

The therapeutic effectiveness of thyme extract in naturally infected puppies with ascariasis

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ABSTRACT

Objective: This study aimed to investigate the therapeutic efficacy of thyme extract in puppies naturally infected with ascarids.

Material-Method: The study consisted of 20 puppies of different sexes, 2-4 months old, naturally infected with ascarid. There were given an oral 20% concentration of thyme extract for 3 days to puppies, and faecal egg counts were conducted on the 0th, 1st, 2nd, 3rd, and 7th days after the treatment was started (day 0). Also, serum urea, creatinine, AST, ALT levels were monitored on the 0th and 3rd days, together with daily clinical examination, to monitor possible toxic effects.

Result: In 2 puppies (10%), the fecal egg count was highly variable at post-treatment examinations, but no reduction in egg count was observed. Egg shedding in 7 (35%) of the treated puppies was zero. It was observed that egg shedding was not completely zero in 11 of the puppies (55%). However, the egg counts decreased by 25% to 98.3%. It was also observed that the values of the measured blood biochemical values were within reference range except serum urea levels and the puppies did not show any clinical sign of toxicity during the treatment.

Conclusion: It was concluded that the thyme extract did not have any toxic effect in the puppies at the concentration studied, and it could be effective in the treatment of ascariasis.

Keywords: Toxocara spp., Puppy, Ascariasis, Treatment, Thyme extract

INTRODUCTION

Ascarids are among the most common gastrointestinal parasites encountered in dogs (Overgaauw and Van Knapen, 2013; Becskei et al., 2020a). Globally, *Toxocara canis* and *Toxascaris leonina* in particular, are the most common canine ascarids (Becskei et al., 2020b). These parasites, which settle in the small intestines of their main hosts, carnivores can also infest paratenic hosts

such as; humans, mice, earthworms, ticks, chickens, sheep, pigs and birds (Glickman and Schantz, 1981). Ascarids are important zoonoses because they can be easily transmitted to humans from cats and dogs with whom we are in constant contact in our daily lives (Despommier, 2003).

The development and course of the infection in dogs; the age, sex of the animal, the number of infected eggs exposed, hormonal status, immune system and the migration route of the larvae

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directly affect it (Lloyd et al., 1998). It can cause growth retardation, diarrhoea, dehydration, abdominal bloating, intestinal obstruction, ascarid toxicity and death, especially in young dogs (Burrows et al., 1995).

In the fight against ascariasis in dogs, it is recommended to first deworm puppies from parasites, and then apply ascarid treatment to adult dogs four times a year, even if there is no effective and regular examination (Becskei et al., 2020b). Due to its zoonotic importance, it has been reported that dogs that come into contact with individuals with compromised immune systems should be treated monthly (Companion Animal Parasite Council, 2020). In the treatment of ascariasis in dogs, medications such as; pyrantel pamoate, ivermectin, selamectin, eprinomectin, moxidectin, nitroscanate, mebendazole, milbemycin, sarolaner, spinosad, lotilaner, praziquantel are used either alone or in combinations (Genchi et al., 1990; Clark et al., 1991; Bowman et al., 1998; McTier et al., 2000; Kozan et al., 2008; Cardenas et al., 2017; Young et al., 2021). However, there is no 100% effective treatment protocol for T. canis in dogs, since its biology is very complex and there are still aspects of it that cannot be clarified (Doğanay et al. 2018).

In its simplest form, phytotherapy can be defined as the treatment made with plants. Phytotherapy, together with chemical treatments or as an alternative treatment, has attracted the attention of researchers in recent years (Sarışen and Çalışkan, 2005). When the phyto-therapeutic potentials of thyme were examined; it was reported to have antibacterial, antiviral, antiparasitic, antifungal and antioxidant potentials (Burt, 2004). The essential oils of thyme contain compounds such as; carvacrol, borneon, tanen, pimen, cymol, and flavones in addition to their intense thymol quantity. Their thymol ratio is around 50% (Benli and Yiğit, 2005).

In this study, it was aimed to investigate the treatment efficacy of thyme extract as an alternative to chemical treatments in naturally infested puppies with ascarid.

MATERIALS and METHODS

The study was carried out with the permission of Kırıkkale University Animal Experiments Local Ethics Committee, dated 19/12/2019 and numbered 63. The study consisted of 20 puppies of different sexes, 2-4 months old, which were brought to Kırıkkale University, Veterinary Faculty, Animal Hospital for medical examination and vaccination.

Puppies that were mono-infected with ascaris upon stool examinations were included in the study.

Before the administration of thyme to the animals included in the study, blood and stool samples were taken, it was administered to orally 3 times in total on the 0th, 1st and 2nd days, by diluting the commercial thyme extract (Thy717, BioArt, Turkey) to a concentration of 20%, at a dose of 1 ml/kg.

Stool samples were taken from the puppies for a total of 5 times, on the first 4 days and on the 7th day of the beginning of the treatment, to count eggs in each gram faeces. In order to monitor for possible toxicity; serum urea, creatinine, aspartate (AST) aminotransferase and alanine aminotransferase (ALT) values were measured by taking blood samples twice, before thyme extract administrations (day 0) and after thyme extract administrations (3th day).

To determine the presence of Toxocara spp. eggs in stool samples, the Fülleborn flotation technique was used. The Mc Master technique was adopted to determine the number of eggs in 1 g stool before, during, and after the treatment. For this, the average of 40 glass beads were thrown into 100 ml jars. Three g of faeces was weighed and put into the jar and 42 ml of saturated salt water was added. Stool was homogenized by closing the lid of the jar. The resulting solution was transferred into centrifuge tubes and centrifuged at 1500 rpm for 3 minutes. The upper liquid was removed without moving the sediment at the bottom, and the same amount of saturated salt water was added to the faecal residue at the bottom of the tube, and the centrifuge tube was turned upside down 5-6 times to homogenize the sediment. The chambers of the Mc Master slide were filled with the help of a Pasteur pipette. Eggs were counted at x10 magnification under the light microscope and the number of eggs detected in both chambers was multiplied by 100 to determine the total number of eggs in 1 gram of stool (Senlik, 2006). Faecal egg reduction test (FECRT) was also adopted to determine the efficacy of the thyme extract against T. canis (Doğanay et al., 2018). Egg reduction was calculated according to the formula below.

 $FECRT = \frac{EPG \text{ value before treatment - EPG value after treatment}}{EPG \text{ value before treatment}} X100$

RESULTS

The daily change in *Toxocara* spp. egg counts in the puppies given the thyme extract is presented in Table 1. According to the data, it was determined

that the number of eggs was zeroed after the administration of the thyme extract in 7 out of 11 puppies which had faecal egg counts between 200 and 1650 on day 0, and in 1 out of 9 puppies which had faecal egg counts more than 1650. In total, faecal egg counts zeroed in 7 dogs (35%).

| Table 1. Faecal egg counts results of | the puppies |
|---------------------------------------|-------------|
|---------------------------------------|-------------|

There was no decrease in the number of eggs laid in 2 of the 20 puppies. In 11 puppies, the egg laying rate showed a wide distribution between 25% and 98.3% (Table 1). Considering the general distribution, the percentage of egg reduction was found to be higher in puppies with relatively low egg counts prior to treatment.

| No. | Day 0 | Day 1 | Day 2 | Day 3 | Day 7 | % of Reduction |
|-----|-------|-------------|-------|-------|-------|----------------|
| 1 | 200 | 0 | 0 | 0 | 0 | 100 |
| 2 | 200 | 700 | 400 | 0 | 0 | 100 |
| 3 | 500 | 0 | 0 | 200 | 0 | 100 |
| 4 | 500 | Not Sampled | 50 | 0 | 250 | 50 |
| 5 | 500 | 150 | 0 | 0 | 50 | 90 |
| 6 | 750 | Not Sampled | 0 | 0 | 50 | 93.3 |
| 7 | 1000 | 900 | 1350 | 5200 | 3700 | No Reduction |
| 8 | 1100 | 7450 | 0 | 0 | 0 | 100 |
| 9 | 1500 | 1650 | 1200 | 1250 | 0 | 100 |
| 10 | 1500 | Not Sampled | 0 | 100 | 900 | 40 |
| 11 | 1650 | Not Sampled | 700 | 50 | 0 | 100 |
| 12 | 2100 | 1950 | 4950 | 9800 | 4600 | No Reduction |
| 13 | 2350 | 3900 | 5000 | 6000 | 1150 | 51.1 |
| 14 | 2500 | 4100 | 7100 | 6000 | 1050 | 58 |
| 15 | 3000 | 3000 | 2350 | 500 | 50 | 98.3 |
| 16 | 4250 | 12000 | 11650 | 0 | 0 | 100 |
| 17 | 8400 | 5800 | 1200 | 4000 | 6300 | 25 |
| 18 | 8400 | 5100 | 6250 | 12150 | 6050 | 28 |
| 19 | 15000 | 18200 | 2700 | 6250 | 1450 | 90.3 |
| 20 | 28150 | 5050 | 9600 | 16250 | 17900 | 36 |

| Table 2. N | Mean chang | ges in blood | l biochemical | parameters | before and | after treatment. |
|------------|------------|--------------|---------------|------------|------------|------------------|
| | | 7 | | | | |

| Demonsterne | Before Treatment (n=20) | | | After Treatment (n=20) | | | Deferrer et Velvee | |
|--------------------|-------------------------|------|----------------|------------------------|------|----------------|--------------------|--|
| Parameters | Min | Max | \overline{x} | Min | Max | \overline{x} | Kelerence values | |
| Urea (mg/dl) | 10 | 28 | 15.38 | 13 | 57 | 24.52 | 10-26 | |
| Creatinine (mg/dl) | 0.16 | 0.48 | 0.26 | 0.12 | 0.57 | 0.32 | 0.5-1.3 | |
| AST (U/L) | 15 | 88 | 28.84 | 7 | 58 | 21.15 | 10-88 | |
| ALT (U/L) | 9 | 21 | 15.15 | 3 | 26 | 15.63 | 10-90 | |

An adult worm was detected on the 7th day after the treatment in puppy number two and on the 3rd day in puppy number 10. On the 3rd day after the treatment, a large number of adult parasite excretions were observed in dogs with 17 and 19 numbers. It was observed that the general condition of dog number 15 was deteriorated prior to the treatment, however, the general condition improved after 3 days of treatment.

The mean values obtained from the measurements made from the blood samples taken before and after the treatment are given in Table 2. It was observed that serum urea, creatinine, AST and ALT values in the blood samples taken 24 hours after the last treatment dose were within the normal reference ranges. Although the mean value of urea after treatment approached the upper limit, it was still within the normal reference limit.

All the animals included in the study were clinically completely healthy during the study and on the 7th day examinations. During this period, there were no findings such as diarrhoea, vomiting, depression and anorexia that could indicate clinical toxication.

DISCUSSION

Canine ascariasis can cause fatal diseases, especially in susceptible dogs, and draws attention as it is a zoonotic condition (Macpherson, 2013; Hassanain et al., 2015). Especially in areas where people and dogs are in close contact, such as parks and gardens, the parasite can easily be transmitted to both susceptible dogs and humans by faecal contamination (Fankhauser et al., 2016). For this reason, in the fight against ascariasis in dogs, it is recommended to break the parasitic cycle by spraying dogs at least 4 times a year, even if no parasite is found (Becskei et al., 2020b). Due to widespread resistance to anti-helminthics (chemical drugs) used in the treatment of canine ascariasis, there is the need to constantly seek for new remedies (Coles et al., 1992; Hoekstra et al., 1997; Silvestre and Humbert, 2002). Among the strategies developed against helminths is phytotherapy (Jahangir et al., 2001).

Considering studies investigating the antiparasitic activities of thyme: Hafez et al. (2019) and Amin et al. (2016) T. canis in experimentally infected rats; Amin and El-Kabany (2013), T. vitulorum; Luis et al. (2016) anti-helminthic against Haemonchus contortus in experimentally infected sheep; Malatyalı et al. (2009) antileishmanial in vitro; Behnia et al. (2008) Entamoeba histoytica trophozoites in vitro; Gaur et al. (2018) in cell culture, Kara et al. (2022) antiprotozoan on Cryptosporidium parvum in experimentally infected rats; Attia et al. (2015), intestinal and cystic stages of Trichinella spiralis; Morsy et al. (1998) Lucilia sericata larvae; Remmal et al. (2011), Abbas et al. (2012), Arczewska-Wlosek and Swiatkiewicz (2012) studied the anticoccidial activity of thyme.

Studies have shown that the ant-parasitic activity of thyme extract is mediated by thymol, which constitutes 50% of the active ingredients (Ferreira et al., 2016). It has been reported that the antihelmintic effect of tymol is based on its paralysing of the parasite, similar to the mechanism of action of macrocyclic lactones, and the inhibition of movement and feeding functions (Kotze et al., 2012; Lynagh et al., 2014). It has been reported that when macrocyclic lactones such as ivermectin, moxidectin, and eprinomectin are used for treatment in canine ascariasis, it reduces the number of faecal eggs by 100%, while this rate is 99.7% with topical use of selamectin, a member of the same group (Pal et al., 1995, Gargili et al., 1999; Payne- Johnson et al., 2000; Kozan et al., 2008). Despite its success in treatment, it should not be forgotten that ivermectins are neurotoxic, especially in sensitive breed dogs such as Collies (Pronk and Schefferlie, 1998).

In canine ascariasis, both the toxic/side effects of chemical agents and the development of antiparasitic resistance, and the costs of continuous new drug development have led researchers to seek for treatment options (Pronk and Schefferlie, 2022; Jackson and Miller, 2006). Hassanain et al. (2015) compared the efficacy of mebendazole and sugar lemon (Citrus aurantifolia Swingle) seed extract in dogs naturally infected with Ancylostoma caninum and T. canis. According to the study, the number of faecal eggs was taken as a reference in the evaluation of treatment efficacy, faecal egg numbers; decreased by 74.10% in the group given only mebendazole, by 91.08% in the group given only sugar lemon extract, and by 98.20% in the group given mebendazole and sugar lemon extract. In our study, egg laying decreased by 25-100% in 18 dogs given thyme extract, while egg laying did not decrease in 2 dogs.

Amin et al. (2016) evaluated the therapeutic efficacy of thyme oil in experimentally infected rats with *T*. canis; they took the number of larvae detected in the brain tissue and brain damage as success criteria and reported that the treatment with thyme oil was found to be effective compared to the control groups. Hafez et al. (2019) examined the therapeutic efficacy of thyme oil in rats experimentally infected with T. canis larvae in a similar study. In the evaluation of the efficacy of the treatment, the histological changes observed in the testis tissue due to larval migration were taken as reference; while moderate and reversible changes were observed in the testicular tissue of the group treated with the thyme oil, serious histopathological changes were detected in the control group. In both studies, thyme oil was found to be effective in the treatment of experimental T. canis infection in rats. This result obtained in the experimental animals inspired this clinical study in dogs, the definitive host of T. canis.

In our study, it was determined that the number of eggs in the faecal samples taken 24 hours after the first treatment application increased significantly. This is thought to be due to increase intestinal peristalsis of thyme extract, induced fragmentation of adult parasites and increase in the number of eggs excreted in faeces due to fragmentation of adult female parasites.

Yıldız et al. (2011) in a study in which they examined the efficacy of Artemisia absinthium extract in cats naturally infected with Toxocara cati, they reported that egg laying stopped in only two of the 8 cats given the extract, and in the remaining cats, although the number of eggs decreased compared to the first day, the laying did not stop. In our study, egg laying stopped in 8 dogs (40%) after the administration of the thyme extract against T. canis, which is in the same family as T. cati. Similar to the related study, in 3 dogs egg shedding was reduced by 90.3-98.3%, while there was no decrease in 2 dogs. In 7 dogs, egg laying decreased by 25-58%. When the results of these studies were compared, it was found that the zeroing rate of thyme extract (35%) was higher compared to the zeroing rate of A. absinthium extract against ascarides (25%).

Hassanain et al. (2015) applied *Citrus aurantifolia* (lemon blossom) extract against *T. canis* in Egypt. In that study, they reported that the number of eggs decreased by 91.8% in the group in which only *C. aurantifolia* was administered. In our study, when the egg reduction in the stools examined before and after the treatment was calculated, it was determined that the egg reduction varied individually in the animals and the egg reduction was between 25-100%. In addition, it was recorded in 2 dogs that thyme extract had no effect on the number of eggs excreted in the faeces.

Although some of the cat and dog owners believe that natural products are safer than synthetic chemicals in parasitic control, it should not be forgotten that essential oils and plant extracts also have toxic effects on animals (Villar et al., 1994; Woolf, 1999; Genovese et al., 2012). Genovese et al. (2012), in their retrospective study covering a 2-year period, in which they compiled the cases of poisoning reported as a result of the use of topical products containing essential oil in cats and dogs; they evaluated different products made of oils such as peppermint, cinnamon, lemongrass, clove, thyme, cedarwood, rosemary, wheat germ oil. In the evaluation, clinical findings such as lethargy, weakness, desire to lie down, hyperactivity, tachycardia, hypothermia, hyperthermia, seizures, skin erythema, vomiting, diarrhea, edema, ataxia, agitation, anorexia, fasciculation, hiding, hypersalivation, panting, tremor were observed in animals. They mentioned that kidney failure may occur. Bischoff and Guale (1998) reported that liver enzymes such as ALT and AST increased in cats exposed to tea tree (Melalleuca alternifolia) oil poisoning. In this study, in which ascariasis was

treated with the oral use of thyme extract, no clinical symptoms of poisoning were observed during or after the study. In order to monitor whether the treatment with thyme extract caused organ failure or not, urea and creatinine were measured for kidney failure, and AST and ALT values were measured for liver failure. Although all values before and after treatment were within their normal physiological reference limits, it was noted that the urea value approached the upper reference values after treatment. In subsequent studies, if the treatment period of 3 days is extended, it would be beneficial especially, in monitoring values for kidney functionality closely. Although there are reports of cats and dogs with a history of poisoning after topical applications with products such as thyme oil, no signs of poisoning were observed in any of the 20 dogs in this study. The products used in the summarized literature consisted of a combination of many oils and carrier substances but, in this study, no other substance was used aside from the thyme extract (as revealed by its content analysis) thus, it is thought that the non-toxicity of the thyme extract may be due to the fact that no other active substance was used.

CONCLUSION

In this study, it was concluded that oral use of thyme extract once a day for 3 days is safe for dogs at the concentration and dose used in this study. Considering that the rate of zeroing egg excretion is high in dogs with lower egg numbers in our study, it is predicted that our study will make a significant contribution to research works to be done on thyme extract in different doses or application days in patients with high egg excretion.

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REFERENCES

- Abbas RZ, Iqbal Z, Khan A, *et al.* Options for integrated strategies for the control of avian coccidiosis. Int J Agric Biol. 2012; 14:1014-1020.
- Amin MM, El-Kabany H. Evaluation of protective and treatment of Thyme (Thymus vulgaris) oil on *Toxocara vitulorum* infected rats. J Rad Res Appl Sci. 2013; 6(1):209-232.
- Amin MM, Hafez EN, Mona A Abd Raboo. Assessment of radiation-attenuated vaccine or thyme oil treatment on controlling DNA damage and nitric oxide synthesis in brain of rat infected with *Toxocara canis*. Arab Journal of Nuclear Science and Applications. 2016; 49(2):199-210.
- Attia RAH, Mahmoud AE, Farrag HMM, Makboul R, Mohammed ME, Ibraheim Z. Effect of myrrh and thyme on Trichinella spiralis enteral and paranteral phases with inducible nitric oxide expression in mice. Mem Inst Oswaldo Cruz Rio de Janeiro. 2015; 110(8):1035-1041.
- Arczewska-Wlosek A, Swiatkiewicz S. The effect of a dietary herbal extract blend on the performance of broilers challenged with Eimeria oocysts. J Anim Feed Sci. 2012; 21:133-142.
- **Becskei C, Kryda K, Fias D**, *et al*. Field efcacy and safety of a novel oral chewable tablet containing sarolaner, moxidectin and pyrantel (Simparica Trio[™]) against naturally acquired gastrointestinal nematode infections in dogs presented as veterinary patients in Europe and the USA. Parasites & Vectors. 2020a; 13:70.
- **Becskei C, Kryda K, Thys M,** *et al.* Efcacy of a new oral chewable tablet containing sarolaner, moxidectin and pyrantel (Simparica Trio[™]) against induced ascarid infections in dogs. Parasites & Vectors. 2020b; 13:71.
- Behnia M, Haghighi A, Komeylizadeh H, Tabaei SJS, Abadi A. Inhibitory Effects of Iranian Thymus vulgaris Extracts on in Vitro Growth of *Entamoeba histolytica*. Korean J Parasitol. 2008; 46(3):153-156.
- Benli M, Yiğit N. Ülkemizde yaygın kullanımı olan kekik (Thymus vulgaris) bitkisinin antimikrobiyal aktivitesi. Orlab On-Line Mik Derg. 2005; 3(8):1-8.
- **Bischoff K, Guale F.** Australian tea tree (Melaleuca alternifolia) oil poisoning in three purebred cats. J Vet Diag Invest. 1998; 10:208-210.
- Bowman DD, Parsons JC, Grieve RB, Hepler DI. Effect of milbemycin on adult *Toxocara canis* infections in dogs with experimentally induced infection. Am J Vet Res. 1998; 49:1986-1989.
- Burrows CF, Batt RM, Sherding RG. Diseases of small intestine. In: Ettinger SJ, Feldman EC (eds) Textbook of Veterinary Internal Medicine. 4th ed. Saunders, Philadelphia; 1995. p.1169-1231.
- **Burt S.** Essential oils: their antibacterial properties and potential applications in foods a review. Int J Food Microbiol. 2004; 94(3):223-253.
- Cardenas RH, Nunaz CR, Contreras LM. Efficacy of two anthelmintic treatments, spinosad/milbemycin oxime and ivermectin/praziquantel in dogs with natural *Toxocara* spp. infection. Vet Parasitol. 2017; 247:77-79.
- Clark JN, Daurio CP, Barth DW, Batty AF. Evaluation of a beefbased chewable formulation of pyrantel pamoate against induced and natural infections of hookworms and ascarids in dogs. Vet Parasitol. 1991; 40(1-2):127-133.

- **Coles GC, Bauer C, Borgsteede FH M**, *et al.* World Association for the Advancement of Veterinary Parasitology (WAAVP) methods for the detection of anthelmintic resistance in nematodes of veterinary importance. Vet Parasitol. 1992; 44(1):35-44.
- Companion Animal Parasite Council. CAPC guideline on ascarids for dogs; 2020. Avaible at: https://capcvet.org/guidelines/ascarid/. Accessed July 22, 2020.
- **Despommier D.** Toxocariasis: Clinical aspects, epidemiology, medical ecology, and molecular aspects. Clin Microbiol Rev. 2003; 16(2):265-272.
- **Doğanay A, Öge H, Öge S, Yıldız K.** Nematoda (Yuvarlak Helmintler). In: Helmintoloji. Doğanay A (eds). Ankara, Türkiye: Ankara Nobel Tıp Kitabevleri Ltd. Şti.; 2018. P. 149-340.
- Fankhauser R, Hamel D, Dorr P *et al.* Efficacy of oral afoxolaner plus milbemycin oxime chewables against induced gastrointestinal nematode infections in dogs. Vet Parasitol. 2016; 225;117-122.
- Ferreira LE, Benincasa BL, Fachin AL, *et al.* Thymus vulgaris L. essential oil and its main component thymol: Anthelmintic effects against *Haemonchus contortus* from sheep. Vet Parasitol. 2016; 228:70-76.
- Gaur S, Kuhlenschmidt TB, Kuhlenschmidt MS, Andrade JE. Effect of oregano essential oil and carvacrol on *Cryptosporidium parvum* infectivity in HCT-8 cells. Parasitol Int. 2018; 67:170-175.
- Gargılı A, Tüzer E, Gülanber A, et al. Efficacy of moxidectin against *Toxocara canis* in experimentally infected dogs. Turk J Vet Anim Sci. 1999; 23:159-161.
- Genchi C, Traldi G, Manfredi MT. Field trials of the anthelmintic efficacy of nitroscanate and mebendazole in dogs. Vet Rec. 1990; 27:77-80.
- Genovese AG, McLean MK, Khan SA. Adverse reactions from essential oil-containing natural flea products exempted from environmental protection agency regulations in dogs and cats. J Vet Emerg Crit Care. 2012;22(4):470-475.
- Glickman LT, Schantz PM. Epidemiology and pathogenesis of zoonotic toxocariasis. Epidemiol Rev. 1981; 3(1):230-250.
- Hafez EN, Hafez MN, Amin MM. Effect of vaccination with irradiated *Toxocara canis* larvae or thyme oil treatment on testicular histochemical and immunohistochemical changes of rats. Trop Biomed. 2019; 36(2):430-442.
- Hassanain MA, Shaapan RM, Abou-El-Dobal SKA. Synergistic anthelmintic effect of Citrus aurantifolia swingle seeds and mebendazole in Egyptian dogs infected with Ancylostoma caninum and *Toxocara canis*: Trial to solve drug resistance problem. IJRSB. 2015; 3(9):104-111.
- Hoekstra R, Visser A, Wiley LJ, Weiss AS, Sangster NC, Roos MH. Characterization of an acetylcholine receptor gene of *Haemonchus contortus* in relation to levamisole resistance. Molec Biochem Parasitol. 1997; 84(2):179-187.
- Jackson F, Miller J. Alternative approaches to control Quo vadit? Vet Parasitol. 2006; 139:371-384.
- Jahangir M, Maqbool A, Tanveer A, Mahfooz A. Prevalence and therapy of ancyclostomiasis in dogs with Nigella sativa (Kalongi) and Saussurea lappa (Qust-e-Shireen). Pakistan Haryana Vet J. 2001; 40;48-51.

- Kara E, Yasa Duru S, Gökpınar S *et al.* Investigation of the prophylactic and therapeutic effectiveness of oral thyme extract in rats experimentally infected with *Cryptosporidium parvum*. Vet Res Commun. 2022; 3:1-11.
- Kotze AC, Hines BM, Ruffell AP. A reappraisal of the relative sensitivity of nematode pharyngeal and somatic musculature to macrocyclic lactone drugs. Int J Parasitol. Drugs Drug Resist. 2012; 2:29-35.
- Kozan E, Kırcalı Sevimli F, Birdane FM, Adanır R. Efficacy of eprinomectin against *Toxocara canis* in dogs. Parasitol Res. 2008; 102:397-400.
- Lloyd S, Toksikoz SR, Palmer EJL, Soulsby DHI. Zoonozis. Oxford: Oxford University Press; 1998. p. 841-854.
- Luis E, Ferreira BI, Benincasa AL *et al.* Thymus vulgaris L. essential oil and its main component thymol: Anthelmintic effects against *Haemonchus contortus* from sheep. Vet Parasitol. 2016; 228:70-76.
- Lynagh T, Cromer BA, Dufour V, Laube B. Comparative pharmacology of flatworm and roundworm glutamategated chloride channels: Implications for potential anthelmintics. Int. J Parasitol Drugs Drug Resist. 2014; 4:244-255.
- **Macpherson CN.** The epidemiology and public health importance of toxocariasis: a zoonosis of global importance. Int J Parasitol. 2013; 43(12-13):999-1008.
- Malatyalı E, Özçelik S, Gürsoy N. Kekik (Thymus vulgaris), kimyon (Cuminum cyminum) ve mersin (Myrtus communis) bitkilerinden elde edilen yağların *in-vitro* antileishmanial etkileri. Turk Hij Den Biyol Derg. 2009; 66(1):7-13.
- McTier TL, Siedek EM, Clemence RG *et al.* Efficacy of selamectin against experimentally induced and naturally acquired ascarid (*Toxocara canis* and *Toxascaris leonina*) infections in dogs. Vet Parasitol. 2000; 91:333-345.
- Morsy TA, Shoukry A, Mazyad SA, Makled KM. The effect of the volatile oils of Chenopodium ambrosioides and Thymus vulgaris against the larvae of *Lucilia sericata* (Meigen). J Egypt Soc Parasitol. 1998; 28(2):503-510.
- **Overgaauw PA, Van Knapen F.** Veterinary and public health aspects of *Toxocara* spp. Vet Parazitol. 2013; 193:398-403.

- Pal B, Mitra SK, Sacmal NK, Biswas D. Comparative efficacy of piperazine, ivermectin and albendazole against experimentally induced *Toxocara canis* infection in pups. Indian Vet J. 1995; 72:52-55.
- Payne-Johnson M, Maitland TP, Sherington J et al. Efficacy of selamectin administered topically to pregnant and lactating female dogs in the treatment and prevention of adult roundworm (*Toxocara canis*) infections and flea (*Ctenocephalides felis felis*) infestations in the dams and their pups. Vet Parasitol. 2000; 91:347-358.
- Pronk MEJ, Schefferlie GJ. Toxicological evaluation of certain veterinary drug residues in food, WHO Food Additives Series 41. WHO, Geneva. Available at: <u>http://www.inchem.org/documents/</u> jecfa/jecmono/v041je02.htm) Accessed Dec 12, 2022.
- **Remmal A, Achahbar S, Bouddine L, Chami N, Chami F.** In vitro destruction of Eimeria oocysts by essential oils. Vet Parasitol. 2011; 182:121-126.
- Sarışen Ö, Çalışkan D. Fitoterapi: Bitkilerle tedaviye dikkat (!). STED. 2005; 14(8):182-187.
- Silvestre A, Humbert JF. Diversity of benzimidazole-resistance alleles in populations of small ruminant parasites. Int J Parasitol. 2002; 32(7):921-928.
- Şenlik B. Teşhis yöntemleri. In: Helmintoloji. TINAR, R (eds). Ankara, Türkiye: Nobel Yayın Dağıtım ; 2006. p.463-535.
- Villar D, Knight MJ, Hansen SR, Buck WB. Toxicity of melaleuca oil and related essential oils applied topically on dogs and cats. Vet Hum Toxicol. 1994; 36:139-142.
- Woolf A. Essential oil poisoning. Clin Tox. 1999; 37(6):721-727.
- **Yıldız K, Başalan M, Duru Ö, Gökpınar S.** Antiparasitic efficiency of art emisia absinthium on *Toxocara cati* in naturally infected cats. Türkiye Parazitol Derg. 2011; 35:10-14.
- Young LM, Wiseman S, Crawley E, Bowman DD, Reinemeyer C, Sntder DE. Efectiveness of Credelio® Plus, a novel chewable tablet containing milbemycin oxime and lotilaner for the treatment of larval and immature adult stages of *Toxocara canis* in experimentally infected dogs. Parasites & Vectors. 2021; 14:256.