evaluation of CT images from facial, cranial of dental deformities but also for determining the sex based on bone morphometry and for taxonomical studies. However further studies are necessary for comparing the data obtained from 3D modeling and actual measurements on skulls by including larger sample size.

#### **Financial Support**

This study was supported by Scientific Research Center of Harran University (Project number: 18004).

#### **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### References

- 1. Bärmann EV, Wronski T, Lerp H, et al (2013): Morphometric and genetic framework for Gazella. Zoo J Linnean Soc, 169, 673-696.
- Best KC, Garvin HM, Cabo LL (2018): An investigation into the relationship between human cranial and pelvic sexual dimorphism. J Forensic Sci, 63, 990-1000.
- **3.** Breiman RS, Beck JW, Korobkin M, et al (1982): Volume determinations using computed tomography. AJR Am J Roentgenol, **138**, 329-333.
- 4. Chanpanitkitchote P, Tangphokhanon W, Nomsiri R, et al (2015): *Ratio of cranial and skull volumes in 25 mammalian species*. Vet Integr Sci, **13**, 165-177.
- Christiansen P, Harris JM (2012): Variation in craniomandibular morphology and sexual dimorphism in Pantherines and the Sabercat Smilodon fatalis. PLoS One, 7, e48352.
- Fandos P, Vigal CR (1993): Sexual dimorphism in size of the skull of Spanish ibex Capra pyrenaica. Acta Theriol, 33, 103-111.
- Groves CP (1983): Notes on the gazelles IV: the Arabian gazelles collected by Hemprich and Ehrenberg. Z Saugetierkd, 48, 371–381.
- Hasan HA (2017): Three dimensional computed tomography morphometric analysis of the orbit in Iraqi population. Int Med J, 24, 147-149.
- Hayatgheib D, Karami M, Farahmand H, et al (2011): Morphological study and comparison of western and eastern goitered gazelle (Gazella subgutturosa: Guldenstaedt, 1780) populations in Iran. Int J Environ Res, 5, 225-232.
- **10.** Ince NG, Demircioglu I, Yılmaz B, et al (2018): Martılarda (Laridae spp.) cranium'un üç boyutlu modellemesi. Harran Univ Vet Fak Derg, **7**, 98-101.

- **11. Jaslow JR** (1989): Sexual dimorphism of cranial suture complexity in wild sheep (Ovis orientalis). Zoo J Linnean Soc, **95**, 273-284.
- 12. Kamal S (2006): Morphological and craniometrical studies on the skull of Kagani goat (Capra hircus) of Jammu region. Int J Morphol, 24, 449-455.
- **13.** Kanchan T, Krishan K, Gupta A, et al (2014): A study of cranial variations based on craniometric indices in a South Indian population. J Craniofac Surg, **25**, 1645-1649.
- 14. Karimi I, Onar V, Pazvan G, et al (2011): The cranial morphometric and morphologic characteristics of Mehraban sheep in Western Iran. Glob Vet, 6, 111-117.
- **15. Kataba A** (2010): Biometric and morphologic characteristics of the skull of the Gwembe Valley Dwarf Goat (Capra Hircus) breed of Zambia. MSc, University of Zambia, Lusaka, Zambia.
- **16.** Kranioti EF (2019): Radiometry versus osteometry in sex assessment: a study of the Cretan radius. Aust J Forensic Sci, **51**, 135-148.
- **17. Krupa P, Krsek P, Cernochova P, et al** (2004): 3D real modelling and CT biomodels application in facial surgery. In: Neuroradiology, Springer, Berlin.
- **18. Leslie DM** (2010): *Procapra picticaudata (Artiodactyla: Bovidae)*. Mamm Species, **42**, 138-148.
- Mallon DP, Kingswood SC (2001): Antelopes. Part 4: North Africa, the Middle East, and Asia. Global Survey and Regional Action Plans. SSC Antelope Specialist Group. IUCN, Gland, Switzerland and Cambridge, UK.
- **20. Nomina Anatomica Veterinaria** (2017): International committee on veterinary gross anatomical nomenclature

(ICVGAN), Published by the Editorial Committee, Hannover.

- 21. Özcan S, Aksoy G, Kürtül İ, et al (2010): A comparative morphometric study on the skull of the Tuj and Morkaraman sheep. Kafkas Univ Vet Fak Derg, 16, 111-114.
- 22. Özkurt A (2002): Üç boyutlu örneksel veriden yüzey modeli üretimi. DEUFMD, 4, 27–36.
- Páral V, Tichý F, Fabiś M (2004): Functional structure of metapodial bones in Cattle. Acta Vet Brno, 73, 413-420.
- 24. Parés I, Casanova PM, Kamal S, et al (2010): On biometrical aspects of the cephalic anatomy of Xisqueta sheep (Catalunya, Spain). Int J Morpol, 28, 347-351.
- 25. Parés-Casanova PM (2014): Osteometric study of the Rasquera White goat. J Appl Anim Res, 42, 177-185.
- **26.** Tecirlioğlu S (1983): Sırtlan ve köpeğin iskelet kemikleri üzerinde makro-Anatomik araştırmalar. Ankara Univ Vet Fak Derg, **30**, 149-166.
- 27. Teo EC, Holsgrove T, Haiblikova S (2017): 3D Morphometric analysis of human vertebrae C3-T3 using CT images reconstruction. J Spine, 6, 391.
- 28. Thouless CR, Al Bassri KA (1991): Taxonomic status of the Farasan island gazelle. J Zool Lond, 223, 151-159.
- **29.** Von Den Driesch A (1976): A guide to the measurement of animal bones from archaeological sites. Peabody Museum Bulletin 1. Cambridge, MA, Harvard University.
- **30.** Wronski T, Sandouka M, Plath M, et al (2010): Differences in sexual dimorphism among four gazelle taxa (Gazella spp.) in the Middle East. Animal Biol, **60**, 395-412.
- **31.** Zhu L. (2012): Craniometrical studies on the skull of Tibetan Gazelle (Procapra Picticaudata). Int J Morphol, **30**, 196-198.

Ankara Üniv Vet Fak Derg, **67**, 169-174, 2020 DOI: 10.33988/auvfd.603928

## First molecular data on the validity of *Myxobolus ichkeulensis* (Cnidaria: Myxosporea) from *Mugil cephalus* (Mugilidae) in Turkish waters

#### Cenk Soner BÖLÜKBAŞ<sup>1,a</sup>, Gökmen Zafer PEKMEZCİ<sup>2,b,⊠</sup>, Banu YARDIMCI<sup>2,c</sup>, Melek ÖZPIÇAK<sup>3,d</sup>, Savaş YILMAZ<sup>3,e</sup>, Nazmi POLAT<sup>3,f</sup>

<sup>1</sup>Ondokuz Mayıs University, Faculty of Veterinary Medicine, Department of Parasitology, Samsun; <sup>2</sup>Ondokuz Mayıs University, Faculty of Veterinary Medicine, Department of Aquatic Animal Diseases, Samsun; <sup>3</sup>Ondokuz Mayıs University, Faculty of Sciences and Arts, Department of Biology, Samsun, Turkey.

> <sup>a</sup>ORCID: 0000-0002-4863-696X; <sup>b</sup>ORCID: 0000-0002-7791-1959; <sup>c</sup>ORCID:0000-0001-7232-554X; <sup>d</sup>ORCID:0000-0003-3506-4242; <sup>e</sup>ORCID: 0000-0003-2859-4886; <sup>f</sup>ORCID: 0000-0001-9785-9927

⊠Corresponding author: zpekmezci@omu.edu.tr Received date: 08.08.2019- Accepted date: 04.11.2019

**Abstract:** To date, there is no report on molecular characterisation of *Myxobolus ichkeulensis* in *Mugil cephalus* (Mugilidae) in Turkish marine waters. The aim of the current research was to provide the first molecular data on *M. ichkeulensis* from *M. cephalus* in Turkish Black Sea coasts. A total of 40 *M. cephalus* specimens were examined for the presence of *M. ichkeulensis* in the Turkish Black Sea coasts during January 2017 and April 2017. In the present study, *M. ichkeulensis* was identified based upon the myxospore morphology, host and tissue specificity, and SSU rRNA sequence. Phylogenetic analysis using neighbor-joining also revealed that our isolate (#GZP-2018-Samsun1) was clustered with other *Myxobolus* species that infect *M. cephalus*. The current study includes the morphological and histopathological descriptions and first molecular data on *M. ichkeulensis* in *M. cephalus* in Turkish marine waters. *M. ichkeulensis* was also reported from the *M. cephalus* in the coast of the Black Sea in Turkey for the first time. Moreover, our current genetic data was recorded as the new valid SSU rRNA sequence (MH374629) for *M. ichkeulensis* in the GenBank database.

Keywords: Histopathology, morphology, Myxobolus ichkeulensis, SSU rRNA, Turkey

# Türkiye sularında *Mugil cephalus*'tan (Mugilidae) *Myxobolus ichkeulensis*'in (Cnidaria: Myxosporea) geçerliliğine ilişkin ilk moleküler veriler

Özet: Bugüne kadar Türkiye karasularındaki *Mugil cephalus* (Mugilidae)'da *Myxobolus ichkeulensis* türünün moleküler karakterizasyonu ile ilgili kayıt yoktur. Bu araştırmanın amacı Türkiye'nin Karadeniz kıyılarında *M. ichkeulensis* türünün *M. cephalus*'lardaki varlığını hakkında ilk moleküler verileri sağlamaktır. Ocak 2017 ile Nisan 2017 tarihleri arasında Türkiye'nin Karadeniz kıyılarından *M. ichkeulensis* türünün varlığı için toplam 40 adet *M. cephalus* incelendi. Bu araştırmada *M. ichkeulensis* türü spor morfolojisi, konak ve doku spesifitesi ile SSU rRNA sekansına dayanarak teşhis edildi. İzolatımız (#GZP-2018-Samsun1) neighbor-joining metodu kullanılan filogenetik analizde *M. cephalus*'ları enfekte eden diğer *Myxobolus* türleri ile kümelendi. Mevcut araştırma, Türkiye sularında *M. cephalus*'ta *M. ichkeulensis* türünün geçerliliği üzerine morfolojik ve histopatolojik tanımlamalar ile ve ilk moleküler verileri içermektedir. Türkiye'nin Karadeniz kıyılarından ilk kez *M. cephalus*'ta *M. ichkeulensis* türü de bildirildi. Ayrıca mevcut genetik verilerimiz GenBank veri tabanındaki *M. ichkeulensis* türü için yeni geçerli SSU rRNA dizisi (MH374629) olarak kaydedildi.

Anahtar sözcükler: Histopatoloji, morfoloji, Myxobolus ichkeulensis, SSU rRNA, Türkiye

#### Introduction

Thousands of myxosporean parasites are known to cause diseases in various marine and freshwater fish (18, 19). The genus *Myxobolus* Bütschli, 1882 is the largest group of Myxobolidae contains over 850 species described (7, 8). Based on spore morphology, host/ tissue tropism with the molecular marker are especially provided

for identification of a new or existing myxozoa species and re-description of incompletely described species (11, 20). *Myxobolus ichkeulensis* was firstly described from the grey mullet *Mugil cephalus* in Ichkeul lagoon in Tunisia by Bahri and Marques (4). Later, there have been few reports on *M. ichkeulensis* in *M. cephalus* from Lake Ichkeul (Tunisia), Ebro River Delta (Spain), Baje de Gorée (Senegal), Black and Azov Seas (Crimea, Ukraine), Camlık Lagoon (north-eastern Mediterranean, Turkey) and Japan Sea (3, 9, 23, 24, 28, 31). Whereas, this *Myxobolus* species has been reported in different marine sources of the world, there were only two SSU rRNA sequences of *M. ichkeulensis* molecularly characterized and submitted to the GenBank with the accession numbers: AF378337 and AY129315 (3, 14).

Up to now, only four *Myxobolus* species have been morphologically identified and reported in *M. cephalus* from Turkish marine waters: *M. episquamalis*, *M. exiguus*, *M. ichkeulensis* and *M. muelleri* (2, 6, 23, 30). Whereas those species were morphologically described, there is no detailed molecular and histopathological data of *M. ichkeulensis* in *M. cephalus* in Turkish marine waters. The aim of the current research was to provide first molecular data on the validity of *M. ichkeulensis* from *M. cephalus* in Turkish marine waters.

#### **Material and Methods**

Sampling, morphological and histopathological examination: For this study, ethics committee approval was not needed because no handling of live fish specimens were involved. A total of forty freshly caught specimens of *M. cephalus* were periodically purchased from commercial fishermen at Kızılırmak Delta, Samsun coast located by the Black Sea, Turkey (41°44'04.2"N 35°57'23.0"E) in the period between January 2017 and April 2017. After purchase and transportation to the laboratory, M. cephalus were examined for М. ichkeulensis infections under a dissecting microscope (18). After whitish plasmodia were detected in the gills, plasmodia were isolated with a needle and opened on a slide. Infected gills containing mature plasmodia were fixed in 10% formaldehyde and embedded in paraffin. Paraffin blocks were cut into 5  $\mu$  slices using microtome and stained with Hematoxylin-Eosin (H&E). The position of the plasmodia in the gills was classified to Molnár (21). Some fresh spores were prepared in glycerine-jelly into the slide for morphological examination. Subsamples of fresh spores were preserved in absolute ethanol for molecular identifications. Myxospores were morphologically examined as previously reported Lom and Arthur (17) by measuring 20 freshly isolated mature spores in reference slide preparations. The myxospores were photographed and measured by Nomarski DIC optics connected to a digital camera. All measurements are presented in micrometres (µm) with mean and range in parentheses.

**DNA extractions and PCR analysis:** Myxobolus spores were centrifuged at low speed, suspended in digestion solution and incubated at 56°C overnight. DNA was then extracted using a commercial DNA extraction kit

(GeneJET Genomic DNA Purification Kit, ThermoFisher Scientific) according to the procedure recommended by the manufacturer. Nested PCR assay was used for identification of Myxobolus species. The small subunit ribosomal RNA (SSU rRNA) gene (~1900 bp) was amplified by first PCR using ERIB1 and ERIB10 primer pairs (5). Fifty µl PCR reactions were contained 20-50 ng DNA, 2X Hot start PCR master mix, 0.4 µM of each primer. Amplification of first PCR conditions were: 30 sec at 95°C, 50 sec at 43°C, and 120 sec at 72°C for 30 cycles, and a 10 min extension at 72 °C. Then, second PCR were carried out in a final volume of 50 µl, which contained 1 ul of the first PCR amplicon, 2X hot start PCR master mix, 0.4 µM of each primer. The ~900 bp amplification products using the MyxospecF-MyxospecR primer sets (12) were run with 30 sec at 95°C, 50 sec at 40 °C, 90 sec at 72°C for 35 cycles, and products were subjected to a final extension at 72 °C for 10 min. After amplification, PCR amplicons were electrophoresed on 1% agarose gel in a TBE buffer. Second PCR amplicons were sequenced both directions with MyxospecF-MyxospecR primer pairs by Sanger method (Macrogen, Netherlands).

Phylogenetic analyses: The obtained sequences were controlled by Vector NTI Advance 11.5 (Invitrogen, USA) using phred values. Then, sequences were de novo assembled and edited with using Contig Express (NTI Advance 11.5, Invitrogen, USA) and the created sequences were compared with GenBank accessions using the BLAST research (1). The SSU rRNA sequences were aligned with other known Myxobolus species from M. cephalus in previous studies (3, 14, 15, 28). Sequence aligments were performed with ClustalW in MEGA 7.0 (29) and adjusted manually. Phylogenetic tree was constructed using neighbor-joining (NJ) analysis in MEGA 7.0 (16). The Kimura two-parameter (K2P) model was used in the analysis. The species Ceratomyxa shasta was chosen as the out-group. The mugilid infecting Myxobolus sequence sets were built with 1000 bootstrap replications for the NJ reconstruction (10). Bootstrap values  $\geq$  70 were considered well supported (13).

#### Results

Whitish cyst-like plasmodia were detected macroscopically along the conjunction line of the gill filaments and arches of *M. cephalus*. We concluded that our isolate (#GZP-2018-Samsun1) is *M. ichkeulensis* based on the spore morphology, biological traits (host/organ specificity and tissue tropism) and molecular data.

#### Taxonomy of Myxobolus ichkeulensis (4)

*Host:* Grey mullet, *Mugil cephalus* (Mugilidae)

*Locality:* Kızılırmak Delta, Samsun, Turkey (41°44'04.2"N 35°57'23.0"E)



**Figure 1.** Microscopic photographs of *M. ichkeulensis* infecting the gill arch of *M. cephalus*. **A.** Fresh myxospores of *M. ichkeulensis* in glycerine-jelly, scale: 10 µm. **B.** Myxospore of *M. ichkeulensis* inside plasmodium (p), histopathological section, H&E staining, scale: 10 µm. **C.** *M. ichkeulensis* plasmodium (p) located in the connective tissue elements of the gill arch, histopathological section, H&E staining, scale: 100 µm. **D.** A large plasmodium (p) of *M. ichkeulensis* in the gill arch, histopathological section, H&E staining, scale: 500 µm.

*Site of infection:* Plasmodia were observed macroscopically as whitish cysts along the conjunction line of the gill filaments and arches.

*Type material:* Reference glycerine-jelly and histopathological sections were deposited in the laboratory. The SSU rRNA sequence of *M. ichkeulensis* was recorded in GenBank as MH374629.

*Prevalence of infections:* 12.5 % (5 out of 40), 12 to 18-cm-sized fish

*Myxospores:* The myxospores were round or spherical. The spores were 13.3 (12.06 to 13.72)  $\mu$ m long (n = 20), 11.42 (10.5 to 12.4)  $\mu$ m wide (n = 20), and 8.24 (7.75 to 8.53)  $\mu$ m thick (n = 10). The two polar capsules were oval, equal in size, 5.97 (5.7 to 6.65)  $\mu$ m long (n = 20) and 3.81 (3.4 to 4.18)  $\mu$ m wide (n = 20), and their posterior end reached half the length of the spore. Eight to ten sutural edge markings were easily observed (Figure 1A-B).

*Histopathological findings:* 250 to 2000  $\mu$ m in diameter plasmodia were found. The histopathological analysis revealed the development of the cyst-like plasmodia as gill arch type. Moreover, plasmodia were located in the connective tissue elements of the gill arch (Figure 1C-D).

*Molecular data:* No intraspecific nucleotide variability within three isolate of *M. ichkeulensis* from the Black Sea were observed in the SSU rRNA sequences. A BLAST search indicated that SSU rRNA sequence of our isolate GZP-2018-Samsun1 (MH374629) from *M. cephalus* showed 99.05% similarity to *Myxobolus* sp. voucher Spain6-tp (MF118764), which is also a tail-infecting species and identified as *M. ichkeulensis*, and shared 98.82% similarity with that of *M. ichkeulensis* (AF378337; AY129315). For this reason, our *Myxobolus* isolate (GZP-2018-Samsun1) thus identified molecularly to belong to *M. ichkeulensis*.

#### **Discussion and Conclusion**

The flathead grey mullet *M. cephalus* (Mugilidae) is a cosmopolitan coastal fish species distributed worldwide. Several myxosporean parasites have been reported as serious pathogens of mugilid fish species (22, 31). Moreover, a great number of myxosporean species were recorded in *M. cephalus* among other mugiliform fish species. To date, thirty six species of myxosporeans have been reported and amongst them are *M. muelleri*, *M. ichkeulensis*, *M. spinacurvatura*, *M. exiguus*, *M. parvus* and *M. episquamalis* are only six cosmopolite species in *M. cephalus* from worldwide (31).

*M. ichkeulensis* was firstly described in *M. cephalus* by Bahri and Margues (4) based on traditional criteria, including tissue tropism and detailed light and electron microscopic examination of spore morphology and subsequently Bahri et al. (3) provided a supplemental data on *M. ichkeulensis* from the host type with molecular data of the SSU rDNA sequence (AY129315). Within the current study, the SSU rDNA sequences of our isolate

(#GZP-2018-Samsun1) showed 98.82% identity with reference sequence of *M. ichkeulensis* (AY129315). Therefore, we molecularly identified our Myxobolus species as M. ichkeulensis. Currently, based on spore morphology, host/organ specificity and tissue tropism with the molecular marker are mainly useful for new myxosporean species and re-description of insufficiently described species (3, 25-28, 31). Thus, these approaches combined (morphology, biological traits, and molecular markers) were used for the validation of M. ichkeulensis in M. cephalus from Turkish waters for the first time. Moreover, this is also first report of *M. ichkeulensis* in *M.* cephalus from the coast of the Black Sea in Turkey. Furthermore, the phylogenetic tree showed that our isolate was clustered with M. ichkeulensis species previously known to be sequenced from *M. cephalus* (Figure 2). A comparison of *M. ichkeulensis* Bahri and Marques (4), spore morphometric data isolated from M. cephalus at different geographical areas is presented in Table 1.



0.05

**Figure 2.** Phylogenetic tree generated by NJ analysis of the SSU rRNA sequences of *M. ichkeulensis* and other *Myxobolus* species infecting mugiliform.

Numbers at nodes indicate the bootstrap values. Ceratomyxa shasta was used as the out group.

•		ŭ )		1		
Spore length	Spore width	Spore thickness	Polar capsule length	Polar capsule width	Locality	Reference
13.5 (13-14)	12.5 (12-13)	-	5.5 (5-6)	4.2 (4-4.3)	Tunisia: Ichkeul Lagoon	Bahri and Marques (4)
13.5±0.54	12.5±0.54	-	5.5±0.54	4.25±0.27	Tunisia: Ichkeul Lake	Bahri et al. (3)
13.32 (12.49-14.15)	12.24 (11.58-12.9)	7.69 (7.24-8.14)	6.38 (6-6.76)	4.18 (3.95-4.41)	Turkey: Mediterranean coast, Camlık Lagoon	Ozak et al. (23)
13.13 (12.06-13.72)	11.42 (10.5-12.4)	8.24 (7.75-8.53)	5.97 (5.7-6.65)	3.81 (3.4-4.18)	Turkey: Black Sea coast, Kızılırmak Delta	Present study

Table 1. Comparison of spore morphometric data (µm) of *M. ichkeulensis* infection in *M. cephalus.* 

Myxosporean plasmodia localize in or among gill lamellae, in gill filaments and inside the gill arch cartilage (20). In the present study, histopathological sections showed M. ichkeulensis plasmodia in the connective tissue elements of the gill arch (Figure 1C-D). Myxosporean species is strictly connected to a specific tissue of the host (20). Myxosporean plasmodia may develop within the connective tissue layer in skin doublets between the fin rays (20, 21). Supportively, Myxobolus plasmodia embedding in the fin tail connective tissue was found in M. cephalus and was 99% similar to M. ichkeulensis (28). Our study and Sharon et al. (28) indicated that M. ichkeulensis in M. cephalus has an affinity for connective tissues both in the gill arch and tail fin. Moreover, small and single cysts of M. ichkeulensis reveal a basi-filamental type of plasmodial development in the *M. cephalus* (23).

In conclusion, supplementary data of histopathology and SSU rDNA analysis of *M. ichkeulensis* infecting *M. cephalus* as host type were provided in the present study for the first time from Turkish waters. Moreover, the new valid SSU rRNA sequence (MH374629) obtained from *M. cephalus* from Turkish waters has been submitted to the GenBank. This sequence can be also used to construct a phylogenetic tree with other mugiliform-infecting *Myxobolus* species.

#### Acknowledgements

A poster of this work was presented at the International Symposium on Ecology, Kastamonu, Turkey on June 20-23, 2018.

#### **Financial Support**

This research received no grant from any funding agency/sector.

#### **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### References

- 1. Altschul S, Madden T, Schaffer A, et al (1997): Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Res, 258, 3389-3402.
- Altunel FN (1983): Parasitism on mullets (Mugil spp.). J Fac Sci Ege Univ B, 1, 364-378.
- 3. Bahri S, Andree KB, Hedrick RP (2003): Morphological and phylogenetic studies of marine Myxobolus spp. from mullet in Ichkeul Lake, Tunisia. J Eukaryot Microbiol, 50, 463-470.
- Bahri S, Marques A (1996): Myxosporean parasites of the genus Myxobolus from Mugil cephalus in Ichkeul lagoon, Tunisia: description of two new species. Dis Aquat Organ, 27, 115-122.
- 5. Barta JR, Martin DS, Liberator PA, et al (1997): Phylogenetic relationships among eight Eimeria species infecting domestic fowl inferred using complete small subunit ribosomal DNA sequences. J Parasitol, 83, 262-271.
- 6. Demirkale I, Cengizler I (2016): Identification of the infested gray mullets (Mugil cephalus L. 1758) by Myxobolus ichkeulensis level in the coast of Adana. TURJAF, 4, 805-812.
- 7. Eiras JC, Molnár K, Lu YS (2005): Synopsis of the genus Myxobolus Bütschli, 1882 (Myxozoa: Myxosporea: Myxobolidae). Syst Parasitol, **61**, 1-46.
- Eiras JC, Zhang J, Molnár K (2014). Synopsis of the species of Myxobolus Bütschli, 1882 (Myxozoa: Myxosporea, Myxobolidae) described between 2005 and 2013. Syst Parasitol, 88, 11-36.
- **9.** Fall M, Kpatcha KP, Diebakate C, et al (1997): Observations sur des Myxosporidies (Myxozoa) du genre Myxobolus parasites de Mugil cephalus (Poisson, Téléostéen) du Sénégal. Parasite, **2**, 173-180.
- **10.** Felsenstein J (1985): Confidence intervals on phylogenies: an approach using the bootstrap. Evolution, 39, 783-791.
- Fiala I, Bartošová–Sojková P, Okamura B, et al (2015): Adaptive Radiation and Evolution within the Myxozoa. 69-84. In: B Okamura, Gruhl A, Bartholomew JL (Eds), Myxozoan Evolution, Ecology and Development. Springer International Publishing, Switzerland.
- Fiala I (2006): The phylogeny of Myxosporea (Myxozoa) based on small subunit ribosomal RNA gene analysis. Int J Parasitol, 36, 1521-1534.

- **13. Hillis DM, Bull JJ** (1993): An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. Syst Biol, **42**, 182-192.
- 14. Kent ML, Andree KB, Bartholomew JL, et al (2001): Recent advances in our knowledge of the Myxozoa. J Eukaryotic Microbiol, 48, 395-413.
- Kim WS, Kim JH, Oh MJ (2013): Morphologic and genetic evidence for mixed infection with two Myxobolus species (Myxozoa: Myxobolidae) in gray mullets, Mugil cephalus, from Korean waters. Korean J Parasitol, 51, 369-373.
- Kumar S, Stecher G, Tamura K (2016): MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. Mol Biol Evol, 33, 1870-1874.
- **17.** Lom J, Arthur JR (1989): A guideline for the preparation of species description in Myxosporea. J Fish Dis, **12**, 151-156.
- **18.** Lom J, Dyková I (1992): Protozoan Parasites of Fishes. Elsevier, Amsterdam.
- **19.** Lom J, Dyková I (2006): *Myxozoan genera: definition and note on taxonomy, life-cycle terminology and pathogenic species.* Folia Parasitol, **53**, 1-36.
- 20. Molnár K, Eszterbauer E (2015): Specificity of Infection Sites in Vertebrate Hosts. 295-313. In: B Okamura, Gruhl A, Bartholomew JL (Eds), Myxozoan Evolution, Ecology and Development. Springer International Publishing, Switzerland.
- **21.** Molnár K (2002): *Site preference of myxosporeans in the gill.* Dis Aquat Organ, **48**, 197-207.
- 22. Ovcharenko M (2015): *Microparasites of worldwide mullets*. Ann Parasitol, **61**, 229-239.
- 23. Ozak AA, Demirkale I, Cengizler I (2012): Two new records of Myxobolus Bütschli, 1882 (Myxozoa, Myxosporea, Myxobolidae) species from Turkey. Turk J Zool, 36, 191-199.

- 24. Pedro-Andres MB, Marques A, Gracia-Royo MP (2011): Myxosporean infection of grey mullet in the Ebro Delta: identification and ultrastructure of Myxobolus ichkeulensis Bahri & Marques, 1996 infecting the gills of Mugil cephalus L. Acta Protozool, **50**, 67-71.
- **25.** Pekmezci GZ, Yardimci B, Yilmaz S, et al (2014): Myxobolus anatolicus sp. nov. (Myxozoa) infecting the gill of Anatolian khramulya Capoeta tinca (Cyprinidae) in Turkey. Dis Aquat Organ, **109**, 213-222.
- 26. Pekmezci GZ, Yardimci B, Yilmaz S (2015): Supplementary studies and the first molecular data on Myxobolus scardinii Reuss, 1906 (Myxozoa: Myxosporea) infecting the gill filaments of rudd, Scardinius erythrophthalmus (L.). Parasitol Res, 114, 3619-3625.
- 27. Rocha S, Azevedo C, Oliveira E, et al (2019): *Phylogeny* and comprehensive revision of mugiliform-infecting myxobolids (Myxozoa, Myxobolidae), with the morphological and molecular redescription of the cryptic species Myxobolus exiguus. Parasitology, **146**, 479-496.
- Sharon G, Ucko M, Tamir B, et al (2019): Co-existence of Myxobolus spp. (Myxozoa) in gray mullet (Mugil cephalus) juveniles from the Mediterranean Sea. Parasitol Res, 118, 159-167.
- **29.** Thompson JD, Higgins DG, Gibson TJ (1994): *CLUSTALW: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice.* Nucleic Acids Res, **2**, 4673-4680.
- **30.** Umur S, Pekmezci GZ, Beyhan YE, et al (2010): First record of Myxobolus muelleri (Myxosporea: Myxobolidae) in flathead grey mullet Mugil cephalus (Teleostei, Mugilidae) from Turkey. Ankara Univ Vet Fak Derg, **57**, 205-207.
- **31. Yurakhno VM, Ovcharenko MO** (2014): Study of Myxosporea (Myxozoa), infecting worldwide mullets with description of a new species. Parasitol Res, **113**, 3661-3674.

Ankara Üniv Vet Fak Derg, **67**, 175-184, 2020 DOI: 10.33988/ auvfd.632957

### Influence of different water access status of suckling calves on performance, water intake behavior and some blood metabolites

#### Mohsen ZAREI<sup>1,a</sup>, Jamal SEIFDAVATI<sup>1,b,⊠</sup>, Gholam Reza GHORBANI<sup>2,c</sup>, Hosein ABDI-BENEMAR<sup>1,d</sup>, Reza SEYEDSHARIFI<sup>1,e</sup>, Abdolhamid KARIMI<sup>3,f</sup>

<sup>1</sup>University of Mohaghegh Ardabili, Faculty of Agriculture and Natural Resources, Department of Animal Sciences, Ardabil; <sup>2</sup>Isfahan University of Technology, College of Agriculture, Department of Animal Science; <sup>3</sup>Fars Agricultural and Natural Resources Research and Education Center, Shiraz, Iran.

> <sup>a</sup>ORCID: 0000-0003-3002-8031; <sup>b</sup>ORCID: 0000-0001-6794-4450; <sup>c</sup>ORCID: 0000-0002-2136-6090; <sup>d</sup>ORCID: 0000-0001-5318-4585; <sup>e</sup>ORCID: 0000-0003-4593-2058; <sup>f</sup>ORCID: 0000-0003-4147-5734

<sup>⊠</sup>Corresponding author: jseifdavati@uma.ac.ir Received date: 14.09.2019 - Accepted date: 03.12.2019

**Abstract:** This study was conducted to determine the effects of water accessibility of pre-weaning calves during milk feeding meal on performance, water intake behavior and some blood metabolites. Forty-eight Holstein calves  $(39.2 \pm 5.8 \text{ kg} \text{ of BW})$  were assigned to 4 treatments, which consisted of different water accessibility: 1) Ad libitum water (AW), 2) Water not available (WNA), 3) Water available after milk feeding (WAM) and 4) Water available before milk feeding (WBM). Resulted showed different water accessibility changed water consumption of calves. The water intake of AW (1.03 kg/d) and WBM (1.07 kg/d) treatments were higher than others. Starter intake of WBM decreased (P<0.05) as compared with others. Calves of WBM had higher (P<0.05) activity of serum AST. Blood Cl and K did not show the difference (P<0.05) among treatments. Serum TAC levels were higher (P<0.05) in WNA and WAM calves. Serum protein and HDL in WNA calves were higher (P<0.05) than others. These groups had higher (P<0.05) serum Na concentration than others. The different water accessibility of calves had no significant effects on health score, respiratory, fecal score and growth parameters. Treatments did not affect behaviors parameters. But the number of visiting water bucket of AW and WBM were higher than other groups. Acetic acid and propionic acid of AW calves significantly increased (P<0.05) as versus other treatments. Drinking water around milk feeding meal change water consumption, performance and improve some blood metabolites and rumen parameters of calves but did not influence growth parameters.

Keywords: Blood metabolites, calves, growth factor, performance, water intake.

#### Introduction

Water is an important nutrient in dairy production (26). According to previous research, 69.5 to 71.6% of the calf body weight is made up of water (6). Too often, it is assumed that if a calf is being fed a liquid diet, its needs for water will be satisfied (27). According to the German enactment of animal welfare and farming of animals (33) calves over 2 weeks of age have to be allowed ad libitum access to water. Feeding calves ad libitum milk during the pre-weaning period is one method to provide more water to the calves at an early stage (17). However, feeding greater amounts of milk have been shown to decrease post-weaning calf growth and decrease starter intake (3). However, milk is the primary source of water during the first few weeks of the calf's life, but represented a decreasing percentage of the total water intake as the calf became older (27). Neonate calves could consume a considerable amount of water via whole milk or milk

replacer but it's not clear that amount alone would fulfill the water requirements to support growth and development (35). The drinking water requirement of calves appears to be insignificant when separated from liquid feed intake (35). Milk-fed calves drink little water until weaning, at which time water intake increases markedly (15), and water intake tends to follow the intake of solid food (19). Calves offered supplemental water gained more body weight and consumed more calf starter compared to calves without access to free water (19). Water intake of Holstein bull calves through 56 days of age was measured in 6 different trials over two years (29). In these studies, water intake was very closely correlated with starter intake. As calves consumed more starter diet they drank more water. The study, covering 30 years (27) found a significant correlation of both live-weight gain and calf starter intake with water intake prior to weaning at 5 weeks. In addition, greater water consumption has

been shown to increase early starter intake (5). Studies have shown that consumption of solid feed in starter diets can contribute to optimized development of the rumen in calves, thereby leading to greater potential for early weaning (5). During the pre-weaning period, when calves consume whole milk or milk replacer it bypasses the rumen and goes to the abomasum via the oesophageal groove. This process also prevents a large portion of consumed water from entering the rumen and may influence bacterial growth and the start of rumen fermentation. Therefore, providing clean fresh drinking water to calves is essential for the rumen microbiota development (7). Wickramasinghe (35) reported that offering drinking water to calves from birth itself appeared to offer positive benefits even in systems promoting a large amount of liquid feed (whole milk or milk replacer) intake. Producers also seem to be hesitant to offer water to new-born calves, assuming it would cause diarrhea (4).

Overall, a lack of scientific evidence on drinking water intake of new-born calves and its effect on their performance likely encumbers the effectiveness of efforts promoting the notion of offering drinking water to newborn calves at birth. Depending on the improved starter intake and weight gain in report Kertz et al. (19), it is often hypothesized that drinking water would enhance rumen development, thus increasing nutrient availability for growth and development of calves. However, no systematic attempt has been made to study this link between water intake and nutrient availability to the animal. This also brought attention to the fact that many dairy farmers wait an hour before and after milk feeding meal to offer drinking water to suckler calves even though offering water a full day is the recommendation. Water ideally should be available at all times to young calves, but as minimum warm water should be offered after feeding and midday in cold climates (5). The hypothesis of this study is the question that "Does water drinking around milk meal affects calf health and performance?" Thus, the objectives of our experiment were to determine the effects of different water access status of suckling calves on performance, water intake behavior and some blood metabolites.

#### **Materials and Methods**

Animals, housing and diets: The research protocol was approved by the Animal Care and Use Committee of the Iranian Council of Animal Care (1995). Forty-eight Holstein calves ((3 d of age;  $39.2 \pm 5.8$  kg of body weight (BW)) were used in this study. The experiment was carried out at Mohandesiye Dairy cattle farm, Esfahan province, Iran. This study was conducted in late 2017 and early 2018. The month average temperature was decreased monthly. The weather data of Esfahan were for the average temperatures (°C) (19.2, 11.2 and 6.4), Maximum

temperature (°C) (31.6, 26.8 and 23.4), Minimum temperature (°C) (6, -2.4 and -4.8) and the average wind speed (m/s) (1.9, 1.1 and 0.9) during October, November and December months (2017), respectively.

The Mohandesiye Dairy cattle farm in Esfahan is one of the biggest farms in Esfahan that had almost 1500 cow. Total parturition days for this study to select and collect the 48 calf were about one week or 7 days. Calves were separated from their mothers within 24 h of birth, weighed, moved into individual pens  $(1.2 \times 1.8 \text{ m}; \text{ width} \times \text{depth})$  where they were fed 10% colostrum of their BW for the first 3 d. The pens were cleaned every 2 d by removing all bedding and replacing it with fresh sawdust bedding. Calves were managed according to the standard operating procedures of this research station, in accordance with guidelines set by the Iranian Council of Animal Care (16).

Calf starter was offered ad libitum, and intake was recorded daily, beginning on day 4. The basal diet was formulated to meet Nutrient Requirements of Dairy Cattle (NRC) (27) requirements for calf nutrients. The ingredients, nutrient composition of the basal diet and drinking water quality are shown in Table 1. Chopped dried wheat straw and alfalfa hay were used after 20 days age of calves in the study. The forage was chopped using a total mixed ration mixer (Loewen Horizontal Mixer, Loewen Welding & Manufacturing Ltd., Matsqui, BC, Canada) at 19,000 rpm for 60 min. Three additional buckets were placed in the middle of the area where the new-born calves were housed and also weighted every 24 hours to calculate the evaporation of water. The daily water intake of a calf was calculated by the difference in weight of the buckets of each calf minus average of three bucket evaporation. All calves were fed the same amount of whole milk. Calves were bucket-fed 4.5 L/d of whole milk divided into 3 feedings, at approximately 0800, 1630 and 0030 h from birth until 30 d of age. From d 31 to 50, calves were fed 8 L/d, using the same procedure as described above. Milk volume was reduced by 20 % that means calves were fed 6.4 L/d from d 50 to 60, with milk weaning occurring on d 60. Therefore, the whole experiment period was sixty days, and for fed colostrum in the first 3 days of life, the main experiment period began on the fourth day of calf birth.

On the 4<sup>th</sup> day, the calves were randomly assigned to 4 treatments: 1) water were ad libitum (AW) 2) Inaccessibility to water an hour before and an hour after milk feeding or water not available (WNA) 3) Inaccessibility before and access to water after milk feeding milk or water available after milk feeding (WAM) and 4) Accessibility before and inaccessibility to water an hour after milk feeding or water available before milk feeding (WBM). A total of 6 water samples were collected and sent directly to a commercial laboratory for analysis (Zagros LAB water quality test, Fars, Shiraz, Z.A.F).

Diet composition	% of DM	Water quality	Mean±SEM
Corn grain ground	27	Number of samples	6
Barley grain ground	35.5	EC (µs/cm)	$2386.00 \pm 36.97$
Rumen growth factor (RGF)	14	$T.D.S^3$ (mg/L)	1538.27±22.85
Soybean meal, 45% CP	10	PH	7.27±0.11
Whole soybean (full fat)	0.8	Nitrate(NO <sub>3</sub> ) (mg/L)	83.25±1.9
Dicalcium phosphate	0.8	Nitrite(NO <sub>2</sub> ) (mg/L)	$0.014\pm0.004$
Sodium bicarbonate	0.6	Phosphate (PO4 <sup>3-</sup> ) (mg/L)	0.03±0.07
Magnesium oxide	0.4	Carbonate(CO <sub>3</sub> <sup>2-</sup> ) (meq/L)	$0.00\pm0.00$
Salt	0.7	$HCO^{3-}$ (meq/L)	5.00±0.3
Vitamin permix <sup>1</sup>	1.5	$Cl^{-}$ (meq/L)	11.50±1.05
Mineral permix <sup>1</sup>	1.5	SO4 <sup>2-</sup> (meq/L)	$6.00 \pm 0.8$
Nutrient composition		Sum Anion	22.50±2.4
ME, <sup>2</sup> Mcal/kg	3.03	$Ca^{2+}$ (meq/L)	7.50±1.7
NEg, <sup>2</sup> Mcal/kg	1.73	$Mg^{2+}$ (meq/L)	2.00±0.06
DM	90	Na <sup>+</sup> (meq/L)	15.58±1.16
СР	22.7	$K^+$ (meq/L)	0.12±0.09
NDF	16.4	Sum Cation	25.20±1.84
ADF	8.4	Escherichia coli (cfu/mL)	$ND^4$
Lipid	2.8		
Ash	8.0		
Ca <sup>2</sup>	0.78		
P <sup>2</sup>	0.55		

**Table 1.** Chemical composition of diet and drinking water quality

<sup>1</sup> Contained per kilogram of supplement: 250,000 IU of vitamin A, 50,000 IU of vitamin D, 1,500 IU of vitamin E, 2.25 g of Mn, 120 g of Ca, 7.7 g of Zn, 20 g of P, 20.5 g of Mg, 186 g of Na, 1.25 g of Fe, 3 g of S, 14 mg of Co, 1.25 g of Cu, 56 mg of I, and 10 mg of Se.

<sup>2</sup> Calculated from NRC (27).

<sup>3</sup> Total dissolved solids based on gravimetric analysis.

<sup>4</sup> Not detected.

The behavioral data recording methods and their description: The ingesting water behavior was defined as the calf is ingesting water by drinking from a bucket and the ingesting grain behavior was described as the calf is consuming grain from a bucket. Also, the ruminating activities were explained as the calf is ruminating. The standing status was recorded as the calf is standing with all 4 legs on the ground. The lying in bed position was noted as the calf is resting on the ground; the head may be supported or unsupported by the neck. The object play was monitored as the calf is standing; butting head against milk or water buckets or hutch in a playful manner. The selfgrooming was observed as the calf's tongue is out of its mouth and in contact with its own body. The non-nutritive sucking was recorded the calf's tongue is out of its mouth and is in contact with or biting any fixtures of the pen; may include bucket if milk is not available at the time of observation. Other behaviors were described as the calf is urinating, defecating, or performing another behavior not described. All observations were by the same person, who was blind to treatment group's allocation to minimize possible bias in data calves. We observed calf behavior for 6 80-min periods every five minutes one by one at before or after approximately 0800, 1630 and 0030 h from after first 4 days of birth to 60 days of life into 3 calves' meals. Therefore, the visits were 80-min before and 80-min after milk meal in 3 times a day accordingly, means total visits were 8 hour a day that is one replicate. The observations replicated three times.

**Data collection and sampling:** Starter intake and orts were recorded daily on an individual basis. Feed and refusals were sampled, mixed thoroughly, and ground to pass a 1-mm screen in a Wiley mill (Ogawa Seiki Co., Ltd., Tokyo, Japan). Standard methods were used for determination of DM (dry matter), CP (crude protein), NDF (neutral detergent fiber), ADF (acid detergent fiber), fat and ash as described in AOAC (1). Calves were weighed every 10 d throughout the study. Structural growth measurements of body length, withers height, hip height, heart girth and hip width were recorded on basis (cm) in two periods including d 4 and 50 of the experiment according to Lesmeister and Heinrichs (24). The four body measurements were obtained on all calves as follows:

The body length was the distance from the point of the shoulders to the ischium; withers height was the distance from the floor beneath the calf to the top of the withers directly above the center of the shoulder; hip height of calves be standing straight on a level, hard surface with its head up, and to measure hip height, used a hip stick and place it between the hip bones at a 45degree angle; heart girth was measured as the minimal circumference around the body immediately behind the front shoulder; hip width was the widest point at the center of the stifle. On the same day the body weights were also obtained. All linear measurements were obtained with specially designed aluminum calipers; or a plastic-coated fiber tape measure available commercially. All measurement devices were in metric graduations (cm). Feed efficiency was calculated according to Khan et al. (20). Fecal scoring was performed every day in the morning (08.00) according to Wisconsin-Madison calf health scoring chart as follows: 0=Normal; 1=Semiformed, pasty; 2=Loose, but stays on top of the bedding; 3=Water, sifts through the bedding. The health criteria of the calves were observed using the procedure described in Heinrichs et al. (14) and was monitored using the procedure described in Heinrichs et al. (14). Behavior was monitored by direct observation of all the calves on 2 consecutive days (on d 35 of the experiment and before weaning once per week). Calves were observed 8 h around milk feeding during the pre-weaning week. Instantaneous scans were performed for occurrences of lying, standing, water and starter intake, ruminating and non-nutritive oral behaviours (28).

Ruminal fluid samples were obtained using a stomach tube 3 h after the morning feeding on d 35, strained through 4-layer cheesecloth. A sample of the filtered ruminal fluid (5 mL) was added to 1 mL of metaphosphoric acid (250 g/L) and retained for short chain fatty acids determination. Rumen samples were analyzed for VFA by gas chromatography (Model CP-9002, Chrompack, Middelburg, Netherlands) with a 50-m (0.32 mm internal diameter) fused-silica column (CP-Wax Chrompack Capillary Column, Varian, Palo Alto, CA).

On d 35 and 50 of the study period, jugular blood samples were also collected in evacuated tubes (5 mL) 3 h after the morning feeding without adding any anticoagulant. These samples were centrifuged at  $1500 \times$  g for 20 min, and the serum samples obtained were stored at -20 °C until subsequent analyses. After unfreezing, in serum samples biochemical indicators (glucose, total protein, chloride (Cl<sup>-</sup>) albumin, cholesterol, HDL, triglycerides and activity of enzymes, ALT, AST) were measured by commercial kits (Pars Azmoon, Tehran, Iran) using an automated analyser (Biotecnica, Targa 3000, Rome, Italy). Glutathione peroxidase (GPx), total antioxidant status (TAS), NEFA, BHBA and MDA was

determined by Ransel ® kits (Randox, UK) with UV/VIS Spectrophotometer. The amounts of sodium (Na) and potassium (K) were measured by a flame photometer (Jenway 6105 Clinical, Jenway LTD Felsted England).

Statistical analysis: The corrections of the collected records were on a 35-day base. All the data collected were analyzed using the MIXED procedure (Version 8, SAS Institute Inc., Cary, NC) as a completely randomized design (32). Calf within treatment was included as a random effect that was used to test the main effect of treatment. Time was modeled as a repeated measurement for starter intake, total dry matter intake (DMI), average daily gain (ADG), feed efficiency, and fecal score within d 4-50. The data on blood metabolites, rumen parameters, BW, skeletal growth and feeding behavior were analysed for two periods while those on nutrient digestibility were analysed for one period (weaning). For BW and skeletal growth, initial values were used as the covariate. The statistical model used for the analyses was  $Y_{ijklm} = \mu + T_i + T_i$  $P_j + T \times P_{ij} + C_{kij} + l(Cov)_m + e_{ijklm}$ , where  $Y_{ijklm} =$ observation or the dependent variable, = the overall mean,  $T_i$  = the effect of treatment i,  $P_i$  = the effect of period j, T  $\times$  P<sub>ij</sub> = the effect of the interaction between treatment i and period j, Ckij = calf random effect, 1 = regression coefficient of observations on birth weight (BW) or skeletal growth as a covariate, and e<sub>ijklm</sub> = random residual effect. Values were presented as least square means. Treatment differences were declared at  $P \leq 0.05$ , with trends towards significance considered at  $P \le 0.15$ .

#### Results

It is evident that different water accessibility around calf milk meal changed water consumption of calves (Table 2). AW and WBM significantly increased water intake of calves as compared to other treatments. Also, the water intake of calves increased with age (P<0.05). Results of different calves' water accessibility showed that the calves in treatments of AW and WBM had higher water intake (respectively AW=1.03 kg/d and WBM=1.07 kg/d). The mean daily water intakes of WNA and WAM treatments decreased and were 0.84 kg/d and 0.84 kg/d, respectively. In this study, AW calves in the second decades (11 to 20 d) drank 1.26 kg/d water, that significantly higher than other groups (P<0.05). During decade 3, water intake of both AW (0.95 kg/d) and WBM (0.96 kg/d) treatments increased compared with other groups. In the 4 decades age of calves, WBM groups had more water intake (1.10 kg/d) than other treatments. The water intake of calves in decade 5, did not differ among treatment (P>0.05). In the last decade, water intake of the WAM group significantly decreased compared with other treatments (P<0.05).

Starter intake of calves is presented in Table 2. Starter intake affected by water accessibility treatments. Calves offered ad libitum water consumed an average of 39% more water than other calves in the second period (P<0.001). At third- and fourth-decade age calve AW and WBM treatments had higher water intake as compared with WNA and WAM treatments. Starter intake did not significantly difference between treatments.

The WBM treatments significantly reduced calves body weight rather than other treatments. Also, the ADG of WBM treatment was the lowest (0.54 kg/d). The result showed that water accessibility of calves before milk feeding had a negative effect on performance. According to Table 2 feed efficiency of calves did not affect by treatments.

**Table 2.** Water intake and starter intake, performance, feed efficiency and ADG of calves (n = 52) with different water accessibility during the preweaning period (Mean  $\pm$  SEM)

Itom		Treatment				Р		
Item	AW	WNA	WAM	WBM	SEM	Treat	Time	Treat×time
Water intake (kg/d)								
Whole period	1.03±0.06 <sup>a</sup>	$0.84 \pm 0.04^{b}$	$0.84 \pm 0.05^{b}$	$1.07 \pm 0.07^{a}$	0.01	0.01	0.01	0.29
1-10 d	$0.94 \pm 0.06$	$0.86 \pm 0.06$	$0.79 \pm 0.06$	$0.89 \pm 0.06$	0.51	0.24	-	-
11-20 d	$1.26 \pm 0.06^{a}$	$0.76 \pm 0.06^{\circ}$	$0.95{\pm}0.06^{bc}$	$0.98 \pm 0.06^{b}$	0.06	0.01	-	-
21-30 d	$0.95 \pm 0.05^{\mathrm{a}}$	$0.69{\pm}0.05^{\rm b}$	$0.78{\pm}0.05^{\rm b}$	$0.96 \pm 0.05^{a}$	0.05	0.01	-	-
31-41 d	$0.92{\pm}0.06^{\rm b}$	$0.73 \pm 0.06^{\circ}$	$0.76 \pm 0.06^{\circ}$	$1.10\pm0.06^{a}$	0.05	0.01	-	-
41-50 d	$1.14 \pm 0.07$	$1.08 \pm 0.07$	$0.94 \pm 0.07$	$1.07 \pm 0.07$	0.03	0.21	-	-
51-60 d	$1.28 \pm 0.09^{ab}$	$1.55 \pm 0.09^{\mathrm{a}}$	$1.04 \pm 0.09^{b}$	$1.48 \pm 0.09^{a}$	0.11	0.02		
Starter intake (kg/d)								
Whole period	0.63±0.03ª	$0.62^{a}\pm0.02$	0.61 <sup>a</sup> ±0.03	$0.54 \pm 0.03^{b}$	0.02	0.01	0.01	0.01
1-10 d	$0.13 \pm 0.01$	$0.12 \pm 0.01$	$0.12 \pm 0.01$	$0.09 \pm 0.01$	0.01	0.17	-	-
11-20 d	$0.29 \pm 0.02$	$0.27 \pm 0.02$	$0.30 \pm 0.02$	$0.28 \pm 0.02$	0.01	0.73	-	-
21-30 d	$0.45 \pm 0.03^{a}$	$0.44^{ab} \pm 0.03$	$0.37^{ab} \pm 0.03$	$0.36 \pm 0.03^{b}$	0.01	0.05	-	-
First month	$0.29 \pm 0.01$	$0.27 \pm 0.01$	$0.26 \pm 0.01$	$0.24{\pm}0.01$	0.01	0.08	0.01	0.16
31-41 d	$0.58 \pm 0.03$	$0.63 \pm 0.03$	$0.55 \pm 0.03$	$0.61 \pm 0.03$	0.01	0.34	-	-
41-50 d	$0.90 \pm 0.03$	$0.81 \pm 0.03$	$0.89 \pm 0.03$	$0.84 \pm 0.03$	0.01	0.12	-	-
51-60 d	$1.46 \pm 0.08^{a}$	$1.46^{a} \pm 0.08$	$1.43^{a} \pm 0.08$	$1.10 \pm 0.08^{b}$	0.04	0.01	-	-
Second month	$0.98 \pm 0.03^{a}$	$0.97^{a} \pm 0.03$	$0.96^{a} \pm 0.03$	$0.85{\pm}0.03^{\rm b}$	0.01	0.01	0.01	0.01
BW, kg								
Initial, 1 <sup>th</sup> day	$39.41 \pm 1.74$	$39.23 \pm 1.74$	$38.92 \pm 1.74$	$39.65 \pm 1.74$	0.84	0.99	-	-
Final, 56th day	$71.69{\pm}2.80^{ab}$	$73.78 \pm 2.80^{a}$	$69.23{\pm}2.80^{ab}$	$65.29{\pm}2.80^{\mathrm{b}}$	2.10	0.03	-	-
ADG (kg/d)	$0.70 \pm 0.04^{a}$	$0.72{\pm}0.04^{a}$	$0.62{\pm}0.04^{ab}$	$0.54{\pm}0.04^{\rm b}$	0.21	0.01	-	-
Feed Efficiency	$0.97{\pm}0.08$	$0.89 \pm 0.08$	$1.12 \pm 0.08$	$1.07{\pm}0.08$	0.03	0.14		

Treatments: WNA=Water not available, AW=Ad libitum water, WAM= Water available after milk feeding, WBM= Water available before milk feeding.

BW: Body weight, ADG: Average daily gain

<sup>a,b</sup> Values within a row with different superscripts differ significantly at P<0.05.

**Table 3.** Blood metabolite concentration for different water accessibility of Holstein calves with common starter and milk replacer(Mean  $\pm$  SEM)

Item	Treatments				SEM	Р		
	AW	WNA	WAM	WBM		Treat	Time	Treat×time
Glucose (mg/dL)	$128.6{\pm}6.98$	$139\pm 6.98$	$130.5{\pm}6.98$	$138 \pm 9.87$	3.36	0.68	0.42	0.72
Cholesterol (mg/dL)	$104.1 \pm 9.8$ <sup>a</sup>	$103.3 \pm 9.8^{a}$	$72.1\pm9.8^{b}$	$71.2 \pm 13.9^{b}$	5.87	0.03	0.09	0.67
Triglycerides (mg/dL)	30.8±3.05	45.8±3.98	53.6±34.10	44.2±3.45	4.03	0.18	0.08	0.62
AST (u/L)	37.5±4.37 <sup>b</sup>	$37.6 \pm 4.37^{b}$	$32.1 \pm 4.37^{b}$	$53.5{\pm}6.18^a$	2.14	0.01	0.01	0.01
ALT (u/L)	$7.6 \pm 0.84$	$10.1 \pm 0.84$	$8.1 \pm 0.84$	$8.0 \pm 1.18$	0.45	0.15	0.60	0.37
TAC (mmol/L)	$0.19 \pm 0.04^{b}$	$0.37{\pm}0.04^{a}$	$0.44 \pm 0.04^{a}$	$0.20{\pm}0.06^{\rm b}$	0.04	0.01	0.01	0.01
BHBA (mmol/L)	$0.10 \pm 0.02$	$0.10 \pm 0.02$	$0.13 \pm 0.02$	$0.08 \pm 0.02$	0.01	0.22	0.47	0.42
Albumin (g/dL)	$3.53 \pm 0.09$	$3.75 \pm 0.09$	$3.71 \pm 0.09$	$3.77 \pm 0.12$	0.04	0.31	0.67	0.96
NEFA (mmol/L)	$0.175 \pm 0.03$	$0.185 \pm 0.03$	$0.190 \pm 0.03$	$0.130 \pm 0.05$	0.01	0.58	0.92	0.82
Protein (g/dL)	$6.51 \pm 0.14^{b}$	$7.10 \pm 0.14^{a}$	$6.86 \pm 0.14^{ab}$	$6.57 \pm 0.19^{b}$	0.08	0.01	0.08	0.14
HDL (mg/dL)	$30.33 \pm 2.88$ <sup>ab</sup>	$36.16 \pm 2.88^{a}$	$23.16 \pm 2.88^{b}$	$23.50{\pm}4.07^{b}$	1.82	0.01	0.09	0.67
MDA (nmol/mL)	$0.91{\pm}0.89$	$1.05{\pm}~0.09$	$1.13 \pm 0.09$	$0.85{\pm}0.13$	0.05	0.21	0.08	0.72

Treatments: WNA=Water Not Available, AW=Adlibitum Water, WAM= Water available after milk feeding, WBM= Water available before milk feeding

<sup>a,b</sup> Values within a row with different superscripts differ significantly at P<0.05.

Table 3 summarizes the blood parameter data for each group and sampling date. Concentrations of glucose, cholesterol, TG, ALT, BHB, albumin, NEFA and MDA were similar among treatments (Table 3). But the serum concentrations of AST, TAC, protein and HDL significantly changed by different water accessibility. At 24–48 h after birth the level of total protein was in reference range but at other ages was below the reference value. In this study WBM has higher activity of serum AST (53.5 u/L) as compared with other treatments.

TAC levels of serum in this study were significantly higher in group WNA and WAM than others (P<0.05). The treat × time interaction was statistically significant for AST and TAC. Serum concentrations of Protein and HDL in WNA treatment is significantly higher than other groups (P<0.05). But were not influenced by either time or treat × time. Some blood electrolytes are presented in Table 4. The elements Cl and K did not significantly different among treatments. But Na concentration of blood serum in AW and WAM is 138.08 and 138.01, respectively. These groups had significantly higher serum concentration of Na than others (P<0.05). Water treatments did not affect the fecal score, temperature, and the respiratory score of calves, these parameters are presented in Table 5. The findings of the current study showed that different water accessibility of calves had no significant effects on health, respiratory and fecal score.

Mean body length, withers height, heart girth, body barrel, hip height and hip width during the pre-weaning periods are given in Table 6. There is no difference in frame growth and body condition scores of calves. Rumen fermentation data showed in Table 7. Acetic acid and

Table 4. Blood concentration of some electrolyte for different water accessibility of Holstein calves (Mean ± SEM)

Item		Treatments					Р	
	AW	WNA	WAM	WBM	_	Treat	Time	Treat× time
Cl (meq/L)	$100.5{\pm}9.33$	$93.83{\pm}9.33$	$108.33{\pm}9.33$	$104.75 \pm 13.20$	4.55	0.74	0.54	0.49
Na (meq/L)	$138.08{\pm}1.16^{a}$	$137.58{\pm}~1.16^{\mathrm{a}}$	$138.01{\pm}1.16^a$	$132.97 \pm 1.64^{b}$	0.75	0.01	0.03	0.23
K (meq/L)	$4.13{\pm}0.11$	$4.26{\pm0.11}$	$4.39{\pm}0.11$	$3.94{\pm}0.15$	0.06	0.13	0.12	0.62

Treatments: WNA=Water not available, AW=Adlibitum water, WAM= Water available after milk feeding, WBM= Water available before milk feeding

<sup>a,b</sup> Values within a row with different superscripts differ significantly at P<0.05.

Item		OEM	р			
	AW	WNA	WNA WAM		- SEM	Р
Health score <sup>1</sup>	$1.53 \pm 0.53$	$2.00 \pm 0.72$	$2.15 \pm 0.51$	$2.15 \pm 0.48$	0.23	0.78
Nasal discharge	$0.30 \pm 0.17$	$0.53 \pm 0.23$	$0.61 \pm 0.17$	$0.61 \pm 0.16$	0.07	0.43
Respiratory score	$0.61 \pm 0.17$	$0.76 \pm 0.23$	$1.23 \pm 0.16$	$1.15 \pm 0.16$	0.13	0.29
Temperature (°C)	39.07	39.16	39.29	39.31	0.05	0.38
Fecal score	$0.76 \pm 0.22$	$1.15 \pm 0.29$	$1.53 \pm 0.21$	$1.15 \pm 0.20$	0.13	0.24

Table 5. Fecal score, temperature, and respiratory score of Holstein calves (Mean  $\pm$  SEM)

Treatments: WNA=Water not available, AW=Ad libitum water, WAM= Water available after milk feeding, WBM= Water available before milk feeding

<sup>1</sup>The day is getting sick until recovery

Table 6. Frame growth measurements and body condition scores for different water accessibility of Holstein calves with common starter and milk replacer (Mean  $\pm$  SEM)

Item (cm)		Treatments					Р		
-	AW	WNA	WAM	WBM	SEM	Treat	Time	Treat×time	
Body length	$60.65{\pm}0.55$	$60.69{\pm}0.55$	$60.42{\pm}0.55$	$60.34 \pm 0.55$	0.58	0.96	0.01	0.21	
Withers height	82.03±0.82	83±0.85	81.42±0.79	82.38±0.81	0.50	0.44	0.01	0.88	
Heart girth	$82.92{\pm}0.80$	$83.88{\pm}0.80$	$81.65{\pm}0.80$	$81.46{\pm}0.80$	0.69	0.11	0.01	0.76	
Body barrel	$76.50{\pm}0.94$	$77.19{\pm}0.94$	$76.30{\pm}0.94$	$75.15{\pm}0.94$	0.67	0.49	0.01	0.92	
Hip height	$85.57{\pm}0.70$	$86.57{\pm}0.70$	$85.57{\pm}0.70$	$85.92{\pm}0.70$	0.51	0.71	0.01	0.77	
Hip width	$19.46 \pm 0.23$	$19.30 \pm 0.23$	$19.34{\pm}0.23$	$19.09 \pm 0.23$	0.16	0.71	0.01	0.81	

Treatments: WNA=Water not available, AW=Ad libitum water, WAM= Water available after milk feeding, WBM= Water available before milk feeding

Item		<b>CEM</b>	ъ			
	AW	WNA WAM		WBM	- SEM	Р
NH <sub>3</sub> (mmol/L)	23.05±1.25	23.42±1.35	28.77±1.48	31.27±1.65	1.98	0.41
Acetic acid (mM)	$58.40{\pm}5.62^{a}$	$57.42\pm5.62^{a}$	$36.19{\pm}5.62^{b}$	$23.83{\pm}5.62^{b}$	5.02	0.01
Propionic acid (mM)	$28.39{\pm}4.42^a$	$22.00{\pm}4.42^{ab}$	$12.37{\pm}4.42^{b}$	$8.06{\pm}4.42^{\rm b}$	3.05	0.04
Butyric acid (mM)	$7.04 \pm 2.11$	$6.59 \pm 2.11$	$8.35 \pm 2.11$	$2.40 \pm 2.11$	1.12	0.29
Isobutyrate (mM)	$0.64 \pm 0.53$	$2.01 \pm 0.53$	$1.96 \pm 0.53$	$0.92 \pm 0.53$	0.29	0.23
Valerate (mM)	$3.17{\pm}0.88$	$2.93{\pm}0.88$	$3.36 \pm 0.88$	$1.17 \pm 0.88$	0.45	0.33
Isovalerate (mM)	$1.76 \pm 0.46$	$1.91 \pm 0.46$	$1.77 \pm 0.46$	$0.88 \pm 0.46$	0.23	0.42

Table 7. Effects of water treatments on rumen fermentation parameters (Mean  $\pm$  SEM)

Treatments: WNA=Water not available, AW=Ad libitum Water, WAM= Water available after milk feeding, WBM= Water available before milk feeding, mM=milimolar

<sup>a,b</sup> Values within a row with different superscripts differ significantly at P<0.05.

Table 8. Total times devoted to perform different behaviors during 8 h of observation in preweaning periods (Mean ± SEM)

Item (h)	Treatments					Р		
	AW	WNA	WAM	WBM	SEM	Treat	Time	Treat×time
Visit water bucket	$0.64 \pm 0.15$ <sup>a</sup>	$0.01 \pm 0.14$	$0.40{\pm}0.15^{\text{ ab}}$	$0.43 \pm 0.14^{a}$	0.08	0.01	0.44	0.01
Visit starter bucket	$1.07 \pm 0.40$	$1.61{\pm}0.35$	$1.93{\pm}0.39$	$1.25{\pm}0.38$	0.19	0.39	0.22	0.07
Rumination	$1.07{\pm}0.45$	$1.11 \pm 0.40$	$1.73 \pm 0.43$	$1.00 \pm 0.42$	0.21	0.66	0.02	0.78
Standing	$1.21 \pm 0.41$	$1.61 \pm 0.36$	$1.06 \pm 0.40$	$1.62{\pm}0.39$	0.18	0.66	0.58	0.95
Lying in bed	$1.85 \pm 0.97$ <sup>b</sup>	$3.61{\pm}0.86^{ab}$	$2.80{\pm}0.94^{\text{ ab}}$	$5.00 \pm 0.91^{a}$	0.45	0.02	0.63	0.85
Object play	$1.92 \pm 0.49$	$1.77 \pm 0.43$	$1.66 \pm 0.47$	$1.50 \pm 0.46$	0.22	0.92	0.54	0.62
Self grooming	$1.71 \pm 0.42$	$1.05 \pm 0.37$	$2.26{\pm}0.40$	$1.50 \pm 0.39$	0.19	0.17	0.24	0.96
Nonnutritive sucking	$3.07{\pm}0.58$	$2.88{\pm}0.51$	$3.60 \pm 0.56$	$3.18 \pm 0.54$	0.26	0.83	0.45	0.66
Other	$1.00 \pm 0.36$	$1.11 \pm 0.32$	$1.60 \pm 0.35$	$1.00 \pm 0.34$	0.16	0.53	0.27	0.75

Treatments: WNA=Water not available, AW=Ad libitum water, WAM= Water available after milk feeding, WBM= Water available before milk feeding.

<sup>a,b</sup> Values within a row with different superscripts differ significantly at P<0.05.

propionic acid of AW group significantly increased as compared with other treatments (P<0.05). As shown in Table 8, treatments did not affect behaviors parameters. But the number of visiting water bucket of AW and WBM were higher than other groups.

#### **Discussion and Conclusion**

De Passillé et al. (8) reported that that water intake of calves is low until weaning (0.17 kg/d), whereas other calf studies measured considerable intakes of water (1 kg/d) (19). According to NRC (27), water intake will decrease with increasing cold until 4.4 °C and will remain constant for lower temperatures. In the different decade's age of calves' water intake, between groups had significantly changed, which may have been due to the accessibility of water in these groups during milk feeding calves. Kertz et al. (19) observed that daily water intakes decreased after 3 to 4 d of age compared with initial intakes. Increased DMI, and increased calf body size, which consequentially led to an increase in water requirements. The researchers indicated (3) demonstrated that total body water content of dairy calves decreased from 80 to 70% during the first 40 d. According to Wenge et al. (34) restrictively-milk fed calves drank 1.1 L water per day within the first 3 weeks of life. The NRC (27) reported that water intake increases from 1 kg/d at first week of age to 2.5 kg/d at 4 weeks of age. Lower water drinking of calves in this study may be because of decreasing the air temperature.

The studies were revealed that starter intake affected by water accessibility as shown by Gonzalez (11) that water appears to have a relationship with starter intake as starter intake is a significant variable in predicting the water intake of a calf, especially after day 21 when starter intake and water intake are both increasing. Prior to 21 days, the relationship between water intake and starter intake is not significant but does trend to be related. These results corroborate the positive relationship normally observed between total DM intake and water intake (19). Conversely, when a greater volume of milk is offered (and indirectly, a greater amount of TS), there is a negative relationship between DM intake and voluntary water intake (21), because the liquid feed fills part of the need for water (2). Unlike water in milk, which is shunted directly to the abomasum, drinking water enters and becomes a part of the developing reticulorumen (hereafter called rumen) of pre-weaned calves (13). Therefore, voluntary water intake of pre-weaned calves (0.75 to 0.82 kg/d) partially represents a water requirement of the developing rumen. In this study higher water intake of AW and WBM groups maybe showed that water in calves after 10 days did not supply the requirements. Thus, calves had the potential to consume more water in this system. These results agree with De Passillé et al. (8) and Hepola et al. (15), who reported higher water intakes in calves fed limited quantities of milk. In contrast, higher starter intakes tend to increase the quantity of water consumed (19).

The contradictory alteration by different water accessibility on blood metabolites was shown by different studies, as Beiranvand et al. (5) reported that addition of water to the starter (up to 50% DM) had no significant effects on blood glucose concentration. Serum albumin concentrations partially reflect hepatic synthesis and its increasing could be related to compensation of decreasing serum osmotic pressure due to globulin levels decline. The level of total protein was below reference value at ages except at 24-48 h after birth. It was approximately similar to Knowles et al. (22) reports. Serum total protein levels were below previously reported reference range from birth up to day 84 of age (9) and from birth up to day 90 of age (36). Knowles et al. (22) revealed that albumin levels in calves were below the reference range of adults until after nine days of age.

It is well known that AST is a nonspecific marker of acute liver damage caused by high-grain diets and associated ruminal lactate production (25). The activities of AST were below the reference range. This is consistent with the Egli and Blum (9) study. Zanker et al. (36) reported a postnatal rise of AST activity had no association with the time of first colostrum intake, indicating that the rise of AST activity was the consequence of endogenous production and was independent of colostrum intake. Serum constituents and hematological profiles evaluated were within normal ranges reported by Kaneko (18).

Reece (30) reported that serum concentration of Na, K and Cl in steers were 141.8, 4.6 and 105 (mEq/L), respectively. Roy et al. (31) indicated that Mean serumsodium values at birth were 139-140 mEq/L. They revealed that most of the deaths occurred, when the sodium level was low and the potassium level normal. With the increasing incidence of scouring, a greater fall in mean serum-sodium values occurred. Mean serumpotassium levels at birth were 5.8-6.0 mEq/L. For calves that scoured on the 1st day or not at all the mean serum potassium fell gradually to 5.8 mEq/L as the results of Roy et al. (31) study indicated clearly that, with an increased incidence of scouring the serum-sodium levels of calves decline with a concomitant but a slight rise in potassium values. NRC (27) showed that about eighteen times more water and eleven times more sodium and potassium were lost each day in the feces of scouring calves than in those of normal calves. They suggested that this loss was due to the increased osmotic pressure in the lumen of the gut owing to an increase in the number of smaller molecules, such as volatile fatty acids, resulting from bacterial activity. The higher osmotic pressure in the gut thus caused the infiltration of large amounts of water accompanied by electrolytes. It is well known that an increase in potassium concentration in the serum can occur with a cellular deficiency of potassium (30) and NRC (27) have emphasized that potassium should never be given to dehydrated patients because of the danger of an increase in potassium concentration in the serum and cardiac arrest. Wickramasinghe (35) reported that blood sodium concentrations of calves receiving drinking water from birth and calves that did not receive water during the first 17 d after birth were 134.4 and 136.1 mmol / L, respectively. They revealed that this could be due to a greater intake of electrolyte solution given the greater severity of scours among calves did not receive water until days 17. Roy et al. (31) studied changes in serum sodium and potassium concentrations in 150 experimental calves, some of which became affected with diarrhea. They observed that diarrhea caused a decrease in serum sodium and an increase in serum potassium concentrations. Milk is a poor source of the major electrolytes lost in diarrhea (Na<sup>+</sup>, Cl<sup>-</sup>, HCO3<sup>-</sup>) (27). Reece (30) reported that in 46 % of the plasma samples collected from diarrhoeic calves, the sodium concentrations were below normal. Since the diarrhoeic calves lost weight, and as part of this loss in body weight was presumably due to a reduction in body water (30), it would appear that these particular samples were from calves which were hypotonically dehydrated. However, as pointed out by Roy et al. (31), the plasma electrolyte concentration only shows the relative proportions of water to electrolytes in the plasma and in itself is not an indication of whether deficits or excesses of water and electrolytes are actually present in the body as a whole. Previous studies have confirmed (30) that in diarrheic calves there is characteristically a loss in body weight, negative sodium and potassium balance.

Overall, the calves in our study were healthy. This result agreed with Gottardo et al. (12) that reported calves receiving water showed a good health status throughout the fattening period, reducing the feed refusal days significantly. Water availability did not lead to the excretion of looser feces; therefore, no consequent detrimental effect on calves' body cleanliness was observed. Wickramasinghe et al. (35) reported that calves that 17 days delay of the offer first drinking water to newborn calves.

The excessive intake of drinking water might affect microbial fermentation in the rumen and thus rumen development. No study has examined the effect of water on rumen development of calves. Few studies focusing on the relationship between drinking water and rumen functions in mature cows provide some insight on the potential effects of drinking water in calves. For instance, Fraley et al. (10) found lower molar proportions of butyrate and propionate in the rumen, which were associated with lower liquid weight in the rumen and high fractional liquid passage rates in lactating cows with high water intake. Butyrate and propionate are often considered to be major drivers of morphological and metabolic developments of the rumen in calves (23). Therefore, the increased water intake of WBM calves could be hypothesized to inversely affect rumen development.

The number of drinking bouts recorded by Beiranvand et al. (5) resulted in mean values between 3 and 5 visits per animal per day and this was at a similar level compared to our own results. According to Gottardo et al. (12), the results of the two indicators illustrated that suckler cows showed climate-induced behavioral changes, whereas water intake was not influenced by increasing cold.

The results indicate that suckler calves with different water accessibility had different water intake. AW and WBM treatments drank more water than other groups (P<0.05). Water accessibility was also found to increase AST and Na concentration of serum but serum concentrations of protein and HDL and TAC levels were significantly higher in groups WNA and WAM than others (P<0.05). Different water accessibility of calves had no significant effects on respiratory and fecal score, growth parameters and behaviors. But acetic acid and propionic acid of AW groups significantly increased as compared with other treatments (P<0.05). Overall, in agreement with our hypotheses, giving water to young calves around milk meal had some beneficial effects but resulted in similar growth performance compared with calves that did not access to water. As a conclusion, it can be recommended that water accessibility around milk meal for suckling calves will be resulted in healthier calves.

#### Acknowledgments

Authors are grateful to the Isfahan University of Technology and the University of Mohaghegh Ardabili for their help.

#### **Financial Support**

This research received no grant from any funding agency/sector.

#### **Conflict of Interest**

The authors declare that they have no competing interests.

#### References

- AOAC (2000): Method 973.18. Official methods of analysis. Association of Official Analytical Chemists, International, Gaithersburg, MD, USA.
- 2. Azevedo A, Machado FS, Campos MM, et al (2016): The effects of increasing amounts of milk replacer powder added to whole milk on feed intake and performance in dairy heifers. J Dairy Sci, 99, 8018-8027.
- 3. Bar-Peled U, Robinzon B, Maltz E, et al (1997): Increased weight gain and effects on production parameters of Holstein heifer calves that were allowed to suckle from birth to six weeks of age. J Dairy Sci, **80**, 2523–2528.
- Beede DK (2005): *The most essential nutrient: Water*. In: Proc. 7th Western Dairy Management Conf. Reno, NV, March 9-11, pp. 13-31.
- 5. Beiranvand H, Khani M, Omidian S, et al (2016): Does adding water to dry calf starter improve performance during summer? J Dairy Sci, 99, 1903-1911.
- Chapman CE, Stone Wilkinson P, Murphy MR, et al (2017): Evaluating nuclear magnetic resonance spectroscopy for determining body composition in Holstein dairy calves using deuterium oxide dilution methods. J Dairy Sci, 100, 2807–2811.
- 7. **Church DC** (1991): Livestock feeds and feeding. 3rd ed. Prentice Hall, Englewood Cliffs, NJ.
- De Passillé AM, Borderas TF, Rushen J (2011): Weaning age of calves fed a high milk allowance by automated feeders: Effects on feed, water, and energy intake, behavioral signs of hunger, and weight gains. J Dairy Sci, 94, 1401-1408.
- Egli CP, Blum JW (1998): Clinical, hematological, metabolic and endocrine traits during the first three months of life of suckling simmentaler calves held in a cow-calf operation. J Vet Med A, 45, 99-118.
- Fraley SE, Hall MB, Nennich TD (2015): Effect of variable water intake as mediated by dietary potassium carbonate supplementation on rumen dynamics in lactating dairy cows. J Dairy Sci, 98, 3247–3256.
- Gonzalez F (2009): Association among fluid, grain intake and weight gain in Holstein bull calve. M.sc. Thesis of Texas A & M University.
- 12. Gottardo F, Mattiello S, Cozzi G, et al (2002): The provision of drinking water to veal calves for welfare purposes. J Anim Sci, 80, 2362–2372.
- Govil K, Yadav DS, Patil AK, et al (2017): Feeding management for early rumen development in calves. J Entomol. Zool Stud, 5, 1132–1139.
- 14. Heinrichs AJ, Jones CM, VanRoekel LR, et al (2003): A system of dairy calf workforce management, training, and evaluation and health evaluation. J Dairy Sci, 86, 115-120.
- 15. Hepola HP, Hänninen LT, Raussi SM, et al (2008): Effects of providing water from a bucket or a nipple on the performance and behavior of calves fed ad libitum volumes of acidified milk replacer. J Dairy Sci, **91**, 1486–1496.
184 Mohsen Zarei - Jamal Seifdavati - Gholam Reza Ghorbani - Hosein Abdi-Benemar - Reza Seyedsharifi - Abdolhamid Karimi

- 16. **Iranian Council of Animal Care** (1995): *Guide to the care and use of experimental animals*, vol. 1. Isfahan University of Technology, Isfahan, Iran.
- 17. Jasper J, Weary DM (2002): Effects of ad libitum milk intake on dairy calves. J Dairy Sci, 85, 3054–3058.
- Kaneko JJ (1997): Serum proteins and the disproteinemias. In: Clinical biochemistry of domestic animals, Kaneko JJ, Harvey J.W and Bruss ML, (Ed.), pp. 117-138, Academic Press, San Diego, California,.
- Kertz AF, Reutzel LF, Mahoney JH (1984): Ad libitum water intake by neonatal calves and its relationship to calf starter intake, weight gain, feces score and season. J Dairy Sci, 67, 2964–2969.
- 20. Khan MA, Bach A, Castells L, et al (2014): Effects of particle size and moisture levels in mixed rations on the feeding behaviour of dairy heifers. Animal, 8, 1722–1727.
- 21. Kiezebrink DJ, Edwards AM, Wright TC, et al (2015): Effect of enhanced whole-milk feeding in calves on subsequent first-lactation performance. J Dairy Sci, 98, 349-356.
- 22. Knowles TG, Edwards JE, Bazeley KJ, et al (2000): Changes in the blood biochemical and haematological profile of neonatal calves with age. Vet Rec, 147, 593-598.
- 23. Lane MA, Baldwin VIRL, Jesse BW (2000): Sheep rumen metabolic development in response to different dietary treatments. J Anim Sci, 78, 1990–1996.
- Lesmeister KE, Heinrichs AJ (2004): Effects of corn processing on growth characteristics, rumen development and rumen parameters in neonatal dairy calves. J Dairy Sci, 87, 3439–3450.
- 25. Mori A, Urabe S, Asada M, et al (2007): Comparison of plasma metabolite concentrations and enzyme activities in beef cattle raised by different feeding systems in Korea, Japan and New Zealand. J Vet Med A Physiol Pathol Clin Med, 54, 342–345.
- Murphy MR (1992): Water metabolism of dairy cattle. J Dairy Sci, 75, 326–333.
- 27. **NRC** (2001): Nutrient requirements of dairy cattle. 7th. rev. ed. National Acad. Sci. Washington, DC.

- 28. Pempek JA, Eastridge ML, Swartzwelder SS, et al (2016): Housing system may affect behavior and growth performance of Jersey heifer calves. J Dairy Sci, 99, 569– 578.
- Quigley J (2000): Predicting water intake in young calves. Available at: http://www.calfnotes.com. (Accessed Nov 11, 2000).
- 30. **Reece WO** (1984): Acid-base balance and selected hematologic, electrolytic, and blood chemical variables in calves nursing cows: one week through fifteen weeks. Am J Vet Res **45**, 666-669.
- 31. Roy RN, Chance GW, Radde IC, et al (1976): Late hyponatraemia in very low birth weight infants less than 1.3 kg. Pediatr Res, 10, 526-531.
- 32. **SAS Institute**, 'SAS/STATuser's guide: Statistics. Version 9.1, (SAS Institute Inc. Cary, NC), 2001.
- TierSchNutzt.V (Tierschutz-Nutztierhaltungsverordnung) (2006): Regulation on the protection of livestock and other animals kept for the production of animal products in their attitude. 11. Monitoring, caring and feeding of calves. Bundesjustizministerium (Federal Ministry of Justice), Berlin, Germany.
- Wenge J, Steinhöfel I, Heinrich C, et al (2014): Water and concentrate intake, weight gain and duration of diarrhea in young suckling calves on different diets. Livest Sci, 159, 133–140.
- Wickramasinghe HKJP, Kramer AJ, Appuhamy JADRN (2019): Drinking water intake of newborn dairy calves and its effects on feed intake, growth performance, health status, and nutrient digestibility. J Dairy Sci, 102, 377-387.
- Zanker I A, Hammon HM, Blum JW (2001): Activities of γ-Glutamyltransferase, alkaline phosphatase and aspartateaminotransferase in colostrum, milk and blood plasma of calves fed first colostrum at 0–2, 6–7, 12–13 and 24–25 h after Birth. J Vet Med A Physiol Pathol Clin Med, 48, 179-185.

Ankara Üniv Vet Fak Derg, **67**, 185-192, 2020 DOI: 10.33988/auvfd.623821

# The effects of fumaric and malic acids on the *in vitro* true digestibility of some alternative feedstuffs for ruminants

# Buğra GENÇ<sup>1,a,∞</sup>, Mustafa SALMAN<sup>2,b</sup>, Bora BÖLÜKBAŞ<sup>2,c</sup>, İsmail KAYA<sup>2,d</sup>, Mustafa AÇICI<sup>3,e</sup>

<sup>1</sup>Ondokuz Mayıs University, Faculty of Veterinary Medicine, Department of Laboratory Animals, Samsun; <sup>2</sup>Ondokuz Mayıs University, Faculty of Veterinary Medicine, Department of Animal Nutrition and Nutritional Diseases, Samsun; <sup>3</sup>Ondokuz Mayıs University, Faculty of Veterinary Medicine, Department of Parasitology, Samsun, Turkey.

<sup>a</sup>ORCID:0000-0002-7561-4993; <sup>b</sup>ORCID:0000-0003-0828-5998; <sup>c</sup>ORCID:0000-0002-0732-0192; <sup>d</sup>ORCID:0000-0002-2570-0877; <sup>c</sup>ORCID:0000-0002-8406-9739

> <sup>⊠</sup>Corresponding author: bugragenc@omu.edu.tr Received date: 24.09.2019- Accepted date: 24.12.2019

**Abstract:** The aim of this study was to determine the effects of the addition of different amounts of fumaric acid (FA) and malic acid (MA) to the leaves of *Robinia pseudoacacia* (Black locust, acacia), *Prunus laurocerasus* (cherry laurel), *Quercus cerris* (oak), and *Camellia sinensis* (tea factory wastes, TFW), to improve their value as alternative feeds for ruminants. The parameters examined were the *in vitro* true digestibility of feed (IVTD<sub>As fed</sub>), dry matter (IVTD<sub>DM</sub>), organic matter (IVTD<sub>OM</sub>), neutral detergent fiber (IVTD<sub>NDF</sub>) and count of protozoans. The digestibility experiments were performed with a DAISY incubator system. Organic acids were not added in the control group and 0.1%, 0.2% or 0.3% FA or MA were added to the experimental groups. Each treatment was replicated 6 times. Samples were incubated for 48 hours. Fumaric acid significantly reduced (P<0.01) all digestibility values of *R. pseudoacacia* leaves. When FA was applied at 0.1% to *C. sinensis* factory wastes, the IVTD<sub>OM</sub> increased significantly (P<0.05), with the same effect observed for *Q. cerris* (P<0.01). However, for the addition of 0.1% FA, IVTD<sub>As Fed</sub>, IVTD<sub>DM</sub> and IVTD<sub>NDF</sub> values decreased significantly (P<0.01). Separately, malic acid did not have a significant effect on the *in vitro* true digestibility values determined in this study (P>0.05). Rumen protozoan counts decreased for both organic acid applications compared to counts in the fresh rumen contents. Because 0.1% fumaric acid increased the IVTD<sub>OM</sub> values of both *C. sinensis* factory wastes and *Q. cerris* leaves they can be considered potential alternative feed sources for ruminants.

Keywords: Fumaric acid, in vitro, malic acid, true digestibility

# Ruminantlar için fumarik ve malik asitlerin bazı alternatif yem maddelerinin *in vitro* gerçek sindirilebilirliği üzerine etkileri

Özet: Bu araştırmanın amacı farklı düzeylerdeki fumaric (FA) ve malik (FA) asitin ruminant alternatif yem kaynağı olarak *Robinia pseudoacacia* (yalancı akasya), *Prunus laurocerasus* (karayemiş), *Quercus cerris* (meşe), ve *Camellia sinensis* (fabrika atığı, TFW) yapraklarının sindirilebilirlik değerleri üzerine olan etkilerini incelemektir. Araştırmada materyallerin yem bazında (IVTD<sub>As fed</sub>), kuru madde bazında (IVTD<sub>DM</sub>), organic maddede (IVTD<sub>OM</sub>), nötral deterjen fiber (IVTD<sub>NDF</sub>) *in vitro* gerçek sindirilebilirlikleri ve protozoa sayıları parametreleri üzerine çalışılmıştır. Sindirilebilirlik parametreleri DAISY inkübatör sistemi ile gerçekleştirilmiştir. Kontrol grubuna FA ve MA eklenmezken deneme grupları ayrı ayrı ve sırasıyla %0,1, %0,2, %0,3 FA ve MA içerecek şekilde düzenlenmiştir. Her örnek 6 kez tekrarlı olacak şekilde araştırmaya dahil edilmiştir. Örnekler 48 saat inkubasyona tabi tutulmuştur. Fumarik asit *R. pseudoacacia* yapraklarında tüm sindirilebilirlik değerlerini önemli (P<0,01) derecede azaltırken değerlerin doza bağlı linear değişimi de önemli (P<0,01) bulunmuştur. Fumarik asit TFW materyaline %0,1 düzeyinde uygulandığında IVTD<sub>OM</sub> önemli derecede (P<0.05) yükselmiştir. Aynı etki *Q. cerris* için de görülmüş (P<0,01) ancak IVTD<sub>As Fed</sub>, IVTD<sub>DM</sub> ve IVTD<sub>NDF</sub> değerlerinin önemli seviyede (P<0,01) düştüğü saptanmıştır. Malik asit bu araştırmada *in vitro* gerçek sindirim değerleri üzerine önemli bir etki yaratmamıştır (P>0,05). Rumen protozoa sayıları her iki organik asit uygulamasında da taze rumen içeriğine göre azalmıştır. Sonuç olarak, *C. sinensis* fabrika atıkları ve *Q. cerris* yapraklarına, %0,1 düzeyinde fumarik asit ilavesinin IVTD<sub>OM</sub> değerlerini artırdığından dolayı, *C. sinensis* fabrika atıkları ve *Q. cerris* yapraklarının ruminantlar için potansiyel alternatif yem kaynağı olarak değerlendirilebileceği kanısına varılmıştır.

Anahtar sözcükler: Fumarik asit, gerçek sindirilebilirilik, in vitro, malik asit.

# Introduction

Organic acids have been used for many years for the purpose of improving the effectiveness of beneficial microorganisms and hence of rumen fermentation in ruminants. Organic acids are described as "Generally Recognized As Safe" (32) for addition to animal feed. The rumen has both a complex biota and ecology. Different species of eukaryotes, prokaryotes, archaeans and bacteriophages play important roles in the functioning of the rumen (20). Dicarboxylic organic acids such as fumaric acid (FA) and malic acid (MA) have been used to manipulate the microbial ecology of the rumen (10, 19). Organic acids can pass through the intestinal mucosal barrier by diffusion and are then involved in the Krebs cycle (32). The use of FA and MA increased the growth rate and number of the bacterium Selenomonas ruminantium (25, 32) which uses lactate produced by ruminal bacteria as a source of energy (25). These acids have beneficial effects on fermentation and microbial populations when used in combination with cellobiose and monensin. Organic acids have also been shown to increase the rate of protein hydrolysis (32).

The plant materials used in this research can be found in many geographies and are easy to obtain and naturally contain tannins. The search for alternative feed sources for ruminants has increased the interest in leaves containing tannins as potential resources. Tannins are water-soluble, polyphenolic compounds usually found in plants with high fibre levels. Although there can be adverse effects from tannins, improved feed consumption, feed efficiency and growth have been reported (35). Furthermore, tannins have been included in animal feed for research purposes due to their antioxidant effects against free radicals, metal binding properties and lipid peroxidation inhibition properties (18). However, Tieman et al. (36) reported that plants with high tannin content have low cellulose digestibility. In spite of some anti-nutritional properties, plant sources containing tannins have been used in the feeding of different kinds of animals because they are easy to access and cheap (35).

Against that background, the aim of this study was to use *in vitro* methods to determine the effects of the addition FA and MA on the actual digestibility parameters and count of protozoans associated with four alternative roughage sources.

### **Materials and Methods**

For this study, ethics committee approval was not needed because the rumen samples were collected only from slaughtered animals.

The tree leaves (*Robinia pseudoacacia* (black locust, acacia), *Prunus laurocerasus* (cherry laurel), *Quercus cerris* (oak)), used in the study were collected according to the methodology described in British Columbia

Ministry of Forests (5) from Samsun Canik Town forest (41°15' 29" N, 36°21'56" E and 41° 15' 48" N, 36° 22' 05" E), which is located approximately 150 m above sea level. The tea (*Camellia sinensis*) factory waste was obtained from tea factories located in Rize Province at the eastern end of the Black Sea region in Turkey. All tree leaves and tea factory waste samples were collected in May 2019. The FA ( $\geq$ 99% purity) and MA ( $\geq$ 95% purity) were obtained from Sigma Aldrich<sup>®</sup> (Istanbul, Turkey).

Fresh tree leave samples were weighed as fresh and then dried at 65 °C for 48h. The dried samples were ground in a mill and then passed through a sieve with a mesh diameter of 1 mm for chemical analysis. Dry matter (DM) content of each sample was determined in air circulation drying oven at 105 °C for 4 hours and the ash content was determined by burning the dried material in an ash oven at 550 °C for 4 hours. The Kjeldahl method was used to determine the crude protein (CP) percentage. Ether extract (EE) was performed according to the methods of AOAC (2). The neutral detergent fiber (NDF) and acid detergent fiber (ADF) contents of the materials that form the cell wall components of the feeds used in this study were determined in accordance with the method reported by Van Soest et al. (37) and were analyzed in a ANKOM 200 Fiber Analyzer (ANKOM Technology Corp. Fairport, NY, USA). The in vitro true digestibility (IVTD) analysis was performed with the ANKOM Daisy Incubator (ANKOM Technology Corporation), according to the methodology described in ANKOM (1). The rumen fluid was collected post-mortem from the rumens of four Holstein x Yerlikara hybrid cattle aged three years that were slaughtered in a commercial abattoir in Samsun, Turkey. Animals were fed twice daily with a diet containing grass hay and maize silage (60%) and concentrates (40%). A thermos was used to store and transport the rumen fluid, which was preheated to 39°C and had CO<sub>2</sub> added. The rumen fluid was collected manually by squeezing two handfulls of ruminal contents from each animal's rumen into the same thermos. In the laboratory, the rumen fluid was filtered through 4 layers of gauze. The F57 bags to be used in the analysis were rinsed with acetone (99.5%) for 3 minutes and then the acetone was evaporated at room temperature. All the bags were marked with both acid and alkaline resistant pen. The bags were dried in a drying cabinet at 105 °C for 2 hours. The tare weights of the bags were recorded. Samples of plants weighing 0.5 g were transferred to separate F57 bags which were then closed. The buffer solution to be used in the analysis was prepared according to the Ankom Daisy in vitro fermentation system described in ANKOM (1). Four digestion units, each with a volume of 2 L, were used in this test. The buffer solution was heated to 39 °C and 1.6 L was poured into each digestion unit. Four hundred mL of rumen fluid was added to each unit. A total

of 24 feed samples were used in each digestion unit, and 6 replicates were formed from each feed sample at the same time. The tests with FA and MA were carried out separately but with otherwise identical procedures. While no acid was added to the control group units, 0.1, 0.2 or 0.3% FA or MA was added to the experimental group units. A total of 24 feed samples were added to each of the control and experimental group units, that is, F57 bags, with 6 replicates of each feed sample. The samples were incubated for 48 hours. After the incubation period, all the liquid in the digestion units was removed and the bags were washed under running water. NDF analysis was performed as per the method outlined in ANKOM (1) by placing the bags in the Ankom Fiber Analyzer device. After analysis, the bags were retained in the drying cabinet until they reached a constant weight at 105 °C. The IVTD values of all samples were calculated with the formula reported in ANKOM (1),

 $IVTD\% = \frac{100 - (W3 - (W1xC1))x100}{W2}$ where: W1= Bag tare weight W2 = Sample weight

W3= Final bag weight after *in vitro* process and sequential ND treatment

C1= Blank bag correction (final oven-dried weight/original blank bag weight)

For the protozoan count, a mixture of 0.6 g methyl green, 8 g sodium chloride (NaCl) and 100 ml 37% formaldehyde was prepared (31). The mixture volume was made up to 1,000 ml with distilled water. One milliliter of the mixture and 1 mL of the liquid containing protozoans that had been taken from the digestion unit were transferred to a Fuchs Rosenthal counting chamber apparatus. For the protozoa count, fresh rumen fluid and samples taken from each digestion unit after the incubation period were studied in parallel. The protozoans were counted on an object slide under a light microscope (Nicone eclipse 80i) with a Fuchs-Rosenthal counting chamber (depth: 0.2 mm, small square area: 0.0625 mm<sup>2</sup>) (31).

Cell number per CMM

= Number of cells counted  $X \frac{1}{Area counted (mm^2)} X \frac{1}{Depth (mm)} X Dilution$ 

*Statistical analysis:* The Kolmogorov-Smirnov Test was used to check for normal distribution of the data, and for homogeneity of variance, the data were evaluated with the Levene Test. All traits on digestibility in the study were summarized as the mean of the group and standard error of means (SEM). For the determination of the differences among the groups, the one-way ANOVA model was fitted to the data for chemical composition, IVTD<sub>As Fed</sub>, IVTD<sub>DM</sub>, IVTD<sub>OM</sub> and IVTD<sub>NDF</sub>. ANOVA equation is:

 $Y_{ij} = \mu + a_i + e_{ij} \ (1),$ 

Where  $Y_{ij}$  is the value for i. group and j. observation;  $\mu$  is the population mean; and  $e_{ij}$  are the individual error terms distributed as N~(0, 1).

To evaluate the differences among the three concentrations of the organic acids, second degree orthogonal polynomial contrasting was used (13). One-way ANOVA and the other statistical tests and calculations were executed with SPSS Software (34).

#### Results

Chemical composition of the four alternative feeds are stated in Table 1. The IVTD<sub>As Fed</sub>, IVTD<sub>DM</sub>, IVTD<sub>OM</sub>, and IVTD<sub>NDF</sub> values for all doses of FA applied to *R. pseudoacacia* leaves were significantly different from the control (P<0.01) (Table 2). Fumaric acid negatively affected *in vitro* digestion across all parameters in *R. pseudoacacia* leaves. In contrast, MA did not significantly affect *in vitro* digestion in any of the examined digestion parameters (P>0.05) in *R. pseudoacacia* leaves. However, when MA was administered at 0.1%, the values of all parameters for *in vitro* digestion were numerically higher in *R. pseudoacacia* leaves.

Fumaric acid was found to be significantly (P<0.05) effective only on IVTD<sub>OM</sub> and the highest increase was seen in 0.1% dosing in TFW (Table 3). On the other hand, *in vitro* digestion was found to be numerically higher to all parameters when administered at doses of 0.1% and 0.2% in TFW. Malic acid did not significantly (P>0.05) affect *in vitro* digestion of TFW. However, it was observed that *in vitro* digestion values increased numerically to all parameters although it was not statistically (P>0.05) significant depending on the doses in TFW.

Table 1. Nutrient composition (g/100g DM) of alternative feed sources for ruminants.

Feed sources	DM	Ash	СР	EE	ADF	NDF
C. sinensis	93.5	4.8	18.2	1.16	34.6	40.5
Q. cerris	95.7	4.3	10.1	3.2	32.2	40.1
R. pseudoacacia	92.3	5.5	27.5	2.4	15.8	18.2
P. laurocerasus	90.7	10.8	8.5	1.1	11.3	23.1

DM: Dry matter, CP: Crude protein, EE: Ether extract, ADF: Acid detergent fiber, NDF: Neutral detergent fiber.

Fumaric acid	IVTDAs Fed	<b>IVTD</b> <sub>DM</sub>	<b>IVTD</b> <sub>OM</sub>	<b>IVTD</b> <sub>NDF</sub>
Control 0%	60.98±0.01	60.66±0.01	60.81±0.03	17.46±0.02
0.1%	60.24±0.31	59.85±0.34	60.02±0.34	16.22±0.53
0.2%	58.13±0.27	57.58±0.29	57.73±0.20	12.72±0.45
0.3%	57.94±0.02	57.37±0.02	57.55±0.01	12.41±0.03
Р				
Combined	0.001	0.001	0.001	0.001
Linear	0.002	0.002	0.001	0.002
Malic acid	IVTD <sub>As Fed</sub>	<b>IVTD</b> <sub>DM</sub>	IVTD <sub>OM</sub>	<b>IVTD</b> <sub>NDF</sub>
Control 0%	59.75±0.62	59.32±0.68	59.43±0.69	15.40±1.05
0.1%	60.80±0.16	60.46±0.17	60.52±0.15	17.16±0.26
0.2%	60.86±0.39	60.53±0.42	60.69±0.44	17.27±0.66
0.3%	57.93±0.96	57.35±1.04	57.39±0.91	12.38±1.60
Р				
Combined	0.077	0.077	0.058	0.077
Linear	0.943	0.943	0.861	0.943

**Table 2.** Effects of the addition of different concentrations (%) of organic acids to *R. pseudoacacia* leaves (n=6) on *in vitro* true digestibility values (Mean $\pm$ SEM).

IVTDAs Fed: *In vitro* true digestibility as fed, IVTD<sub>DM</sub>: *In vitro* true digestibility of dry matter, IVTD<sub>OM</sub>: *In vitro* true digestibility of organic matter, IVTD<sub>NDF</sub> : *In vitro* true digestibility of neutral detergent fiber.

Table 3. Effects of addition of different concentrations (%) of organic acids (fumaric acid, malic acid) to <i>C. sinensis</i> (n=6) factory
waste product (TFW) on <i>in vitro</i> true digestibility values (Mean±SEM).

Fumaric acid	IVTDAs Fed	IVTDDM	<b>IVTD</b> OM	<b>IVTD</b> <sub>NDF</sub>
Control 0%	50.96±0.90	49.21±0.96	49.73±0.96	12.19±1.27
0.1%	53.21±0.10	51.63±0.11	52.13±0.10	15.36±0.14
0.2%	51.26±0.35	49.54±0.38	50.12±0.37	12.67±0.51
0.3%	50.06±0.36	48.25±0.38	48.73±0.36	10.94±0.53
Р				
Combined	0.051	0.051	0.048	0.054
Linear	0.057	0.057	0.061	0.062
Malic acid	IVTD <sub>As Fed</sub>	IVTD <sub>DM</sub>	IVTD <sub>OM</sub>	<b>IVTD</b> <sub>NDF</sub>
Control 0%	50.20±0.23	48.41±0.25	48.89±0.24	11.17±0.35
0.1%	51.38±0.87	49.67±0.93	50.26±0.94	12.79±1.24
0.2%	51.97±0.75	50.30±0.81	50.91±0.80	13.65±1.03
0.3%	52.98±0.37	51.38±0.40	51.97±0.42	15.06±0.56
Р				
Combined	0.128	0.128	0.117	0.131
Linear	0.536	0.536	0.525	0.527

IVTDAs Fed: *In vitro* true digestibility as fed, IVTD<sub>DM</sub>: *In vitro* true digestibility of dry matter, IVTD<sub>OM</sub>: *In vitro* true digestibility of organic matter, IVTD<sub>NDF</sub> : *In vitro* true digestibility of neutral detergent fiber.

Fumaric acid produced only a numerical increase (P> 0.05) in the *in vitro* digestion parameters of the leaves of *P. laurocerasus* (Table 4) when applied at a dose of 0.3%, however MA had no significant (P>0.05) effect on any of the parameters.

It was found that *in vitro* digestion levels were significantly (P<0.005) reduced for  $IVTD_{As Fed}$ ,  $IVTD_{DM}$  and  $IVTD_{NDF}$  parameters by adding FA to *Q.cerris* (Table 5) leaves however,  $IVTD_{OM}$  digestion was found to be

significantly (P<0.005) higher when 0.1% dose was applied. At the same time dose-dependent changes of the differences were also significant (P<0.05). It was found that MA had an enhancing effect on the *in vitro* digestion of *Q. cerris* leaves at a dose of 0.1% for all parameters, but the increase was not statistically significant (P>0.05).

The effects of different concentrations of organic acids on the total count of rumen protozoans are stated in Table 6. Compared to the fresh rumen content group, a decrease was observed in the experimental groups. Fumaric acid at all level of 0.1% and 0.3% increased the count of protozoans numerically in comparison to the control group and in contrast MA numerically reduced the count of protozoa inversely proportional to increasing dose.

**Table 4.** Effects of addition of different concentration (%) of organic acids (fumaric acid, malic acid) to *P. laurocerasus* (n=6) leaves on *in vitro* true digestibility values (Mean±SEM).

Fumaric acid	IVTDAs Fed	IVTDDM	<b>IVTD</b> <sub>OM</sub>	<b>IVTD</b> <sub>NDF</sub>
Control 0%	59.76±0.58	58.90±0.64	58.99±0.63	14.33±0.94
0.1%	59.49±0.71	58.60±0.79	58.98±0.86	13.89±1.16
0.2%	59.04±1.49	58.11±1.64	58.45±1.61	13.16±2.41
0.3%	60.51±0.11	59.73±0.12	59.96±0.11	15.55±0.18
Р				
Combined	0.706	0.706	0.747	0.706
Linear	0.736	0.736	0.719	0.736
Malic acid	IVTDAs Fed	IVTD <sub>DM</sub>	IVTD <sub>OM</sub> IVTD <sub>N</sub>	DF
Control 0%	61.17±0.56	60.46±0.61	60.63±0.61	16.62±0.90
0.1%	61.73±0.14	61.07±0.15	61.23±0.17	17.51±0.22
0.2%	61.58±0.01	60.91±0.02	61.04±0.02	17.28±0.03
0.3%	61.17±0.42	59.73±0.12	60.63±0.47	16.61±0.69
Р				
Combined	0.634	0.634	0.659	0.634
Linear	0.787	0.787	0.750	0.787

IVTDAs Fed: *In vitro* true digestibility as fed, IVTD<sub>DM</sub>: *In vitro* true digestibility of dry matter, IVTD<sub>OM</sub>: *In vitro* true digestibility of organic matter, IVTD<sub>NDF</sub> : *In vitro* true digestibility of neutral detergent fiber.

Table 5. Effects of addition of different levels (%) of organic acids (fumaric acid, malic acid) to Q. cerris (n=6) leaves on in vitro true
digestibility values (Mean±SEM).

Fumaric acid	IVTDAs Fed	<b>IVTD</b> <sub>DM</sub>	<b>IVTD</b> <sub>OM</sub>	<b>IVTD</b> <sub>NDF</sub>
Control 0%	46.75±0.14	45.47±0.15	45.40±0.89	12.86±0.14
0.1%	46.64±0.11	45.35±0.12	46.01±0.12	11.88±0.55
0.2%	45.82±0.25	44.49±0.26	44.94±0.37	9.64±0.62
0.3%	45.69±0.29	44.36±0.30	44.81±0.15	9.17±0.07
Р				
Combined	0.004	0.004	0.004	0.005
Linear	0.020	0.020	0.017	0.020
Malic acid	IVTD <sub>As Fed</sub>	<b>IVTD</b> <sub>DM</sub>	<b>IVTD</b> <sub>OM</sub>	<b>IVTD</b> <sub>NDF</sub>
Control 0%	46.95±0.09	45.68±0.10	46.36±0.09	11.48±0.12
0.1%	47.52±0.43	46.28±0.46	46.94±0.46	12.33±0.58
0.2%	46.76±0.62	45.47±0.66	46.08±0.56	11.21±0.98
0.3%	46.73±0.75	45.44±0.79	46.13±0.80	11.16±1.15
Р				
Combined	0.499	0.499	0.464	0.520
Linear	0.227	0.227	0.190	0.240

IVTDAs Fed: *In vitro* true digestibility as fed, IVTD<sub>DM</sub>: *In vitro* true digestibility of dry matter, IVTD<sub>OM</sub>: *In vitro* true digestibility of organic matter, IVTD<sub>NDF</sub> : *In vitro* true digestibility of neutral detergent fiber.

Table 6. Effects of addition of different concentrations (%) of fumaric acid (FA) and malic acid (MA) on the total ruminal protozoa count per mL.

Additives	Protozoa count in fresh rumen liquid		Protozoa count after 48 hours incubation				
		Control 0%	0.1%	0.2%	0.3%		
FA	917262	161938	186731	143438	190625		
MA	950000	198929	182929	165954	161928		

# **Discussion and Conclusion**

Chemical composition (DM, Ash, CP, EE, ADF and NDF) of feedstuffs is known to be important in terms of animal nutrition. For *P. laurocerasus*, the parameters mentioned above have not been investigated previously. Özyılmaz (28) reported that TFW had 93.42% DM, 14.07% CP, 4.69% ash, 1.07% EE, 47.76% NDF and 40.93% ADF levels and that these values varied according to the organic or conventional cultivation of the tea plants and their harvest periods. Parissi et al. (29) and Luginbuhl and Mueller (24) reported that *R. pseudoacacia* leaves had 27.3% CP (g/100g DM) and 28.0% (g/100g DM) respectively, which is consistent with the results of the present study.

For *Q. cerris*, Kaya and Kamalak (15) reported 91.6% DM, 4.3% CP, 23.6% NDF and 18% ADF. Also for *Q. cerris* leaves, Canbolat et al. (8) reported 94.6% DM, 8.4% CP, 5.5% ash, 43.5% NDF and 36% ADF levels. While some of the values obtained in our study are in relative agreement with the results presented for other studies, other results differed markedly. These differences may have been due to the growing of the plants in different environments, different variety of plants and harvesting of leaf samples at different times.

In the current study, the effects of FA and MA on the *in vitro* true digestibility and protozoa count of four alternative feedstuffs for cattle were varied considerably. In a trial (4) in which these acids were used in conjunction with paddy straw, no concentrations had a statistically significant effect on IVTD (P > 0.05). In another study reported that rumen digestion and sodium retention were not affected by adding a salt of MA to corn silage (21). Furthermore, Ebrahimi et al. (12) reported that both FA and MA had no effect on digestibility as measured through DM, OM, NDF and ADF. Similarly, in our study, both FA and MA did not have a significant effect on IVTD values when they were used in combination with *P. laurocerasus* leaves (P>0.05).

In our study, the fact that FA dramatically reduces the *in vitro* digestion values of acacia leaves may be related to its high level of crude protein. As a matter of fact, Chen (11) attributed the low *in vitro* digestion values of *R. pseudoacacia* leaves to higher levels of crude protein compared to carbohydrate levels, despite low levels of NDF and ADF. They also pointed out that the *R. pseudoacacia* leaves had high level of condensed tannin and lectin that could prevent fermentation. A similar description of the *in vitro* digestive properties of *R. pseudoacacia* leaves are\_described by Burner et al. (6).

Sirohi et al. (33) reported that FA (0, 5, 10 and 15 mM) added to rations that included berseem, sorghum and wheat straw containing different proportions of cellulose had a positive effect on IVTD<sub>DM</sub>. In this study (33), the highest digestion percentage was seen in the group with

the lowest cellulose level and 10 mM concentration of fumaric acid. There was a significant increase (P<0.05) in microbial biomass for all cellulose levels in the berseem group. Furthermore, for the sorghum group, FA at 15 mM concentration significantly (P<0.05) decreased the IVTD<sub>DM</sub>, proportional to the amount of cellulose. In addition, the count of protozoa was significantly reduced in all sorghum and berseem groups (P < 0.05) and this effect was not correlated with the amount of cellulose (33). The fact that the data obtained in our research on IVTD<sub>DM</sub> is not compatible with the study of Sirohi et al. (33) may be attributed to the different nutrient and cell wall structure of plant materials. Sirohi et al. (33) reported that FA increased the IVTD<sub>DM</sub>, but the true digestibility values for the groups that received the highest concentrations of FA and MA were low, which is in agreement with the results of the current research. The same researchers reported that the different types of feed may have had different effects on IVTD<sub>DM</sub> values; this argument is supported by the results of our studies.

In the researches, the findings of the effects of MA administered on different doses and durations on rumen fermentation vary. Carro and Ranilla (9) reported that a 10 mM/L of malate and a 17-hour incubation period was insufficient for the complete fermentation of treatments that included corn, barley, wheat and sorghum separately. Among the feeds used, the most fermentation occurred in the corn group. However, Callaway and Martin (7) reported that the application of malate at a concentration of 7.5 mM/L resulted in complete fermentation in ruminal fluid within 10 to 24 hours. It therefore appear that no any significant (P<0.05) enhancing effects of MA on *in vitro* digestion were observed in the present study for any plants because of the low content of fermentable material they have.

According to Castillo et al. (10), organic acid salts may be more useful in facilitating rumen fermentation due to their buffering properties. Montano et al. (26)\_reported that MA added to a highly concentrated feed had no effect on ruminal digestion of OM, ADF, starch level, microbial numbers, microbial digestion and protein level. Similar findings were reported for a high roughage ration supplemented with MA (21). The high solubility of this acid contributes to the effects it has on chemical reactions (25). In the study of Kara (14) in which MA was added to corn silage at 0.5%, 1%, and 1.5%, IVTD<sub>OM</sub> was not affected (P>0.05) by the applications. This finding is supported by the results of our research. Kara (14) suggested that the effects of MA on NDF were due to the increased solubility of the cellulose in silage. Khampa et al. (16) demonstrated that Dimethyl (DL) -malate had no effect on digestion of DM, OM, CP and NDF but increased ADF digestion for cassava (P>0.05). The use of MA at high doses did not have a statistically significant effect on

*in vitro* digestion in other studies (4, 7, 9, 12). Consistent with these data, in our study, when MA was used at 0.1% dose, it was observed that *in vitro* digestion levels increased only numerically in *R. pseudoacacia*, TFW and *P. laurocerasus* leaves. Different findings obtained from *in vitro* digestion studies with different feed materials can be explained by the efficacy of antinutritional factors such as tannin and other polyphenols that affect rumen microbial activity.

In the present study, only 0.1% and 0.3% dose of FA increased protozoa counts numerically and MA showed a reducing effect on protozoa counts with increasing dose. This finding is consistent with the reports of other studies (17, 27, 32) investigating the effects of organic acids on rumen microorganisms. Ok et al. (27) reported that FA and MA have different effects on bacteria, protozoa and other microbial community. They reported that these acids had an increasing effect on rumen bacteria, but decreased the number of methanogenic archaea species that could form complexes with protozoa. Sahoo and Jena (32) reported that MA increases the number of lactate utilizing S. ruminantium in rumen, leading to a decrease in lactic acid levels. Therefore the count of ruminal protozoa may also be adversely affected due to the change in pH. This situation may be explained as the anionic effects of organic acids may adversely affect microbial life (32).

Li et al. (22) stated that the numbers of fumarateutilising bacteria (*Fibrobacter succinogenes, S. ruminantium*) did not change significantly in the presence of FA and also that DM digestibility ratios were not affected by the application. Lopez et al. (23)\_reported that the application of sodium fumarate to rumen fluid did not change the total number of bacteria during a 48-hour incubation period but increased the number of cellulolytic bacteria three fold (P<0.01).

Partanen (30) stated that the effects of organic acids on rumen bacteria vary according to the chemical properties of the acids. Gram (+) bacteria are sensitive to long chain acids whereas Gram (-) bacteria are sensitive to acids with less than 8 carbon atoms. The authors of the study suggest that these effects should also be investigated with respect to protozoans. In addition, Asanuma and Hino (3) stated that the increasing effects of higher amounts of organic acids on DM digestibility are associated with an increase in the cellulolytic bacteria population, along with an increase in H<sub>2</sub> transfer. Therefore, the author(s) of the present study recommend that the determination of true digestibility should also take into consideration the counts of protozoans, ruminal bacteria and methanogen archaea together.

In this study, it was observed that the digestion of organic matter of *C. sinensis* factory wastes and *Q. cerris* leaves could be increased by treating with 0.1% fumaric acid. It can be concluded that this application may be

beneficial in using *C. sinensis* factory wastes which are considered as undesirable material as an economical alternative feed source in ruminant nutrition. Malic acid has no negative effect on *in vitro* true digestibility values for *R. pseudoacacia, C. sinensis* factory waste, *P. laurocerasus* and *Q. cerris* in this study. On the other hand, *in vivo* studies with similar organic acids and alternative feed raw materials are needed.

## Acknowledgements

The authors would like to thank Mr. Gregory T. Sullivan for editing the English in this manuscript.

#### **Financial Support**

This research received no grant from any funding agency/sector.

## **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### References

- 1. ANKOM (2002): Operator's manual. Ankom 200/220 fiber analyzer. Ankom Technology Corporation, Fairport.
- AOAC (2006): Official Methods of Analysis, 18th edn. Association of Official Analytical Chemists, Inc., Arlington, VA.
- Asanuma N, Hino T (2000): Activity and properties of fumarate reductase in ruminal bacteria. J Gen Appl Microbiol, 46, 119–125.
- 4. Bharathidhasan A, Karunakaran A, Pugazhenthi TR, et al (2016): The effect of supplemental organic acid on methane reduction to decrease the global warming from dairy cattle. IJACSA, 3, 4.
- British Columbia Ministry of Forests (1996): Collection of Specimens. p.2-5. In: Techniques and procedures for collecting, preserving, processing, and storing botanical specimens. Res. Br., B.C. Min. For., Victoria, British Columbia.
- 6. Burner DM, Carrier DJ, Belesky DP et al (2008): Yield components and nutritive value of Robinia pseudoacacia and Albizia julibrissin in Arkansas. Agroforest Syst, 72, 51-62.
- 7. Callaway TR, Martin SA (1997): Effects of cellobiose and monensin on in vitro fermentation of organic acids by mixed ruminal bacteria. J Dairy Sci, 80, 1126–1135.
- 8. Canbolat O, Kamalak A, Özköse E, et al (2005): Effect of polyethylene glycol on in vitro gas production, metabolizable energy and organic matter digestibility of Quercus cerris leaves. LRRD, 17, 4.
- Carro MD, Ranilla MJ (2003): Effect of the addition of malate on in vitro rumen fermentation of cereal grains. Brit J Nutr, 89, 181–188.
- Castillo C, Benedito JL, Méndez J, et al (2004): Organic acids as a substitute for monensin in diets for beef cattle. Anim Feed Sci Tech, 115, 101–116.
- **11.** Chen Y, Zhao Y, Fu ZY et al (2011): Chemical composition and in vitro ruminal fermentation

characteristics of tetraploid black locust (Robinia pseudoacacia L.). Asian J Anim Vet Adv, **7**, 706-714

- 12. Ebrahimi SH, Datta MM, Heidarian V, et al (2015): Effects of fumaric or malic acid and 9,10 anthraquinone on digestibility, micobial protein synthesis, methane emission and performance of growing calves. Indian J Anim Sci, 85, 1000-1005.
- **13.** Gill JL (1978): Design and analysis of experiments in the animal and medical sciences. Vol 1. The Iowa State Univ. Press, Ames, Iowa, USA.
- Kara K (2015): In vitro methane production and quality of corn silage treated with maleic acid. Ital J Anim Sci, 14, 3994.
- **15.** Kaya E, Kamalak A (2012): Potential nutritive value and condensed tannin contents of acorns from different oak species. Kafkas Univ Vet Fak, **18**, 1061-1066
- 16. Khampa S, Wanapat M, Wachirapakorn C, et al (2006): Effect of levels of sodium DL-malate supplementation on ruminal fermentation efficiency of concentrates containing high levels of cassava chip in dairy steers. Asian Austral J Anim, 19, 368-375.
- 17. Kluge H, Broz J, Eder K (2004): Untersuchungenzum Einfluss von benzoesäureals futterzusatz auf leistungs parameter, nährstoffverdaulichkeit, n-bilanz, mikroflora und parameter des mikrobiellen stoffwechselsim verdauungstrakt von absetzferkeln. Tagungfür Schweine und Geflügelernährung Halle (Saale) Germany, 42-45.
- **18.** Koleckar V, Kubikova K, Rehakova Z, et al (2008): Condensed and hydrolysable tannins as antioxidants influencing the health. Mini-Rev Med Chem, **8**, 436-447.
- Kolver E, Aspin P (2006): Supplemental fumarate did not influence milk solids or methane production from dairy cows fed high quality pasture. Proc Nz Soc Anim Prod, 66, 409–415.
- **20.** Kumar S, Puniya AK, Puniya M, et al (2009): Factors affecting rumen methanogens and methane mitigation strategies. World J Microb Biot, **25**, 1557-1566.
- **21.** Kung L, Huber JT, Krummrey JD, et al (1982): *Influence* of adding malic acid to dairy cattle rations on milk production, rumen volatile acids, digestibility, and nitrogen utilization. J Dairy Sci, **65**, 1170–1174.
- 22. Li Z, Liu N, Cao Y, et al (2018): Effects of fumaric acid supplementation on methane production and rumen fermentation in goats fed diets varying in forage and concantrate particle size. J Anim Sci Biotechno, 9, 21.
- Lopez S, Valdes C, Newbold CJ, et al (1999): Influence of sodium fumarate addition on rumen fermentation in vitro. Brit J Nutr, 81, 59–64.
- 24. Luginbuhl JM, Mueller JP (2000): Evaluation of fodder trees for meat goats. 77-79. In: L. Gruner and Y. Chabert

(Ed). Nutrition and Feeding Strategies. 7th International Conference on Goats. Tours, France.

- 25. Martin SA, Streeter MN, Nisbet DJ, et al (1999): Effects of Dl-malate on ruminal metabolism and performance of cattle fed high-concentrate diet. J Anim Sci, 77, 1008–15.
- 26. Montano MF, Chai W, Zinn-Ware TE, et al (1999): Influence of malic acid supplementation on ruminal pH, lactic acid utilization, and digestive function in steers fed high-concentrate finishing diets. J Anim Sci, **77**, 780–784.
- 27. Ok JU, Ha DU, Lee SJ, et al (2012): Effects of organic acids on in vitro ruminal fermentation characteristics and methane emission. J Life Sci, 22, 1324-1329.
- 28. Özyılmaz N (2019): Organik ve konvansiyonel yöntemlerle üretilen çayların (Camellia sinensis) fabrika çay atıklarında besin madde içeriği ve in vitro sindirilebilirlik değerlerinin belirlenmesi. Yüksek Lisans Tezi, Ondokuz Mayıs Üniversitesi Sağlık Bilimleri Enstitüsü, Samsun.
- **29.** Parissi ZM, Abraham EM, Roukos C, et al (2018): Seasonal quality assessment of leaves and stems of fodder ligneous species. Not Bot Horti Agrobo, **6**, 426-434.
- 30. Partanen K (2001): Organic acids their efficacy and modes of action in pigs. pp. 201-218. In: Piva E, Knudsen B, Lindberg JE (Ed), Gut environment of pigs. Nottingham University Press, Nottingham, UK.
- **31.** Ranilla MJ, Carro MD, Valdés C, et al (1997): A comparative study of ruminal activity in Churra and Merino sheep offered alfalfa hay. Anim Sci, 65, 121-128.
- **32.** Sahoo A, Jena B (2014): Organic acids as rumen modifiers. IJSR, **3**,11.
- **33.** Sirohi SK, Pandey P, Goel N (2012): Response of fumaric acid addition on methanogenesis, rumen fermentation, and dry matter degradability in diets containing wheat straw and sorghum or berseem as roughage source. ISRN Vet Sci, 1-8.
- **34. SPSS** (2007): Statistical packages for social science. Version 21, SPSS In., Illinois, USA.
- **35.** Stukelj M, Valencak Z, Krsnik M, et al (2010): The effect of the combination of acids and tannin in diet on the performance and selected biochemical, haematological and antioxidant enzyme parameters in grower pigs. Acta Vet Scand, **52**, 19.
- **36.** Tieman TT, Lascano CE, Kreuzer M, et al (2008): The ruminal degradability of fibre explains part of the low nutritional value and reduced methanogenesis in highly tanniniferous tropical legumes. J Sci Food Agr, **88**, 1794–1803.
- Van Soest PJ, Robertson JB, Lewis BA (1991): Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. J Dairy Sci, 74, 3583-3597.

Ankara Üniv Vet Fak Derg, **67**, 193-199, 2020 DOI: 10.33988/auvfd.640288

# Insulin-like growth factor-1 gene expression in the endometrium of repeat breeder cows after the administration of presynch-10 and ovsynch protocol

# Ali Reha AĞAOĞLU<sup>1,a,⊠</sup>, Özgecan KORKMAZ AĞAOĞLU<sup>2,b</sup>, Özge SİDEKLİ<sup>2,c</sup>, Mesih KOCAMÜFTÜOĞLU<sup>1,d</sup>

<sup>1</sup>Burdur Mehmet Akif Ersoy University, Faculty of Veterinary Medicine, Department of Obstetrics and Gynecology, Burdur; <sup>2</sup>Burdur Mehmet Akif Ersoy University, Faculty of Veterinary Medicine, Department of Animal Science, Burdur, Turkey. <sup>a</sup>ORCID: 0000-0002-6545-8800; <sup>b</sup>ORCID: 0000-0002-7414-1725; <sup>c</sup>ORCID: 0000-0002-4891-1968; <sup>d</sup>ORCID: 0000-0003-3228-6835

> Corresponding author: rehaagaoglu@mehmetakif.edu.tr Received date: 30.10.2019- Accepted date: 30.12.2019

**Abstract:** The aim of this study was to investigate the effects of using presynch-10 and ovsynch synchronization protocols on endometrial expression and serum levels of insulin-like growth factor-1 (*IGF-1*) that thought to play roles during endometrial healing process in cows diagnosed with repeat breeder syndrome (RBS) and subclinical endometritis (SE). A total of 40 Holstein-Friesian cows identified as having RBS and SE were used. The animals in Group 1 (G1, n=20) were applied presynch-10 and ovsynch furthermore, biopsy and blood samples were collected both before and after administration. Biopsy and blood samples were taken from the animals in Group 2 (G2, n=20) without performing any synchronization. The *IGF-1* gene expression profiles were determined in all samples using RT-qPCR. Even though the level of *IGF-1* expression increased in samples taken after synchronization in G1, it was not statistically significant (P>0.05). There was no statistical difference in serum *IGF-1* levels between G1 and G2 (P>0.05). In conclusion, the expression of the *IGF-1* gene in the endometrial tissue of RBS cows with SE was identified, but the evidence suggests that the synchronization protocols in question had no significant effect on the gene expression profile being investigated.

Keywords: Gene expression, IGF-1, ovsynch, presynch-10, repeat breeder

# Presynch-10 ve ovsynch protokolleri uygulamasından sonra repeat breeder ineklerde endometriyal insülin benzeri büyüme faktörü-1 gen ekspresyonu

**Özet:** Bu çalışmada; repeat breeder sendromu (RBS) olan ve subklinik endometritis (SE) olduğu belirlenen ineklerde uygulanan presynch-10 ve ovsynch senkronizasyon protokollerinin, endometriyal iyileşme sürecinde rol oynadığı düşünülen insülin benzeri büyüme faktörü-1'in (*IGF-1*) endometriyal ekspresyonu ve serum düzeyleri üzerine etkilerinin araştırılması amaçlanmıştır. Çalışmada RBS olan ve SE tespit edilmiş toplam 40 baş Holstein-Friesian inek kullanılmıştır. Grup 1'de bulunan hayvanlara (G1, n=20), presynch-10 ve ovsynch uygulanmış, ayrıca uygulama öncesi ve sonrasında biyopsi ve kan örnekleri alınmıştır. Grup 2'de bulunan hayvanlara (G2, n=20) herhangi bir senkronizasyon uygulaması yapılmadan sadece biyopsi ve kan örnekleri alınmıştır. Tüm örneklerde RT-qPCR ile *IGF-1* geninin ekspresyon profilleri belirlenmiştir. G1'de senkronizasyon sonrası alınan örneklerde *IGF-1* ekspresyon düzeyinde bir yükselme şekillenmiş olsa da istatistiksel olarak bir önem bulunamamıştır (P>0,05). G1 ve G2 arasında serum *IGF-1* seviyesi açısından da istatistiksel bir fark oluşmamıştır (P>0,05). Sonuç olarak; SE tespit edilen RBS'li ineklerin endometriyal dokularında *IGF-1* geninin eksprese olduğu belirlenmiş ancak kullanılan senkronizasyon protokollerinin, incelenen genin ekspresyon profili üzerine anlamlı bir etkisinin olmadığı sonucuna varılmıştır.

Anahtar sözcükler: Gen ekspresyonu, IGF-1, ovsynch, presynch-10, repeat breeder.

# Introduction

Repeat breeder syndrome (RBS) is a substantial problem that causes infertility in the dairy herds (36) and occurs at the frequency of 7-36% (19, 38, 39). RBS causes significant economic losses since especially it prolongs calving-conception interval and increases the culling rates of cows in herds (14, 34). Even though there are a number of factors that play a role in the etiology of RBS, the major causes can be listed as subclinical endometritis (SE), luteal deficiencies, delayed ovulation and malnutrition, which cause embryonic death or fertilization errors (1, 19, 23, 38). One of these problems is luteal insufficiency that leads to low serum progesterone levels and embryonic deaths (20). Low progesterone levels may lead to lower interferon-tau secretion of the embryo, which causes early lysis of the corpus luteum (21). Similarly, endocrine asynchronization, which means that hormones are not released when they should be, causes delayed or absence preovulatory luteinizing hormone (LH) surge. Depending on this situation; progesterone level remains low and, embryonic deaths and RBS may occur (3, 4, 11). Another condition that causes RBS is SE. It is a superficial inflammation characterized by degradation and congestion in the epithelial cells of the cows' endometrium and infiltration of inflammatory cells without any purulent discharge and uninterrupted continuation of the sexual cycle (15). Cows with SE do not show any clinical signs of endometritis and they have very low chances to become pregnant after artificial insemination (AI). Studies have shown that the prevalence of SE in cows with RBS varies between 15% and 50% (12, 24, 28). A number of factors play a role in the development of these disorders that cause RBS. In this context, reproductive activities in cows, as in all other mammals, are regulated by the harmonious interaction of the hypothalamic-hypophyseal-ovarian axis. It has been revealed that during this interaction, growth hormone and insulin-like growth factor play crucial roles, like as hypothalamic-hypophyseal-ovarian axis derived hormones (18, 31). IGF-1 is synthesized and secreted in the liver due to the effect of growth hormone (26). IGF-1 does not only affects ovarian functions, oocyte quality and viability of the embryo, but also the endometrium (33). For instance; it was determined the members of the IGF system that have a stimulating effect on the regeneration of damaged epithelial cells in the process of tissue repair and in the production of new cells (5). Endometrial cells are also specific epithelial cells of the uterus and, IGFs are expressed in varying amounts in endometrium in various species such as rodents (8), pigs (29), cattle (7) and, sheep (32). Moreover, it has been demonstrated that IGFs have both progressive or regressive effects on uterine tissue depending on the stage of the sexual cycle. In this context, it was determined that post-partum cows with low IGF-1 concentrations are more sensitivity to endometritis (35). There are many causative factors in the etiology of bovine RBS, therefore, treatment methods are also preferred according to the etiology. In the treatment for RBS that develops due to hormonal dysfunction or SE, it should be focused on the treatment of endometritis or regulation of ovarian function. From this perspective, one of the preferred treatments of RBS with endometritis is the application of the "ovsynch" protocol which controls ovarian functions (25). The ovsynch method is preferred the treatment of RBS with SE because it ensures regular follicular development and ovulation, stimulates the regeneration of endometrial epithelium. Furthermore, it

stimulates the immune system due to the resulting estrus and, the prostaglandins used in this method are thought to stimulate the local immune response of the uterus. The aim of this study was to determine how endometrial *IGF-1* gene expression levels and serum *IGF-1* levels in RBS cows cytologically diagnosed with SE, are affected by presynch-10 + ovsynch applications.

#### **Material and Methods**

Animals: The animal material for the study has consisted of 40 Holstein-Friesian cows bred on dairy farms and identified as having RBS. All of the procedures conducted on the cows were approved by the Local Ethics Committee on Animal Experiments (Approval no: 2019-519). Repeat breeder cows were identified by examining farm records and conducting gynecological examinations. Cows that had given birth at least once and were inseminated in three consecutive estrus cycles yet failed to get pregnant were identified based on farm records. Rectal palpations and USG examinations were conducted on these cows to determine whether there were any pathological issues in the uterus or ovaries. Clinical pathological conditions were excluded from the study. The cows included in the study had corpus luteum on the ovaries and body condition scores between 3.0-3.5 under the scoring system with 1 to 5 scales. An endometrial cytology sample was taken from all of the cows included in the study to determine the endometrial polymorphnuclear lymphocyte (PMNL) ratio, an endometrial biopsy to determine gene expressions, and a blood sample from the tail vein to measure serum IGF-1 levels. The days in milk (DIM) information and the calving-first service interval of cows were determined based on farm records. In the cytological evaluation, cows with PMNL ratios  $\geq$ 3% were considered to be SE (28) and included in the study. The cows were randomly divided into two groups (G1 and G2).

**Experiment 1:** The presynch-10 +ovsynch protocol was administered to the cows in Group 1 (G1, n=20) and the same samples were repeated 10 days after the last injection (Figure 1).

*Experiment 2:* No synchronization protocol was administered to the cows in Group 2 (G2, n=20). After waited for the length of time required by the synchronization protocols administered to G1 (34 days), estruses were monitored. Ten days after estruses were determined, rectal examinations were performed and all samples were collected again from cows with corpus luteum (Figure 2).

*Collection of endometrial cytology samples:* A cytobrush (Cytobrush<sup>®</sup>, Ref.17214/2960, Minitube, Germany) was used to determine the neutrophil concentration. In order to prevent the cytobrush from being contaminated during this procedure, it was inserted



Figure 1. Presynch-10 + ovsynch protocol

PGF2α: Cloprostenol, 500 μg, im, Estrumate<sup>®</sup>, Intervet; GnRH: Buserelin acetate, 10 μg, im, Receptal<sup>®</sup>, Intervet; VI: Vaginal Inspection; R/USG: Rectal ultrasonography; RP: Rectal palpation; E/S: Endometrial swab; E/B: Endometrial Biopsy; Syringes: collecting blood samples; Stars: collection tissue samples.



Figure 2. Control group

VI: Vaginal Inspection; R/USG: Rectal ultrasonograpy; RP: Rectal palpation; E/S: Endometrial swab; E/B: Endometrial Biopsy; Syringes: collecting blood samples; Stars: collection tissue samples.

into the uterus lumen inside a stainless-steel metal catheter approximately 50 cm long. After the sample was taken, it was smeared on a slide with the brush. The collected samples were stained using the May Grunwald Giemsa staining method and 200 cells were counted in each preparation microscopically.

RNA isolation and cDNA synthesis: The endometrial biopsy samples were collected from the dorsal wall and intercaruncular site of the uterus with biopsy forceps (Kruuse Biopsy instrument, Denmark). The collected tissues were placed in 1.5 ml microcentrifuge tubes and frozen in liquid nitrogen. The samples were taken to the laboratory in a nitrogen tank and stored in a deep freeze at -86°C until the analyses were performed. Homogenization of frozen endometrial tissue samples was performed using the TRIzol® (Invitrogen, USA) method. DNA digested with DNase-I (Fermentas). And, the commercial kit (Thermo Scientific RevertAid<sup>TM</sup> First Strand cDNA Synthesis Kit) was used to cDNA synthesis.

*RT-qPCR:* YHWAZ was used as the reference gene as it is most stable and suitable for the study (2). And, a pair of specific primers [Forward: 5'-

CTGAGCAAGGAGCTGAATTATC-3' and Reverse: 5'-CTCTGTATTCTCGAGCCATCT-3 for YHWAZ (NM\_174814.2)] was designed using the idt/PrimerQuest and ncbi/tools/primer-blast software to amplify fragment of the YHWAZ gene consisting of 162 bp. (2). For the IGF-1 gene, the primers designed by Yoshida et al. (37) [Forward: 5'-TCAGTTCGTGTGCGGAGACA-3' and Reverse: 5'-ACTTCCTTCTGAGCCTTGGG-3' for IGF-1(NM001077828)] were used to amplify a region consisting of 222 bp. Primer efficiencies were determined using two-fold serial dilutions of cDNA. The expression profiles for YHWAZ and IGF-1 genes were identified with RT-qPCR, using a ROCHE Nano Lightcycler<sup>®</sup>. The reaction was prepared using SYBR Green Master Mix (2X). This procedure was performed twice. Melting curve analyzes were performed.

*Hormone measurements:* Hormone analyses were performed in a private laboratory with international accreditation (TÜRKAK, TS EN ISO/IEC 17025:2005 Experimental Laboratory) using ECLIA<sup>®</sup> (Electrochemiluminescence Immunoassay) for estradiol and progesterone, and *IGF-1* specific ELISA kit (CusaBio<sup>®</sup>, Bovine insulin-like growth factor 1, ELISA Kit). *Statistical analysis:* MINITAB 16.0<sup>©</sup> was used for the statistical analysis of the data. It was later normalized with the data reference gene obtained from the qPCR analyses based on the delta delta Ct mathematical model reported by Livak and Schmittengen (16). The paired t test was used for data collected from the same group while the T test and One-way ANOVA were used to identify differences between the groups. Box plot and Interval Plot graphics were created for a visual representation of the data.

## Results

The calving-first service interval and the DIM numbers were  $85.95\pm19.79$  and  $205.6\pm55.60$  for G1 and  $90.05\pm21.77$  and  $236.25\pm54.51$  for G2 respectively. The mean DIM numbers for the cows in G1 and G2 were  $205.60\pm55.60$  (min. 150, max. 349), so cows with PMNL

>%3 during the cytological evaluation were considered to have SE (31).

*Expression of IGF1 gene:* The *IGF-1* gene was expressed in all samples. The resulting RT-qPCR products using YHWAZ as the reference gene and the *IGF-1* gene in bovine endometrium were separated with electrophoresis on 2% agarose gel and stained with ethidium bromide (Figure 3).

When the *IGF-1* expression profile was examined depending on days of applied synchronization protocols, there was more expression in the endometrial biopsy samples taken after synchronization. However, it was not statistically significant (P>0.05). Expression levels between the biopsy samples taken in G2 remained almost the same. The Box Plot graphics that compare the data obtained after normalization of the *IGF-1* gene RT-qPCR results with each sampling group are provided in Figure 4.



**Figure 3.** Expression of YHWAZ and IGF-1 genes at mRNA levels in bovine endometrium on 2% agarose gel. Lanes 1-4: PCR products of IGF-1 (222 bp), lane 5: YHWAZ (162 bp), M: 100 bp DNA ladder



**Figure 4.** Box plot graphics showing the differences between sampling groups. EBS: endometrial biopsy sample.



# Interval Plot of IGF1 ng/ml

Figure 5. Serum IGF-1 levels according to administration days in G1. a:b, P<0.05; c:b, P<0.05, 1/EBS1/G1: Group 1 Endometrial biopsy sample 1; 2/Pre-PG: presynch-10 1st Prostaglandin injection; 3/Ovs-1<sup>st</sup> GnRH: ovsynch 1<sup>st</sup> GnRH injection; 4/Ovs-2<sup>nd</sup> GnRH: ovsynch 2<sup>nd</sup> GnRH injection; 5/Ovs.S-12: After ovsynch 12 hours; 6/EBS2/G1: Group 1 Endometrial biopsy sample 2.

When serum IGF-1 levels were examined, it was showed that the IGF-1 level of 3.99 ng/ml on the day the first sample was taken in G1 fell to 3.64 ng/ml on the day that the second GnRH administration of ovsynch was performed and that this decline was statistically significant (P<0.05). No statistically significant difference was found between the IGF-1 levels (4.04±0.92 pg/ml) measured on the day when the 1<sup>st</sup> PGF2α injection of presynch-10 was performed and on the day when 2<sup>nd</sup> GnRH injection of ovsynch (3.64±0.54 pg/ml) (P<0.05). There was also no statistically significant difference in terms of serum IGF-1 levels between the day on which the first sample was taken and the days on which the second sample was taken (P>0.05) (Figure 5).

No statistically significant difference was found between IGF-1 levels in the blood samples collected on the days when the first and second samples were taken in G2 (1<sup>st</sup> IGF-1 value: 3.99±0.73, 2<sup>nd</sup> IGF-1 value: 4.09±0.75).

## **Discussion and Conclusion**

The healing of damaged epithelial tissue consists of proliferation and remodeling processes (10). IGF-1 is an important factor that functions in the tissue repair process by stimulating epithelization and regeneration (5). Physiological regeneration of endometrial tissue takes place during the post-partum period in cows. Increased endometrial IGF-1 gene expression has been determined during the post-partum period in cows, and it has been demonstrated that the IGF-1 may play an important role in the endometrial repair process (17). Because a certain amount of endometrial injury occurs in cases of bovine SE, there might be a connection between *IGF-1* and the injury and subsequent healing that occurs. In a study of the issue, endometrial IGF-1 expression was reported to be higher in cows diagnosed with metritis or clinical endometritis than in healthy cows or cows with SE. The same study found that when RBS cows with SE were compared with cows that did not have SE, endometrial IGF-1 expression was higher in the cows with SE. It was concluded that both of these findings could be related to the tissue damage in the endometrium and the healing process (13). Ovarian steroids are reported to have an effect on endometrial IGF-1 expression. In cows, it has been shown that endometrial IGF-1 expression peaks during the estrus stage when the estradiol level is highest in the estrus cycle (27). In this study, presynch-10 and ovsynch administrations given for treatment purposes to RBS cows induced ovulation, meanwhile, increased serum estradiol levels due to the follicular development that occurs. When the IGF-1 expression profile based on days of administration examined that showed an increasing in IGF-1 expression but the difference between the biopsy samples taken before and after synchronization in G1 was not statistically significant. It is thought that the increase in the expression of IGF-1 may be associated with the fact that the endometrium is exposed to estradiol as a result of the synchronization protocols applied.

In a study that examined bovine serum IGF-1 levels during ovsynch administration that the serum IGF-1 levels decrease after the 1st GnRH injection of ovsynch but later increase until day 9 after the PGF2 $\alpha$  in cows and this occurred due to follicular development (6). It has also been demonstrated that IGF-1 is synthesized not only in the follicles of in bovine ovaries but in luteal tissue as well. However, after lysis of the luteal structure due to PGF2 $\alpha$ 

stimulation, there is no significant decrease in *IGF-1* expression has been demonstrated (22). Moreover, it has been shown that ovulation occurred before the peak of plasma estradiol level at the time of second GnRH injection during ovsynch administration in cows (30). This finding indicates that the *IGF-1* concentration in circulation is controlled by several different mechanisms.

It is thought that the reason why there is no difference in serum IGF-1 levels between on the day of the first PGF2 $\alpha$  of presynch-10 and on the day of the second GnRH of ovsynch may be due to this complex biological mechanism. We think that this situation may be the reason that a correlation could not be established between serum IGF-1 levels and endometrial IGF-1 expression levels.

In conclusion, in addition to this study, which examined the IGF-1 expression profile in conjunction with administration of the presynch-10 + ovsynch protocol for the purpose of treating SE, new studies that investigate all of the elements of the IGF system and not only endometrial expression levels but also expression in follicles, corpus luteum and liver tissue could help develop new approaches when it comes to solving the repeat breeder problem, which is important to livestock breeders.

#### Acknowledgements

This study was presented in the Turkish Veterinary Gynecology Association, VIII. National & II. International Congress, 10-13 October 2019, Antalya.

# **Financial Support**

This study was supported by the TUBITAK (Project No. 1130590).

# **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### References

- 1. Adnane M, Kaidi R, Hanzen C, et al (2017): *Risk factors* of clinical and subclinical endometritis in cattle: a review. Turk J Vet Anim Sci, **41**, 1-11.
- 2. Agaoglu AR, Agaoglu OK, Aslan S, et al (2020): The effects of presynch-10 and ovsynch on some endometrial toll and nod-like receptor gene expressions in repeat breeder cows. Kafkas Univ Vet Fak Derg, 26, 111-120.
- **3.** Albihn A, Gustafsson H, Hurst M, et al (1991): Embryonic ability to prolong the interestrous interval in virgin and repeat breading heifers. Anim Reprod Sci, 26, 193-210.
- 4. Albihn A, Gustafsson H, Rodriguez-Martines H, et al (1989): Development of day 7 bovine demi embryos transferred into virgin and repeat breeder heifers. Anim Reprod Sci, 21, 161-176.

- Bitar MS (2000): Insulin and glucocorticoid-dependent suppression of the IGF-I system in diabetic wounds. Surgery, 127, 687–695.
- Burns PD, Spitzer JC, Henricks DM (1997): Effect of dietary energy restriction on follicular development and luteal function in nonlactating beef cows. J Anim Sci, 75, 1078-1086.
- 7. Geisert RD, Lee CY, Simmen FA, et al (1991): Expression of messenger RNAs encoding insulin-like growth factor-I, -II, and insulin-like growth factor binding protein-2 in bovine endometrium during the estrous cycle and early pregnancy. Biol Reprod, 45, 975–983.
- Ghahary A, Chakrabarti S, Murphy LJ (1990): In situ localization of the sites of synthesis and action of insulinlike growth factor-1 in the rat uterus. Mol Endocrinol, 4, 191–195.
- Giudice LA, Dsupin BA, Jin IH, et al (1993): Differential expression of messenger ribonucleic acids encoding insulin-like growth factors and their receptors in human uterine endometrium and decidua. J Clin Endocrinol Metab, 76, 1115–1122.
- Grazul-Bilska AT, Johnson ML, Bilski JJ, et al (2003): Wound healing: the role of growth factors. Drugs Today, 39, 787–800.
- **11.** Gustafsson Hi Larsson K, Kindahl H, et al (1986): Sequential endocrine changes and behaviour during estrus and metestrus in repeat breeder and virgin heifers. Anim Reprod Sci, **19**, 261-273.
- 12. Janowski T, Barański W, Łukasik K, et al (2013): Prevalence of subclinical endometritis in repeat breeding cows and mRNA expression of tumor necrosis factor α and inducible nitric oxide synthase in the endometrium of repeat breeding cows with and without subclinical endometritis. Pol J Vet Sci, 16, 693-699.
- **13.** Kasimanicam R, Kasimanicam V, Kastelic JP (2014): Mucin 1 and cytokines mRNA in endometrium of dairy cows with postpartum uterine disease or repeat breeding. Theriogenology, **81**, 952-958.
- 14. Khair A, Asuduzzaman M, Sultana Z, et al (2018): Economic benefit in repeat breeder cows using intrauterine infusion of penicillin and estrus synchronization followed by timed artificial insemination. J Adv Vet Anim Res, 5, 454– 458.
- **15.** LeBlanc SJ, Duffied TF, Leslie KE, et al (2002): Defining and diagnosing postpartum clinical endometritis and its impact on reproductive performance in dairy cows. J Dairy Sci, **85**, 2223-2236.
- **16.** Livak KJ, Schmittgen TD (2001): Analysis of relative gene expression data using real-time quantitative PCR and the  $2^{-\Delta \Lambda T}$  method. Methods, **26**, 402-408.
- 17. Llewellyn S, Fitzpatrick R, Kenny DA, et al (2008): Endometrial expression of the insulin-like growth factor system during uterine involution in the postpartum dairy cow. Dom Anim Endocrinol, 34, 391–402.
- Lucy MC, Bilby CR, Kirby CJ, et al (1999): Role of growth hormone in development and maintenance of follicles and corpora lutea. J Reprod Fertil, 54, 49–59.
- **19.** Mandefro M, Negash G (2014): Repeat breder syndrome in dairy cows: Influence of Breed and Age on Its Prevalence. World J of Agric Sci, **10**, 200-203.

- **20.** Mann GE, Green MP, Sinclair KD, et al (2003): *Effects* of circulating progesterone and insulin on early embryo development in beef heifers. Anim Reprod Sci, **79**, 71–9.
- **21.** Mann GE, Lamming GE (2001): Relationship between maternal endocrine environment, early embryo development and inhibition of the luteolytic mechanism in cows. Reproduction, **121**, 175–80.
- 22. Neuvians TP, Pfaffl MW, Berisha B, et al (2003): The mRNA express, one of the members of the IGF system in bovine corpus luteum during induced luteolysis. Dom Anim Endocrinol, 25, 359-372.
- **23.** Parkinson TJ (2009): The repeat breeder syndrome. 463-466. In: DE Noakes, TJ Parkinson, GCW England, (Eds), Veterinary reproduction and obstetrics. 9th ed., Saunders Elsevier, Edinburgh.
- 24. Pothmann H, Prunner I, Wagener K, et al (2015): The prevalence of subclinical endometritis and intrauterine infections in repeat breeder cows. Theriogenology, 83, 1249-1253.
- **25.** Pursley JR, Mee MO, Wiltbank MC (1995): Synchronization of ovulation in dairy cows using PGF2α and GnRH. Theriogenology, **52**, 1067-1078.
- 26. Radcliff RP, McCormack BL, Crooker BA, et al (2003): Plasma hormones and expression of growth hormone receptor and insulin-like growth factor-I mRNA in hepatic tissue of periparturient dairy cows. J Dairy Sci, 86, 3920-3926.
- 27. Robinson RS, Mann GE, Gadd TS, et al (2000): The expression of the IGF system in the bovine uterus throughout the oestrous cycle and early pregnancy. J Endocrinol, 165, 231-243.
- Salasel B, Mokhtari A, Taktaz T (2010): Prevalence, risk factors for and impact of subclinical endometritis in repeat breeder dairy cows. Theriogenology, 74, 1271-1278.
- **29.** Simmen RCM, Simmen FA, Hofig A, et al (1990). Hormonal regulation of insulin like growth factor gene expression in pig uterus. Endocrinology, **127**, 2166-2174.

- **30.** Souza AH, Gümen A, Silva EPB, et al (2007) Supplementation with estradiol-17  $\beta$  before the last gonadotropin-releasing hormone injection of the ovsynch protocol in lactating cows. J Dairy Sci, **90**, 4623-4634.
- **31.** Spicer LJ, Echternkamp SE (1995): The ovarian insulin and insulin-like growth factor system with an emphasis on domestic animals. Dom Anim Endocrinol, **12**, 223–245.
- **32.** Stevenson KR, Gilmour RS, Wathes DC (1994): Localization of insulin-like growth factor-I (IGF-I) and -II messenger ribonucleic acid and type 1 IGF receptor in the ovine uterus during the oestrous cycle and early pregnancy. Endocrinology, **134**, 1655-1664.
- **33.** Totey SM, Pawshe CH, Rao KBC (1996): In vitro maturation of buffalo oocytes: Role of insulin and its interaction with gonadotrophins. J Reprod Fertil, **50**, 113-119.
- **34. Valdmann M, Kurykin J, Kaart T, et al** (2018): Relationships between plasma insulin like growth factor-1 and insülin concentrations in multiparous dairy cows with cytological endometritis. Vet Rec, **183**, 1-7.
- **35.** Wathes DC, Cheng Z, Fenwick MA, et al (2011): Influence of energy balance on the somatotrophic axis and matrix metalloproteinase expression in the endometrium of the postpartum dairy cow. Reproduction, 141, 269-281.
- **36.** Wodaje HB, Mekuria TA (2016): *Risk factors of repeat breeding in dairy cattle.* Adv Biol Res, **10**, 213-221.
- **37.** YoshidaY, Miyamura M, Hamano S, et al (1998): Expression of growth factor ligand and their receptor mRNAs in bovine ova during in vitro maturation and after fertilization in vitro. J Vet Med Sci, **60**, 549-554.
- **38.** Yusuf M, Nakao T, Ranasinghe RB, et al (2010): Reproductive performance of repeat breeders in dairy herds. Theriogenology, **73**, 1220-1229.
- **39.** Zobel R, Tkaltcic S, Buic V, et al (2011): Repeat breeder syndrome in dairy cows: influence of breed and age on its prevalence and the success of a hormone therapy. Turk J Vet Anim Sci, **35**, 405-411.

Ankara Üniv Vet Fak Derg, 67, 201-204, 2020 DOI: 10.33988/auvfd.603515

# Short Communication / Kısa Bilimsel Çalışma

# PCR detection of Mycobacterium genavense DNA in fecal samples of caged birds

# Orkun BABACAN<sup>1,2,a,∞</sup>, Bülent BAŞ<sup>2,b</sup>, Barış SAREYYÜPOĞLU<sup>2,c</sup>

<sup>1</sup>Balıkesir University, Kepsut Vocational School, Department of Veterinary, Kepsut, Balıkesir; <sup>2</sup>Ankara University, Faculty of Veterinary Medicine, Department of Microbiology, Dışkapı, Ankara, Turkey <sup>a</sup>ORCID: 0000-0003-0258-1825; <sup>b</sup>ORCID: 0000-0001-9992-8738; <sup>c</sup>ORCID: 0000-0002-2212 2610.

Corresponding author: orkun.babacan@balikesir.edu.tr Received date: 07.08.2019- Accepted date: 04.11.2019

Abstract: In this study, pathogenic mycobacteria were investigated in fecal samples of caged birds by PCR. A total of 47 feces samples collected from 4 different aviaries in Ankara. DNA extraction from fecal samples was performed with a commercial kit using spin column technology. PCR was performed with designed primers respectively amplifying 274 base pairs (bp), 128 bp, 102 bp and 219 bp nucleotide sequences of specific genes (16SrRNA, ISI245, IS901 and hypothetical 21kDa protein gene) of Mycobacterium spp., Mycobacterium avium complex (MAC), Mycobacterium avium subsp. Avium and Mycobacterium genavense, respectively. Five samples were positive and harbored the sequence for the Mycobacterium spp., of 4 of these 5 samples was identified as M. genavense by PCR. As a conclusion of this study, which is the first announcement of the detection of M. genavense DNA in fecal samples of caged birds in Turkey, PCR was seen to be a rapid, sensitive, and a reliable method in detection of avian mycobacteriosis.

Keywords: Bird, feces, mycobacteriosis, Mycobacterium genavense

# Kafes kuşu dışkı örneklerinde Mycobacterium genavense DNA'sının PZR ile saptanması

Özet: Bu çalışmada, kafes kuşlarının dışkı örneklerinde PCR ile patojenik mikobakterilerin varlığı incelendi. Çalışmanın materyalini Ankara'da 4 farklı kushaneden toplanan 47 dıskı örneği olusturdu. Dıskı örneklerinden DNA ekstraksiyonu, spin kolon teknolojisi kullanılarak ticari DNA ekstraksiyon kiti ile yapıldı. PCR, Mycobacterium spp., Mycobacterium avium complex (MAC), Mycobacterium avium subsp. avium ve Mycobacterium genavense'nin spesifik genlerinin (16SrRNA, ISI245, IS901 ve hipotetik 21kDa protein geni) sırasıyla 274 baz çiftini (bp), 128 bp, 102 bp ve 219 bp nükleotit sekanslarını çoğaltan tasarlanmış primerlerle gerçekleştirildi. PZR sonucunda, 5 dışkı örneği Mycobacterium spp. için pozitif bulundu. Örneklerin 4'ü PZR'da M. genavense olarak tanımlandı. Türkiye'deki kafes kuşlarının dışkı örneklerinde M. genavense DNA'sının saptanmasına yönelik ilk duyuru olan bu çalışmanın sonucu olarak, PZR'nin kanatlı mikobakteriyozisi tespitinde hızlı, duyarlı ve güvenilir bir yöntem olduğu görülmüştür. Anahtar sözcükler: Dışkı, kuş, mikobakteriyozis, Mycobacterium genavense

Avian tuberculosis is a chronic wasting disease in cages, exotic, zoo and wild birds in the world. In recent years, for exotic birds, Mycobacterium genavense has been reported as agent of tuberculosis (2,12,16,17,18,20,22). M. genavense, in bird species, especially avian Passeriformes and Psittaciformes, described as the most frequent etiologic agent of avian tuberculosis (4,14). M. genavense was first described on AIDS patients in 1990 (17). In people with weakened immune systems, children and other animal species including the dog could cause the infection (1,3,7,8,11,14,20). The disease occurs sporadically and can also be seen in intensive breeding companies (6).

Mycobacteria can exist for months in the environment. The main sources of infection are infected animals, contaminated soil and water (12,20). M. genavense is transmitted through the digestive system by fecal-oral route (10). Feces of infected birds are one of the important source for infection. In intensive breeding, stool contains a large number of bacteria for bird to bird contact (9,16,20). People may inhale the bacteria as a result of direct contact with infected birds (9,10).

Mycobacteriosis generally occurs in adult birds more than young birds. Avian tuberculosis causes direct and indirect economic losses (19).

М. shows similar clinical and genavense histopathological lesions of Mycobacterium avium complex (14,15,17). The deaths occur after several months because it is a chronic infection. Infected birds usually are greater than one year of age (12). Also, they are weakened with a weight loss (5,12,15,20). Although some infected birds, which may exhibit a normal appearance and behavior, could be found dead. The most common route of transmission is by fecal-oral route. Because of that, lesions can be seen in the digestive tract (20).

*M. genavense* is a fastidious and slow growing bacterium (5). For this reason, it is very difficult to culture this bacteria (11,17,20). Because of this, the diagnosis based on Ziehl-Neelsen staining of asidoresistance bacteria from infected samples (2,17). Since *M. genavense* is a fastidious bacterium, PCR is a rapid and accurate method for detecting the small amount of DNA from clinical samples (2,9,15,19,20). Results are obtained within hours (19). Fresh tissues and feces can be used for the diagnosis of infection in PCR method (20). Because of the risk of zoonotic infection between individuals with a weakened immune system and their animals, rapid diagnosis is very important to prevent serious infection (11).

In this study, presence of *M. genavense* DNA was investigated by PCR from cage bird feces samples.

This study was carried out in Ankara University Faculty of Veterinary Medicine Department of Microbiology. A total of 47 feces samples were collected from four different aviaries in Ankara region, Turkey. There were canaries, parrots, budgerigars, pigeon, phesant and dove birds in aviaries. The number of birds in cages ranged from 1-23. From the first aviary 10 feces samples were taken. 12, 13 and 12 feces samples were taken from the second, third and fourth aviaries, respectively. Feces samples were collected from cages floor and put into the sterile containers. The amount of each feces samples were approxiametly 25 grams. Symptoms were evaluated following clinical examinations and anamnesis was obtained from the owners. Respiratory system symptoms of the birds were changing from slight to heavy. They showed conjuctivitis and diarrhoea, some birds without any symptoms.

PCR was used for the investigation DNA of *Mycobacterium* spp., *Mycobacterium avium* complex (MAC), *M. avium* subsp. *avium* and *M. genavense* from birds feces. DNA extractions were performed with the QIAamp DNA Stool Mini Kit (Qiagen), which is works by spin column technology, from birds feces.

The primers used in this study, were designed by Barış Sareyyüpoğlu with Primer3 (21). PCR was performed with designed primers respectively amplifying 274 base pairs (bp), 128 bp, 102 bp and 219 bp nucleotide sequences of specific genes (16SrRNA, ISI245, IS901 and hypothetical 21kDa protein genes) of Mycobacterium spp., MAC, M. avium subsp. avium and Mycobacterium genavense, respectively. (Table 1). The PCR were performed for all target genes, in a total reaction volume of 25 µl, containing 2.5 µl 10x PCR buffer, 3 µl 25 mM magnesium chloride, 250 µM of each deoxynucleotide triphosphate, 1.25 U Taq DNA Polymerase, 20 pmol of each primer and 25 ng of template DNA. The reaction conditions for the M. genavense specific PCR are 35 cycles of denaturation at 94°C for one minute, annealing at 54°C for one minute and extension at 72°C for one minute, followed by a final extension step at 72°C for 7 minutes. The amplified products were detected by staining with 10 mg/ml ethidium bromide after electrophoresis at 80 V for two hours in 2% agarose gels. Results were screened from agarose gel by molecular imaging system (Gene Genius, Syngene, England).

*M. genavense* DNA used as positive control in PCR tests, was supplied by Enrico Tortoli from Regional Reference Centre for Mycobacteria, Florence, Italy. Also *M. avium* subsp. *avium* strain (German Collection of Microorganisms and Cell Cultures-DSMZ; DSM NO:44156) used as a positive control in PCR tests.

Table 1. Primers, target genes, sequences and product sizes

Primers	Name of primer	Target gene	Product size (bp)	Primer sequence
Mycobacterium spp.	Myco1F	16SrRNA	274	TGGGTACTAGGTGTGGGTTTCC
	Myco1R			TTAACCCAACATCTCACGACAC
MAC	MAC1F	ISI245	128	TGGCCGGCTCGGTACTCGTT
	MAC1R			GGCTGTGGGGGGCAATGGTTT
M. avium subsp. avium	Masa1F	IS901	102	CTCGATGCTCACCGCCATCTT
	Masa1R			ATTTCGCCCGGAGTGCACATAG
M. genavense	Mgen1F	hypothetical 21kDa	219	TGACTGGTCGTTTGAGATGAAT
	Mgen1R	protein genes		GATCGGAGGCAGTTCAATGTAG



**Figure 1.** PCR results for *Mycobacterium genavense*. M: marker (100bp) +: positive control -: negative control 1-5: feces samples.



**Figure 2.** PCR results for *Mycobacterium* spp. (274bp) M: marker (100bp) Line 1-2: positive controls, Line 3: negative control, Line 4-8: positive feces samples for *Mycobacterium* spp.

As a result of the molecular analysis of all samples, a total of 5 samples were found positive in terms of the sequence for the *Mycobacterium* spp. (Figure 2). Four of these 5 samples, positive for the *Mycobacterium* genus, were identified as *M. genavense* DNA by PCR (Figure 1).

In Turkey, *M. genavense* DNA was found for the first time in cage bird's feces with PCR. In birds, the primary source of infection is contaminated environment for mycobacterium infections. Feces of infected birds play an important role for the spread of infection among birds (20). The results of this study showed that feces are not only important for spreading of the infection but also detection of the mycobacteriosis. The important role of fecal contamination in avian tuberculosis has been seen once more with this study.

Importantly, all *M. genavense* positive specimens were detected in cages of the same aviary. This result can indicate that the contamination by horizontal route from bird to bird.

In several studies, PCR has been used to diagnose mycobacterial infections from poultry organ samples. *M. genavense* is difficult to cultivate unlike other Mycobacteria species. PCR is a rapid and reliable method for the diagnosis of *M. genavense* infections (19,20). Tell et al. (19) reported that they found *M. genavense* in 67% of poultry organ samples. The *M. genavense* primers,

which were used in this study, were designed from the hypothetical 21kDa protein gene. Ledwon et al. (14) designed the primers from the same gene for the diagnosis of *M. genavense* from the organ samples of budgerigars and identified positive samples. Patino et al. (13) reported 3 (8 %) *M. genavense* DNA in free living birds' feces using *16s rRNA* gene specific primers of *Mycobacterium genavense*. Addition, Schimitz et al. (16) were used probe targeting the hypothetical 21kDa protein gene in their real-time PCR protocol. According to of all these studies, it was seen that hypothetical 21kDa protein gene could be used for the diagnosis of *M. genavense* in PCR.

*Mycobacterium genavense*-specific primers, tested for the first time in this study in Turkey, can be used in PCR tests performed with clinical materials other than feces (liver, spleen, bone marrow, tracheal swab, air sacs, lungs, intestines) especially in patients with suspected tuberculosis in cage birds. The results of this study showed that fecal screening can be performed especially for cage birds in terms of *M. genavense* for zoonotic risks.

# Acknowledgements

The authors thanks to Dr. Enrico Tortoli (Regional Reference Center for Mycobacteria, Florence, Italy) who supplied *M. genavense* DNA which is used in this study as a positive control in PCR. This study is presented as an oral presentation in IX. Veterinary Microbiology Congress with International Participation, North Cyprus Turkish Republic, 5-7 October 2010.

## **Financial Support**

This research received no grant from any funding agency/sector.

# **Conflict of Interest**

The authors declare that there is no conflict of interest.

#### References

- 1. Aranaz A, Liebana E, Mateos A, et al (1997): *Laboratory diagnosis of avian mycobacteriosis*. Semin Avian Exotic Pet Med, **6**, 9-17.
- 2. Bougiouklis P, Brelleu G, Fragkiadaki E, et al (2005): Outbreak of avian mycobacteriosis in a flock of two-yearold domestic pigeons (Columba livia f. domestica). Avian Dis, **49**, 442- 445.
- **3.** Böttger EC, Hirschel B, Coyle MB (1993): *Mycobacterium genavense* sp. nov. Int J Syst Bacteriol, **43**, 841-843.
- 4. Hoop RK, Böttger EC, Ossent P, et al (1993): Mycobacteriosis due to mycobacterium genavense in six pet birds. J Clin Microbiol, **31**, 990-993.
- 5. Hoop RK, Böttger EC, Pfyffer GE (1996): Etiological Agents of Mycobacterioses in Pet Birds between 1986 and 1995. J Clin Microbiol, **34**, 991-992.

- 6. Ikonomopoulos J, Fragkiadaki E, Liandris E, et al (2009): Estimation of the spread of pathogenic mycobacteria in organic broiler farms by the polymerase chain reaction. Vet Microbiol, 133, 278-282.
- 7. Kiehn TE, Hoefer H, Bottger EC, et al (1996): *Mycobacterium genavense infections in pet animals*. J Clin Microbiol, **34**, 1840-1842.
- 8. Ledwon A, Szeleszczuk P, Malicka E, et al (2009): Mycobacteriosis caused by Mycobacterium genavense in Lineolated Parakeets (Bolborhynchus lineola). A case report. Bull Vet Inst Pulawy, 53, 209-212.
- 9. Lennox AM. (2007): Mycobacteriosis in companion psittacine birds: a review. J Avian Med Surg, 21, 181-187.
- Manarolla G, Liandris E, Pisoni G, et al (2009): Avian mycobacteriosis in companion birds: 20-year survey. Vet Microbiol, 133, 323-327.
- 11. Mendenhall MK, Ford SL, Emerson CL, et al (2000): Detection and differentiation of Mycobacterium avium and Mycobacterium genavense by polymerase chain reaction and restriction enzyme digestion analysis. J Vet Diagn Invest, 12, 57-60.
- **12. OIE Terrestrial Manuel** (2008): Avian tuberculosis. Available at https://www.oie.int/doc/ged/D7710.PDF. (Accessed January 10, 2019)
- **13.** Patino LCW, Monge O, Suzan G, et al (2018): Molecular Detection of Mycobacterium avium avium and Mycobacterium genavense in feces of free-living scarlet macaw (Ara macao) in Costa Rica. J Vildl Dis, **54**, 357-361.
- 14. Portales F, Realini L, Bauwens L, et al (1996): Mycobacteriosis caused by Mycobacterium genavense in birds kept in a zoo: 11- year survey. J Clin Microbiol, 34, 319-323.
- **15.** Ramis A, Ferrer L, Aranaz A, et al (1996): Mycobacterium genavense infections in canaries. Avian Dis, **40**, 246-251.
- **16.** Schmitz A, Korbel R, Thiel S, et al (2018): *High prevalence of Mycobacterium genavense within flocks of pet birds.* Vet Microbiol, **218**, 40-44.
- 17. Shitaye JL, Halouzka R, Svobodova J, et al (2010): First isolation of Mycobacterium genavense in a blue headed parrot (Pionus menstruus) important from Surinam (South America) to the Czech Republic: a case report. Vet Med-Czech, 55, 339-347.
- **18.** Sutherland M, Courtman N, Sacks P, et al (2018): Severe leukemoid response associated with Mycobacterium genavense infection in a pet budgerigar (Melopsittacus undulatus). J Exot Pet Med, **27**, 11-16.
- **19.** Tell LA, Leutenegger CM, Larsen RS, et al (2003): Realtime polymerase chain reaction testing for the detection of Mycobacterium genavense and Mycobacterium avium complex species in avian samples. Avian Dis, **47**, 1406-1415.
- 20. Tell LA, Woods L, Cromie RL (2001): *Mycobacteriosis in birds*. Rev sci tech Off int Epiz, 20, 180-203.
- **21.** Untergasser A, Cutcutache I, Koressaar T, et al (2012): *Primer3- new capabilities and interfaces.* Nucleic Acids Res, **40**, e115.
- 22. Witte CL, Hungerford LL, Papendick R, et al (2008): Investigation of characteristics and factors associated with avian mycobacteriosis in zoo birds. J Vet Diagn Invest, 20, 186–196.

Ankara Üniv Vet Fak Derg, **67**, 205-208, 2020 DOI: 10.33988/auvfd.623717

# Case Report / Olgu Sunumu

# The first case of *Bovicola (Werneckiella) ocellatus* (Piaget, 1880) (Phthiraptera: Ischnocera: Trichodectidae) on a donkey (*Equus asinus* Linnaeus, 1758) in Turkey

# Bilal DİK<sup>1,a</sup>, İpek ERDEM<sup>2,b,⊠</sup>, Aykut ZEREK<sup>2,c</sup>, Mustafa KARAGÖZ<sup>3,d</sup>, Mehmet YAMAN<sup>2,e</sup>

<sup>1</sup>Selçuk University, Faculty of Veterinary Medicine, Department of Parasitology, Konya; <sup>2</sup>Hatay Mustafa Kemal University, Faculty of Veterinary Medicine, Department of Parasitology, Hatay; <sup>3</sup>Ownerless Animal Care and Rehabilitation Center of the Hatay Metropolitan Municipality, Hatay, Turkey.

> <sup>a</sup>ORCID: 0000-0002-7553-5611; <sup>b</sup>ORCID: 0000-0002-0086-8294; <sup>c</sup>ORCID: 0000-0002-8533-387X; <sup>d</sup>ORCID: 0000-0003-3977-441X; <sup>e</sup>ORCID: 0000-0001-5399-8060

⊠Corresponding author: ipekerdem@mku.edu.tr Received date: 24.09.2019- Accepted date: 07.11. 2019

**Abstract:** In the clinical examination of a donkey, which brought to the Ownerless Animal Care and Rehabilitation Center of the Hatay Metropolitan Municipality due to neglecting and senility, local hair losses, ruffling and opacity were observed. A few louse specimens were detected on the hair shafts on the back of the donkey during the macroscopic examination. They were collected with a forceps, and preserved in a tube with alcohol 70%. Then they examined in detail in microscope, and identified as *Bovicola ocellatus* (Piaget, 1880). The presence of *B. ocellatus* on donkey has been reported for the first time in Turkey in this paper.

Keywords: Bovicola ocellatus, donkey, louse, Turkey

# Türkiye'de bir eşekte (*Equus asinus* Linnaeus, 1758) ilk *Bovicola (Werneckiella) ocellatus* (Piaget, 1880) (Phthiraptera: Ischnocera: Trichodectidae) olgusu

Özet: Hatay Büyükşehir Belediyesi Sahipsiz Hayvan Bakım ve Rehabilitasyon Merkezi'ne yaşlılık ve bakımsızlık nedeniyle getirilen bir eşeğin klinik muayenesinde tüylerinin çok karışık ve matlaşmış olduğu, yer yer döküldüğü gözlenmiştir. Yapılan makroskobik incelemede eşeğin sırt bölgesindeki tüylerin dip kısımlarında bir kaç bit saptanmıştır. Bitler bir pens yardımıyla alınmış ve içinde %70'lik alkol bulunan bir tüpte saklanmıştır. Daha sonra mikroskopta ayrıntılı olarak incelenen örnekler *Bovicola ocellatus* olarak teşhis edilmiştir. Bu makaleyle *B. ocellatus*'un Türkiye'deki varlığı ilk kez bildirilmektedir.

Anahtar sözcükler: Bit, Bovicola ocellatus, eşek, Türkiye

The lice are hemimetabolous and obligatory insects. They complete all their life on the hosts. The lice belonging to the Ischnocera and Amblycera suborders are parasitized on mammals and poultry, and they feed on the feathers or hairs of their hosts. Anopluran species has only seen in mammals, they suck blood of their hosts and cause anemia in heavy infestations. The chewing lice belonging to the genus *Bovicola*, in family Trichodectidae, suborder Ischnocera are found on cattle, sheep, horses, donkeys and deers. These species are similar to each other morphologically and they are specific to their host (1, 2, 11). The lice complete their development in onemonth average and their lives vary between 1-2 months (1, 2). Although they can be observed on the hosts during the year, however, they are seen more common in winter

months. Lice infestations cause clinical symptoms such as clutter hairs, dandruff, hair losses, pruritus and anemia. Lice are usually found on the head, neck and sides of the body. In severe infestations, they may spread on the whole body on Equidae (11).

Bovicola (Werneckiella) ocellatus (Piaget, 1880) belong to the family Trichodectidae, in the suborder Ischnocera, order Phthiraptera, class Insecta (4, 12). According to Werneck (13), Piaget described this species as Trichodectes parumpilosus var. ocellata in 1880. Later, this species was described as Trichodectes parumpilosus ocellata by Kellogg in 1908; as Trichodectes ocellatus by Harrison in 1916; as Bovicola ocellata by Bedford in 1932; as Werneckiella ocellata by Eichler in 1939; as Damalinia (Werneckiella) ocellata by Hopkins in 1949. According to Moreby (9), this species was described as *Werneckiella equi asini* by Eichler in 1953.

There is no data that *B. ocellatus* were seen in donkeys, in Turkey. Morphological information will be given about the female of *B. ocellatus* detected on a donkey in Hatay province of Turkey in this paper.

The material of this article consisted of B. ocellatus samples collected from a donkey, which brought to the Ownerless Animal Care and Rehabilitation Center of the Hatay Metropolitan Municipality due to neglecting and senility. Hair losses, ruffling and opacity were observed on the donkey. A few louse specimens were detected on the hair shafts on the back of the donkey during the clinical examination. The louse samples were collected with a forceps and they preserved in a tube with alcohol 70%. After that they sent to Department of Parasitology, Faculty of Veterinary Medicine, Selçuk University. The samples  $(2 \bigcirc 3 \text{ nymphs})$  were cleared in 10% KOH for 24 hours, rinsed in distilled water and passed through series of ethyl alcohol, mounted on the slides in Canada Balsam. After drying, they were examined and identified as *B. ocellatus*. No male of B. ocellatus was detected.

The head of female Bovicola ocellatus slightly rounded in front and it is flat or slightly concave in median. Marginal carina separated from each other by a narrow notch in the anteromedian. The mandibula well developed and notched in the apical. The antenna with three segments; the segment III is slightly longer than the others. The pre-antennal region is slightly shorter and narrower than the post-antennal region. The thorax is narrower and shorter than the head (Figure 1c). It is trapezoidal-shaped, narrowed to posterior. All the legs are weak and with a single claw. The first pair of legs shorter than the others pair and bent in the front. The abdomen is quite larger and longer than the head and thorax. The tergal and sternal plates are well sclerotized in the median, and with a short setae row posteriorly. Tergal plate of the second abdominal segment is thinner than the other tergal plates. The other tergal plates thickened from anterior to posterior. The paratergal plates well developed and became smaller towards to the posterior. The head length of female Bovicola ocellatus (Figure 1b) is 0.5 mm, head width 0.52 mm, thorax length 0.28 mm, thorax width 0.35 mm, abdominal length 1.32 mm, abdominal width 0.87 mm and total length was 2.07 mm.



Figure 1. Bovicola ocellatus and Bovicola equi; a. Bovicola ocellatus, nymph III. b. Bovicola ocellatus, female. c. Bovicola ocellatus head, thorax, original. d. Bovicola ocellatus, female, lower part of tergite IX is pointed (arrow). e. genitalia apertura (arrow). f. Bovicola equi, female. g. head of B. equi h. post-vulval sclerite of B. equi

There were three nymphs; two nymphs III (Figure 1a) and one nymph II. The nymph II is 1.14 mm; the nymph III is 1.5-1.6 mm. The nymphs are very similar to the adult, but tergal, sternal and paratergal plates of nymph II are not developed. The head of nymph III is concave in anteromedian, paratergal plates are weak developed whereas sternal and tergal plates not developed yet.

Moreby (9), stated that five specimens of B. ocellatus from zebra (E. burchellii burchellii), three specimens from horses (E. caballus) and 41 specimens from donkeys (E. asinus) have been collected until that day. Only females of B. ocellatus detected, but male could not find. Firstly, Werneck (13) described male of B. ocellatus and gave its morphological characteristics. Later, he reported that he misidentified this species and corrected this species as *B*. aspilopygus (B. aspilopyga) (14). Although three males of B. ocellatus among the hundreds louse specimens taken from donkeys were reported, but there was no photo or explanation about the male of this species in that paper (3). It is known that B. equi (Denny, 1842), B. bovis (Linnaeus, 1758) and some other lice species reproduce parthenogenetically and produce male only during the rapid growth of the population (15). Probably for this reason, male of *B. ocellatus* could not found in this case.

Bovicola ocellatus can be separable easily from B. equi by its distinctive features. In female of Bovicola ocellatus; the head is widen, and flattened in anterior, the antennal cavity highly sclerotized, pleural plates are large, distinctive and well sclerotized, the post-vulval sclerites widen. In B. equi; the head is narrowed and rounded in anterior (Figure 1g), pleural plates are narrowed, moderately distinctive and weakly sclerotized (Figure 1f), post-vulval sclerites are different in shape (Figure 1h). Although tergite IX. is similar in both species, however in B.equi, it is oval whereas in B. ocellatus it is pointed posteriorly (Figure 1d). In B. equi, the genital opening relatively resembles to hourglass while bell-shaped in *B*. ocellatus (Figure 1e). B. ocellatus can be distinguished from B. aspilopygus and B. zuluensis by having vulvalpost scleritis, and from B. neglectus due to the narrower anterior marginal carina of the head, from B. zebrae due to with the absence of transparent circular areas in the tergite IX. and the wider paratergal plates (9).

Moreby (9) reported that; the mean of the head length of the females of *B. ocellatus* samples collected from zebras was 0.51 mm, the abdominal width was 0.88 mm, while the mean head length of the *B. ocellatus* samples taken from the donkey was 0.46 mm, the width of the abdomen was 0.79 mm, and the average total length of samples collected from both host species was measured as 2.12 mm. In this study, the mean head length of two females of *B. ocellatus* collected from the donkey was 0.5 mm, abdomen width 0.87 mm, total length 2.07 mm. These measurements are consistent with those reported by Moreby.

According to Seguy (10), Harrison considered that B. ocellatus was a different species from B. equi whereas Keler stated that this species was the synonym of B. equi. In Turkey, we did not find any paper about the presence, prevalence and the other features of B. ocellatus as well as B.equi. Although, some authors (7,8) reported that the Bovicola specimens found in horses and donkeys were considered as B. equi. Unfortunately, they (7,8) did not publish any paper or figures of these specimens. In the light of the above information, some other authors (1,6)have also stated that the species seen on horses and donkeys was B. equi. Therefore, it is unclear that the Bovicola samples collected from donkeys are B. equi or B. ocellatus. These findings and considerations can be mistaken, at least partly, and it is not possible to verify all this information today. On the other hand, only one article was found about B. equi was seen in horses in Turkey (5). However, there are no morphological characteristics and figures of B. equi in that paper.

In conclusion, this is the first report of *B. ocellatus* was seen on a donkey in Turkey. In addition, some morphological characteristics and figures of *B. ocellatus* and *B. equi* were explained here. Further studies should be needed to detect the male of *B. ocellatus*, as well as prevalence and biological characters of this species.

## **Financial Support**

This research received no grant from any funding agency/sector.

# **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### References

- 1. Dik B (2015): Veteriner Entomoloji, Ders Kitabı, Selçuk Üniversitesi Basımevi, Konya.
- 2. Durden LA, Mullen GR (2019): *Lice (Phthiraptera).* 79-106. In: Durden LA (Ed), Medical and Veterinary Entomology. 3rd ed. Academic Press, USA.
- 3. Ellse LS (2013): The epidemiology and control of the louse Bovicola ocellatus (Piaget) in donkeys. PhD Thesis, Bristol University, Bristol.
- Emerson KC, Price RD (1981): Host-parasite list of the Mallophaga on mammals. Miscellaneous Publications of the Entomological Society of America.
- Güleğen E, Çırak VY (2005): Bir at çiftliğinde Bovicola (Damalinia) equi (Linnaeus, 1758) enfestasyonu ve tedavisi. Turkiye Parazitol Derg, 29, 183-184.
- İnci A, Yıldırım A, Dik B, et al. (2010): Current knowledge of Turkey's louse fauna. Turkiye Parazitol Derg, 34, 212-220.

- Merdivenci A (1965): Türkiye'nin entomolojik coğrafyası. 114-154. In: Unat EK, Yaşarol Ş, Merdivenci A (Eds), Türkiye'nin Parazitolojik Coğrafyası, Ege Üniversitesi Tıp Fakültesi Yayınları No: 42, Ege Üniversitesi Basımevi, İzmir.
- 8. Mimioğlu M (1973): Veteriner ve Tıbbi Artropodoloji. 343. Ankara Üniv. Vet. Fak. Yayınları, Ankara Üniversitesi Matbaası.
- Moreby C (1978): The biting louse genus Werneckiella (Phthiraptera: Trichodectidae) ectoparasitic on the horse family Equidae (Mammalia: Perissodactyla). J Nat Hist, 12, 395-412.
- **10.** Séguy E (1944): *Insectes ectoparasites:(mallophages, anoploures, siphonaptères)*. 393. 43 Faune de France, Paul Lechevalier et fils, Paris.

- **11. Taylor MA, Coop RL, Wall RL** (2016): *Veterinary Entomology*. 209-210. In: Veterinary Parasitology. 4th ed. Wiley Blackwell, India.
- Vas Z, Rékási J, Rózsa L (2012): A checklist of lice of Hungary (Insecta: Phthiraptera). Annls hist nat Mus natn hung. 104, 5-109.
- **13. Werneck FL** (1950): Os malófagos de mamíferos. part. 2, Ischnocera (continuação de Trichodectidae) e Rhyncophthirina. Edição do Instituto Oswaldo Cruz.
- 14. Werneck FL (1956): A respeito de algunas malafagos de mamiferos. Revta bras. Biol, 16, 25-32.
- **15.** Westrom DR, Nelson BC, Connolly GE (1976): Transfer of Bovicola tibialis (Piaget)(Mallophaga: Trichodectidae) from the introduced fallow deer to the Columbian blacktailed deer in California. J Med Entomol, **13**, 169-173.

Ankara Üniv Vet Fak Derg, **67**, 209-214, 2020 DOI: 10.33988/auvfd.695432

# **Review** / Derleme

# The role of micronutrients in high-yielding dairy ruminants: Choline and vitamin E

# Luciano PINOTTI<sup>1,a,⊠</sup>, Michele MANONI<sup>1,b</sup>, Francesca FUMAGALLI<sup>1,c</sup>, Nicoletta ROVERE<sup>1,d</sup>, Marco TRETOLA<sup>2,e</sup>, Antonella BALDI<sup>1,f</sup>

<sup>1</sup>Milan University, Department of Health, Animal Science and Food Safety, Milan, Italy; <sup>2</sup>Agroscope, Institute for Livestock Sciences, Posieux, Switzerland.

<sup>a</sup>ORCID: 0000-0003-0337-9426; <sup>b</sup>ORCID: 0000-0002-9785-4031; <sup>c</sup>ORCID: 0000-0003-4384-0125; <sup>d</sup>ORCID: 0000-0003-1491-0737; <sup>e</sup>ORCID: 0000-0003-3317-4384; <sup>f</sup>ORCID: 0000-0002-5543-2455.

<sup>⊠</sup>Corresponding author: Luciano.pinotti@unimi.it Received date: 05.02.2020- Accepted date: 27.02.2020

**Abstract:** This review addresses the potential role of antioxidants and methyl-group sources in optimising the metabolic health of dairy ruminants. The productivity of high-yielding dairy cows has increased over the past 40 years and the milk yield has doubled. Such increases in milk production have been observed not only in dairy cows but also to some extent in other dairy ruminants such as ewes, goats and buffaloes (*Bubalus bubalis*). As a consequence, in all specialized dairy ruminants it is essential to optimize the macro and micro-nutrient supply, especially during the most critical period in the animals' production cycle i.e. from parturition until the peak of lactation. In this critical phase, an array of factors can enhance the balance between the intake and demand for nutrients, although the availability and supply of the selected micronutrients is also important. The supplementation of dietary antioxidants or boosting the endogenous methyl group status, via vitamin E, selenium and choline are proposed as possible strategies in maintaining stable metabolic health and optimising milk production.

Keywords: Choline, dairy cows, health, milk production, vitamin E.

# Introduction

In high-yielding dairy ruminants the antioxidant and methyl group status are important in defining the metabolic health of the animals. However, the supplementation of both methyl group sources and antioxidants are usually addressed separately. This review highlights the key knowledge on vitamin E and choline supplementation in dairy ruminants and their role in optimizing milk production and metabolic health.

## Vitamin E

Historically, deficiencies in vitamin E or selenium have been associated with high somatic cell counts (SCCs) in bulk and individual cow milk, and also with increased incidence and severity of intramammary infection (IMI) and mastitis. The positive effect of vitamin E on the SCC depends on adequate dietary Se levels, as reported since the 1980s. When dietary selenium was adequate, vitamin E supplementation significantly reduced the incidence of IMI and clinical mastitis (38). The supplemental vitamin E also improves the killing ability of neutrophils and enhances macrophage function in cows (6, 8, 16, 17, 33). The current requirement for supplemental vitamin E is approximately 500 and 1000 IU/day for lactating and dry cows, respectively. Although NRC requirement published in the 2001 was substantially higher than that of the 1989, other data suggest that higher supplementation rates may be warranted in some situations. Cows supplemented with 4000 IU of vitamin E per day (in combination with 0.1 ppm of supplemental selenium) during the last two weeks before calving and 2000 IU/day during the first week of lactation were found to have significantly fewer mammary gland infections and clinical mastitis compared with 1000 and 500 IU/day during the dry and early lactation periods (42). Subsequent studies (3, 7, 23) confirmed these findings.

A meta-analysis (23) encompassing around 34 published papers between 1984 and 2003 on the relationship between vitamin E and mammary gland health, established that different supplementation levels, up to a maximum of 4000 IU during the dry and early lactation periods, were associated with lower somatic cell counts (SCCs), IMI, and clinical mastitis. Overall, vitamin E supplementation reduced milk SCC by a factor of 0.70,

the risk of IMI by 14%, and the occurrence of clinical mastitis by 30%.

A further disorder in which antioxidant status plays an important role is the retention of fetal membranes (RFM). RFM was one of the first conditions to be attributed to depleted antioxidant defenses and may also be related to impaired neutrophil function. RFM in dairy cows is a cause of endometritis, subsequent ovarian cycle delay and hence delayed pregnancy, resulting in serious economic losses (19). In fact, there is considerable evidence that oxidative stress is enhanced in RFM compared to animals with non-retained placenta. Thus, cows with low fast-acting antioxidants (including alphatocopherol) in plasma, and low glutathione peroxidase in red blood cells have been shown to have a higher incidence of retained placenta than cows with higher antioxidant levels (10, 11). LeBlanc et al. (21) showed that vitamin E (in association with selenium) was involved in RFM, but also highlighted that there are other important contributors, one of which may be the energy supply for immune function.

Supplementing dairy cattle with adequate amounts of vitamin E and Se is thus now a widely-accepted practice. However, adequate levels of these micronutrients do not completely control oxidative stress around calving (3, 6, 27). Numerous studies in dairy cows (34) have shown that

the plasma vitamin E concentration decreases gradually around parturition, reaching the lowest values around calving and then increasing gradually afterwards. The inadequacy of micronutrient supplementation in this situation is probably due to increased milk yields, and a greatly reduced feed intake, resulting in a substantial increase in the recommended intake levels for this animal (24, 43).

In dairy cows the intake of vitamin E is generally considered adequate when alpha-tocopherol plasma levels are over 3-3.5 mg/ml or above 2 mg/mg relative to plasma cholesterol. Beyond these levels, no further benefits are observed (3). Reaching an adequate vitamin E status however, is difficult in dairy cows. Plasma vitamin E levels, expressed as alpha-tocopherol, fall significantly around parturition in all dairy ruminants (cows, ewes, goats, buffaloes - Figure 1), and it is difficult to maintain levels that are thought to be adequate for health. It has been suggested that this is one of the reasons for the decline in peripartum innate and acquired immune defense in dairy ruminants. Supplementation levels should therefore be re-considered, choosing high bioavailability sources. In this respect, the bio-potency and bioactivity of naturally occurring RRR-a-tocopherol are known to be higher than its synthetic all-rac counterpart (34, 41).



**Figure 1.** Changes in plasma alpha-tocopherol around parturition (day 0 in *x* axis) in various dairy ruminant species. Data from Hogan et al. (17), Panda et al. (25), Pinotti et al. (32), Toker (40). x-axes = days relative to parturition; y- axes = alpha-tocopherol, mg/l.

Advances in feed manufacturing technology (including microencapsulation, and polymer coat protection) now make it possible to design products that deliver specific nutrients to the absorption sites of the small intestine. It is also clear that the formulation (natural vs. synthetic; inorganic vs. organic form; protected vs. unprotected) may not only influence intestinal bioavailability, but may also affect the uptake from target organs, tissue utilization, transfer to the new-born animals (1), and food quality (e.g. milk).

# Choline

Choline supplementation increases productivity as well as metabolic health in dairy ruminants (2, 29, 32). Choline is often considered as a vitamin however, unlike classical vitamins, its endogenous synthesis is possible, and a choline deficiency syndrome usually goes undetected in healthy mammals. It has therefore been suggested that choline may be an essential nutrient for mammals, irrespectively of whether or not it is classified as a vitamin-like compound. Choline has an important role in energy and protein metabolism especially for those pathways that take place in the liver. It is involved in lipid metabolism, and more precisely in lipid transport as a lipotropic agent.

Choline, an important labile methyl group donor, takes part in the biosynthesis of other methylated compounds in the body. This latter function is "code shared" with methionine, thus choline and methionine are interchangeable being the two primary methyl donors in animal metabolism.

However, the metabolic pathways of choline and methyl groups are different in ruminants. In adult ruminants, choline is extensively degraded in the rumen; and as a consequence, unable to contribute significantly to the choline body pool. The methyl group metabolism is generally conservative with a relatively low rate of methyl catabolism, and a high rate of de novo synthesis of methyl groups via the tetrahydrofolate (THF) system.

In ruminants there is a conservative methyl group metabolism, which works perfectly in positive energy balance animals (e.g. beef cattle). However, this can become exacerbated in lactating dairy ruminants, for which the availability of dietary choline is still limited, while the output of methylated compounds in milk is high, and precursors from the tetrahydrofolate pathway (such as glucose and other gluconeogenetic precursors) are limiting, especially at the beginning of lactation.

On the basis of these assumptions, the hypothesis that choline may be a limiting nutrient for milk production has been proposed (29, 32). A great number of studies conducted to assess the possible effects of choline in milk production, established that from a technical/feeding point of view, choline must be rumen-protected in order to be effective in ruminant nutrition.

Knowledge about transition and early lactation periods of dairy cows proposes that greater choline availability improves not only milk production (5, 9), but also lipid (12, 13, 26, 30, 32) and methyl group metabolism (5). These results have also been investigated in a few meta-analyses. In 2010, Sales et al. (37) quantified the dietary effects of RPC (rumen-protected choline) on the production traits of dairy cows. In Sales' review, by increasing RPC supplementation from 6 to 50 g/d, which can also be effective in increasing milk yield, no dose response was observed. Milk yield in fact, decreased from 131.5 to 0.037 g of milk for each g of dietary RPC supplemented in the diets from 6 up to 50 g/d. In the same supplementation range, milk fat content decreased linearly at a rate of 0.00339% for a 1g/d increase in dietary RPC, confirming that choline has no effect on the milk fat content. Although the reasons for this are not clear, an interaction between choline and methionine has been proposed (37).

More recently two other meta-analyses on choline in dairy cow nutrition have been published. In 2019 using data from 27 studies, Humer et al. (18) reported that choline supplementation increased postpartum dry matter intake by 4% and milk yield by 3.4% in dairy cows. A higher milk yield also induced a higher milk fat yield and milk protein yield, without affecting their concentration. However also in this meta-analysis, no dose/response effects were observed. Providing RPC had no effect not only on the main plasma metabolites (such as nonesterified fatty acids, beta-hydroxybutyrate, glucose, and cholesterol) but nor on theincidence of ketosis, and mastitis across all studies in this meta-analysis.

These results seem to be in contrast with several studies in which choline was shown to improve the metabolic health and liver function (5, 12, 13, 26, 30). In fact, Zenobi et al. (44) stated that feeding pregnant, non-lactating Holstein cows with increasing amounts of RPC (from 0 to 25.8 g/d) decreased the concentration of hepatic triglycerides in a linear manner. Dietary choline thus seems to improve fat metabolism in the liver and increase the apparent absorption/traffic of triglycerides in the bloodstream, further confirming the beneficial effects of supplemental RPC for adult ruminants.

The most recent meta-analysis conducted by Arshad et al. (2) was based on 21 experiments including a total of 1313 cows, with a range of prepartum choline supplementation from 5.0 to 25.0 g/d. The results confirm that milk yields and energy-corrected milk (ECM) in multiparous cows increase linearly with the amount of dietary choline supplemented during the transition period. In addition, across the studies included in this metaanalysis, feeding RPC tended to reduce retained placenta and mastitis, although the optimum dose of choline was not established. Arshad et al. (2) recommended more than



Figure 2. Meta-regression of choline amount (g/d of choline chloride) on effect size (MY) based on 17 studies.

13 g/d of choline, provided in a rumen-protected form. This quantity, however should be considered with caution since the dose–response relationship between dietary choline and milk yield needs to be addressed more thoroughly (Figure 2). Thus, several physiological and dietary factors are probably related to the obtained responses with dietary RPC supplementation necessitating further research to investigate the precise mechanism of choline action in lactating dairy cows.

In the case of other dairy ruminants, the effects of RPC have been less investigated. In dairy buffaloes 15 g/d of choline in a rumen-protected form were shown to increase milk yield by 15% (15). Using the same RPC dose, Kumar et al. (20) obtained a 20% increase in milk yield in choline-supplemented animals compared to the controls. The effect of RPC on milk yield recorded in these studies on dairy buffaloes are among the highest registered in dairy ruminants. Although the exact mechanism for such a large response is still unknown, the co-presence of supplemental fat in the diets cannot be ruled out. In small ruminants, Lobley et al. (22) studied the effects of choline infusion in sheep on transmethylation reactions, while Emmanuel and Kennelly (14) investigated methionine and choline incorporation into the plasma and milk of lactating goats. In both cases however, the main goal was to assess the fate and metabolism of methyl groups and the interchangeable nature between choline and methionine. Other researchers (4, 32) also assessed the efficacy of RPC supplementation in dairy goats. Pinotti et al. (32) supplemented periparturient dairy goats with 4 g/d of RPC

starting four weeks prior to expect kidding and continuing for five weeks after parturition. Supplemental choline dose was derived from the experiments with dairy cows (32) corrected for the metabolic body weight (BW<sup>0.75</sup>) of the goats at the beginning of the experiment. In RPC supplemented goats during early lactation, milk yield and fat-corrected milk (4% FCM) yield increased by 7 and 12%, respectively, compared to non-supplemented goats. RPC supplement also increased the milk fat concentration, and fat and protein yield while plasma metabolites did not differ between treatments. By contrast, in dairy ewes (39), RPC supplementation in combination with methionine and betaine had no effect on milk yield, although the dose of choline used was extremely low. It is worth noting that the number of studies on ovine species is still limited.

Overall these results suggest that greater choline availability is essential for optimizing milk production in dairy ruminants, although other factors such as dietary composition, dietary crude protein content, and postruminal methionine supply merit further investigation.

This review has highlighted that the dietary supply of antioxidants and methyl group sources may not always be sufficient to maximize milk production in dairy ruminants. Although the requirement of these nutrients may in theory be satisfied by specific supplementation (vitamin E and selenium) or by other nutrients (in the case of choline), it is unlikely that this happens in practice especially at the onset of lactation. The magnitude of the production response is likely to be affected by the basal diet composition, the dose of the nutrient, its form of presentation/supply, and the stage of lactation (5, 24, 28, 41). However, it is also evident that our knowledge is incomplete regarding the potential role of antioxidants and methyl group sources in optimising metabolic health in dairy ruminants. For example, there are indications that both methyl group precursors (including choline) and other co-factors (folate and vitamin B12) are important for the optimal metabolic support of milk production, although methionine may not always be involved in this scenario (35). At the same time there are limited indications (in vitro, 36) that antioxidant and methyl sources interact positively in bovine mammary epithelial cells. In conclusion, a nutritional approach based on vitamin E and choline supplementation in dairy ruminants could be beneficial in terms of optimizing their milk production and metabolic health. However, the supply and use of individual nutrients should be reconsidered in order to meet the novel findings about the synergic effects of these compounds in ruminant diet.

#### **Financial Support**

This review received no grant from any funding agency or sector.

# **Conflict of Interest**

The authors declared that there is no conflict of interest.

#### References

- 1. Abuelo A, Hernández J, Benedito JL, et al (2019): *Redox* biology in transition periods of dairy cattle: Role in the health of periparturient and neonatal animals. Antioxidants, **8**, 20.
- 2. Arshad U, Zenobi MG, Staples CR, et al (2020): Metaanalysis of the effects of supplemental rumen-protected choline during the transition period on performance and health of parous dairy cow. J Dairy Sci, 103, 282-300.
- **3.** Baldi A (2005): *Vitamin E in dairycows*. Livest Prod Sci, **98**, 117-122.
- 4. Baldi A, Bruckmaier R, D'ambrosio F, et al (2011): Rumen-protected choline supplementation in periparturient dairy goats: effects on liver and mammary gland. J Agric Sci, 150, 1-7.
- 5. Baldi A, Pinotti L (2006): Choline metabolism in highproducing dairy cows; metabolic and nutritional basis. Can J Anim Sci, 86, 207-212.
- Baldi A, Pinotti L (2015): Antioxidant nutrition in dairy ruminants. In Book of Abstracts of the 66<sup>th</sup> Annual Meeting of the European Association for Animal Production, Wageningen Academic Publishers, The Netherlands.
- Baldi A, Savoini G, Pinotti L, et al (2000): Effect of vitamin E and different energy sources on vitamin E status, milk quality and reproduction in transition cows. J Vet Med A, 47, 599-608.
- Baldi A, Pinotti L, Giromini C, et al (2018): Nutritional strategies to counteract oxidative stress: benefits and challenges.In Book of abstracts of the 69<sup>th</sup> Annual Meeting

of The European Association for Animal Production. Wageningen Academic Publishers, The Netherlands.

- **9.** Brüsemeister F, Sudekum KH (2006): Rumen-protected choline for dairy cows: the in situ evaluation of a commercial source and literature evaluation of effects on performance and interactions between methionine and choline metabolism. Anim Res, **55**, 93-104.
- Brzezinska-Slebodzinska E, Miller JK, Quigley JD, et al (1994): Antioxidant status of dairy cows supplemented prepartum with vitamin E and selenium. J Dairy Sci, 77, 3087-3905.
- Campbell MH, Miller JK (1998): Effect of supplemental dietary vitamin E and zinc on reproductive performance of dairy cows and heifers fed excess iron. J Dairy Sci, 81, 2693-2699.
- **12.** Chung YH, Brown NE, Martinez CM, et al (2009): Effects of rumen-protected choline and dry propylene glycol on feed intake and blood parameters for Holstein dairy cows in early lactation. J Dairy Sci, **92**, 2729–2736.
- Cooke RF, Silva Del Río N, Caraviello DZ, et al (2007): Supplemental Choline for Prevention and Alleviation of Fatty Liver in Dairy Cattle. J Dairy Sci, 90, 2413-2418.
- Emmanuel B, Kennelly JJ (1984). Kinetics of methionine and choline and their incorporation into plasma lipids and milk components in lactating goats. J Dairy Sci, 67, 1912-1918.
- **15.** Garg MR, Sherasia PL, Bhanderi BM (2012): Effect of supplementing bypass fat with and without rumen protected choline chloride on milk yield and serum lipid profile in Jaffarabadi buffaloes. Buffalo Bull, **31**, 91-97
- Hogan JS, Weiss WP, Todhunter DA, et al (1992): Bovine neutrophil responses to parenteral vitamin E. J Dairy Sci, 75, 340- 399.
- Hogan JS, Weiss WP, Smith KL (1993): Role of vitamin E and selenium in host defence against mastitis. J Dairy Sci, 76, 2795-2908.
- **18.** Humer E, Bruggeman G, Zebeli Q (2019): A Meta-Analysis on the Impact of the Supplementation of Rumen-Protected Choline on the Metabolic Health and Performance of Dairy Cattle. Animals **9**, 566.
- **19. Kankofer M, Lipko J, Zdunczyk S** (2005): *Total antioxidant capacity of bovine spontaneously released and retained placenta*. Pathophysiology, **11**, 215-219.
- 20. Kumar R, Nayak S, Baghel RPS, et al (2017): Effect of Prill Fat and Rumen Protected Choline Supplementation on Feed Intake, Body Weight Changes and Economics of Lactating Murrah Buffaloes. J Anim Res, 7, 355-359.
- LeBlanc SJ, Herdt TH, Seymour WM, et al (2004): Peripartum serum vitamin e, retinol, and beta-carotene in dairy cattle and their associations with disease. J Dairy Sci, 87, 609-619.
- 22. Lobley GE, Connell A, Revell D (1996): The importance of transmethylation reactions to methionine metabolism in sheep: effect of supplementation with creatine and choline. Brit J Nutr, 75, 1, 47-56.
- 23. Moyo N, Nielen M, Kruitwagen C, et al (2005): Vitamin E Supplementation and Udder Health: A Meta-Analysis. 159-165.In: H. Hogeveen (Ed), Mastitis in Dairy Production. Current Knowledge and Future Solutions, Wageningen Academic Publishers, Wageningen, The Netherlands.

- **24.** NRC (2001): Nutrient requirements in dairy cattle. Natl. Acad. Press 7th ed., Washington DC.
- 25. Panda N, Kaur H, Mohanty TK (2005): Reproductive Performance of Dairy Buffaloes Supplemented with Varying Levels of Vitamin E. Asian-Aust. J Anim Sci, 19, 19-25.
- Piepenbrink MS, Overton TR (2003): Liver metabolism and production of cows fed increasing amounts of rumenprotected choline during the periparturient period. J Dairy Sci, 86, 1722-1733.
- 27. Pilotto A, Savoini G, Baldi A, et al (2016): Associations between blood fatty acids, β-hydroxybutyrate, and αtocopherol in the periparturient period in dairy cows: An observational study. J Dairy Sci, 99, 8121-8126.
- 28. Pinotti L (2012): Vitamin-Like Supplementation in Dairy Ruminants: The Case of Choline. 65-86. In: Narongsak Chaiyabutr (Ed), Milk Production - An Up-to-Date Overview of Animal Nutrition, Management and Health, InTech, Narongsak Chaiyabutr, Croatia.
- **29.** Pinotti L, Baldi A, Dell'Orto V (2002): Comparative mammalian choline metabolism with emphasis on the high-yielding dairy cow. Nutr. Res. Rev, 15: 315–331.
- **30.** Pinotti L, Baldi A, Politis I, et al (2003): Rumen protected choline administration to transition cows: effects on milk production and Vitamin E status. J Vet Med A, **50**, 18-21.
- **31.** Pinotti L, Campagnoli A, Dell'Orto V, et al (2005): *Choline: Is there a need in lactating dairy cow*, Livest Prod Sci, **98**, 149-152.
- **32.** Pinotti L, Campagnoli A, D'Ambrosio F, et al (2008): Rumen-Protected Choline and Vitamin E Supplementation in Periparturient Dairy Goats: Effects on Milk Production and Folate, Vitamin B12 and Vitamin E Status. Animal, 2, 1019-1027.
- **33.** Politis I, Hidiroglou N, Batra TR, et al (1995): Effects of Vitamin E on Immune Function of Dairy Cows. Am J Vet Res, **56**, 179-184.
- **34.** Politis I (2012): Reevaluation of vitamin E supplementation of dairy cows: Bioavailability, animal health and milk quality. Animal, 6, 1427–1434

- 35. Preynat A, Lapierre H, Thivierge MC, et al (2009). Effects of supplements of folic acid, vitamin B12, and rumen-protected methionine on whole body metabolism of methionine and glucose in lactating dairy cows. J Dairy Sci, 92, 677-689.
- **36.** Rebucci R, Pinotti L, Fusi E, et al (2013). Role of Choline and Methionine in Bovine Mammary Epithelial Cell Line Exposed to Hydrogen Peroxide. J Nutr Ecol Food Res, 1, 189-193.
- **37.** Sales J, Homolka P, Koukolová V (2010): Effect of dietary rumen-protected choline on milk production of dairy cows: A meta-analysis. J Dairy Sci, 93, 3746-3754.
- Smith KL, Harrison JH, Hancock DD, et al (1984): Effect of Vitamin E and Selenium Supplementation on Incidence of Clinical Mastitis and Duration of Clinical Symptoms. J Dairy Sci, 67, 1293-1300.
- **39.** Tsiplakou E, Mavrommatis A, Kalogeropoulos T, et al (2017): The effect of dietary supplementation with rumenprotected methionine alone or in combination with rumenprotected choline and betaine on sheep milk and antioxidant capacity. J Anim Physiol Anim Nutr, **101**, 1004-1013.
- 40. Toker NY (2007): Blood serum Vitamin A and E concentrationsand distribution into lipoprotein fractionsof pregnant sheep and newborn lambs. Revue Méd Vét, 158, 413-417.
- Vagni S, Saccone F, Pinotti L, et al (2011): Vitamin E Bioavailability: Past and Present Insights. Food Nutr Sci, 2, 1088-1096.
- **42.** Weiss WP, Hogan JS, Todhunter DA, et al (1997): Effect of Vitamin E Supplementation in Diets with a Low Concentration of Selenium on Mammary Gland Health of Dairy Cows. J Dairy Sci, **80**, 1728-1737.
- **43.** Weiss WP (1998): Requirements of Fat-Soluble Vitamins for Dairy Cows: A Review. J Dairy Sci, **81**, 2493-2501.
- 44. Zenobi MG, Scheffler TL, Zuniga JE, et al (2018): Feeding increasing amounts of ruminally protected choline decreased fatty liver in nonlactating, pregnant Holstein cows in negative energy status. J Dairy Sci, 101, 5902– 5923.